

MARKER ASSISTED SELECTION FOR SEED YIELD IN SOYBEAN [*GLYCINE MAX* (L.)  
MERR.] PLANT ROW YIELD TRIALS

BY

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DISSERTATION

Submitted in partial fulfillment of the requirements  
for the degree of Doctor of Philosophy in Crop Sciences  
in the Graduate College of the  
University of Illinois at Urbana-Champaign, 2010

Urbana, Illinois

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## ABSTRACT

Quantitative trait loci (QTL) controlling seed yield in soybean [*Glycine max* (L.) Merr.] have been difficult to confirm among populations. Our objective was to determine whether a method of marker-assisted selection (MAS) for seed yield in elite lines would be applicable to selection in soybean plant row yield trials (PRYTs). Lines from two populations with elite parents were grown in PRYTs in 2008 and tested with markers to identify quantitative trait loci (QTL) associated with seed yield. The first population was tested with 53 single nucleotide polymorphism (SNP) markers and the second population with 26 SNP markers. *F*-tests were conducted to determine which loci were significantly associated with seed yield in the PRYTs. Lines from each population were then selected from the PRYTs to form five groups from each population: high and low seed yield phenotypes, high and low seed yield genotypes, and random. The five groups from each population were planted at eight diverse locations in 2009. In one population, the mean of the genotypic high group was not statistically different than the phenotypic high group. In the other population, the mean of the genotypic high group was within 90 kg/ha<sup>-1</sup> of the mean of the phenotypic high group and was superior to the random group for seed yield. Even with the limited marker coverage, the genotypic selection method used in this study successfully identified lines in PRYTs that would not have been selected due to poor seed yield performance in 2008.

## **ACKNOWLEDGEMENTS**

I would like to thank my wife Shelli, I couldn't have done it without you, and my boys for keeping everything in perspective. In addition, I would like to thank Doug Appl, Jeff Baer, Ryan Boone, Tom Corbin, and Eric Roberts of the soybean research facility of Pioneer Hi-Bred located at Ivesdale, IL, for their help. I would also like to thank the staff of the soybean research projects of Pioneer Hi-Bred at Napoleon, OH, Dallas Center, IA, Hamel, IL, Lawrence, KA, and Princeton, IL. In addition, I would like to thank Scott Sebastian and the members of my committee for their guidance through this project.

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## INTRODUCTION

Soybean [*Glycine max* (L.) Merr.] is a very important oilseed crop (Wilcox, 2004) and in 2001 accounted for 35% of the total oilseed production of the world (Wilcox, 2004). The United States ranked number one in world soybean production in 2007 (FAO, 2010a). From 1961 to 2008, soybean seed yields in the United States increased from 1690 kg/ha<sup>-1</sup> to 2666 kg/ha<sup>-1</sup> (FAO, 2010b). While much of this increase is explained by genetic improvement in soybean varieties, some may also be explained by changes in cultural practices including the use of herbicides and fertilizer (Luedders, 1977). Luedders (1977) reported that one major improvement in soybean over time is a reduction of lodging. Reduced lodging results in more harvestable seed by improving ease of mechanical harvest.

Projections of human population growth indicate there may be in excess of 9 billion people by 2050 (U.S. Census Bureau, 2010). It has been suggested that an increasing world population, along with the increasing rate of population growth, requires a second green revolution (Pardue, 2010). At a 2008 conference of food security, United Nations secretary-general Ban Ki-moon indicated that food production must increase 50% by the year 2030 to meet the demand of a growing population (Ki-moon, 2008).

Marker-assisted selection (MAS) has been arguably the most important plant breeding tool developed in the last 20 years. Marker-assisted selection increases the heritability of selectable traits, and increases the rate of genetic gain, by decreasing the variance of the selected trait. The ability to select only lines with a desired trait phenotype promotes a more efficient use of plant breeding resources and is thus a desirable method of selection. Genetic markers have been developed for many commercially important traits and are being used in soybean breeding programs. Genetic markers useful for the selection of plants with resistance to phytophthora

(Hegstad et al., 1997, Salamath and Bhattacharyya, 1998), soybean cyst nematode (Concibido et al., 1996; Coryell et al., 1999), brown stem rot (Bachman et al., 2001), and more recently, soybean rust (Hyten et al., 2007), have been developed.

Plant row yield trials in a soybean breeding program are typically grown at a single location in a single replicate experiment in relatively short, single row plots (Hegstad et al., 1999; Streit et al., 2001; Cooper, 1990). This allows the breeder to observe lines from many populations and allow for the selection of lines from populations with an above average probability of being superior (Orf et al., 2004). Selections may be made visually, by seed yield data, or a combination of both. Selections that are made from the plant row yield trials are then evaluated at multiple locations and selection is imposed in subsequent generations until the line is either discarded or released as a breeding line or a commercial cultivar. A breeding program that combines both MAS to select plants for line derivation together with selection based on plant row yield trials can make more breeding progress than programs that do not use these methods.

Unfortunately, plant row yield trials have a low predictive value for seed yield. This is partially due to the lack of replications, which results in the confounding of location and genotype and increased errors in the seed yield estimates. Hegstad et al. (1999) reported the broad sense heritability of seed yield from plant row yield trials of five populations at 0.55, 0.49, 0.44, 0.63, and 0.29. They also reported correlation coefficients of seed yield from plant row yield trials with two subsequent years of yield testing from five populations, which averaged 0.17. Only one of the five correlation coefficients was significant at the 0.05 level. Streit et al. (2001) reported the broad sense heritability of four populations at 0.35, 0.61, 0.63, and 0.50. They reported the correlation coefficient between seed yield from plant row yield trials and one

subsequent year of yield testing was 0.41 averaged across four populations. All four coefficients were significant at the 0.05 probability level.

Traits such as seed yield that are difficult or expensive to phenotype and have low correlations between years are prime candidates for MAS. The use of molecular markers to select for seed yield could increase genetic gain through the elimination of lines lacking a beneficial allelic profile for seed yield. Unfortunately, the identification of molecular markers associated with major seed yield genes has been difficult in elite by elite populations (Reyna and Sneller, 2001; Sebastian et al., 2010).

A considerable amount of work has been done to identify quantitative trait loci (QTL) associated with seed yield. In the following discussions, when a marker is identified it is followed in parentheses by the chromosome number and position in Centimorgans (cM).

Orf et al. (1999a) conducted a study to identify seed yield QTL in soybean. They studied F<sub>7</sub> derived recombinant inbred lines (RILs) from the two populations 'Minsoy' x 'Archer' and 'Noir 1' x Archer. The lines were evaluated in three environments for several traits, including seed yield. The RILs were characterized for more than 400 molecular markers. Quantitative trait loci for seed yield were identified in association with Satt277 (6, 107.6 cM), B172\_2 (unknown), Satt561 (16, 71.4 cM), and Satt507 (1, 64.5 cM). The beneficial alleles associated with Satt277 and B172\_2 were derived from Noir 1 and those of Satt561 and Satt507 from Archer. The authors concluded that the beneficial alleles from Archer were likely already common in northern germplasm but not in southern germplasm.

Reyna and Sneller (2001) reported the results of introgressing three seed yield QTL identified in the cultivar Archer by Orf et al. (1999b) into southern germplasm. To test these QTL in southern germplasm, near isogenic line (NIL) populations were developed from the

crosses of Archer x 'Asgrow A5403' and Archer x 'Pioneer 9641'. The NIL populations were developed from F<sub>6</sub> plants that were heterozygous for the markers Satt144 (13, 102.1 cM), Satt002 (1, 64.5 cM), and Sct\_33/SoyHSP176 (unknown). F<sub>7:8</sub> lines homozygous for either the Archer or southern alleles were then selected to form the populations. The families were yield tested at two locations with two replications in 1998 and two locations with three replications in 1999. The seed yield data indicated the alleles from Archer did not contribute to seed yield in combination with southern germplasm. It was hypothesized that the Archer alleles were superior to the alleles of the other northern parent in the original study from Orf et al. (1999b), but may not be superior to the southern alleles in the southern environment. The authors commented that it may be difficult to successfully exploit beneficial alleles for complex traits in genetic backgrounds that are different than where they were originally mapped.

A study from Yuan et al. (2002) identified QTL for seed yield in a population of 100 RILs from the cross 'Essex' x 'Forrest' and a population of 94 RILs from the cross 'Flyer' x 'Hartwig'. The Essex x Forrest RILs were evaluated in two environments each in 1997 and 1998. The Flyer x Hartwig RILs were evaluated in two environments each in 1998 and 1999. A total of 135 polymorphic SSR markers were used to screen the Essex x Forrest RILs. Nine polymorphic simple sequence repeat (SSR) markers were used to screen the Flyer x Hartwig RILs. Markers Satt337 (9, 47.4 cM) (high yield allele from Essex), Satt167 (9, 45.7 cM), which is linked to Satt337 (high yield allele from Essex) and Satt294 (4, 78.7 cM) (high yield allele from Forrest) were found to be significantly associated with yield in the Essex x Forrest population. Two markers were significantly associated with seed yield in the Flyer x Hartwig RILs. These markers are Satt337 (9, 47.4 cM) (high yield allele from Flyer), and Satt539 (9, 2.8 cM) (high



yield allele from Hartwig). The seed yield QTL that mapped to Satt337 is common to both of the populations and is identical by descent in Flyer from Essex.

More recent studies of seed yield QTL have concentrated on the detection of positive QTL alleles from both *Glycine max* and *Glycine soja* plant introductions. Plant introductions often carry undesirable alleles that can be detrimental to breeding programs. However, exotic germplasm may contribute beneficial alleles that have been lost or were never present in the North American germplasm pool. An example of where exotic germplasm has played an important role in breeding programs has been in providing new disease resistance alleles, such as those that contribute to soybean cyst nematode resistance (Concibido et al., 1996).

Concibido et al. (2003) published a study involving the identification of a beneficial seed yield allele from *G. soja* PI 407305. Alleles beneficial to seed yield were identified by growing BC<sub>2</sub>F<sub>1</sub> lines in three locations in 1996 and seven locations in 1997. Lines carrying the allele from PI 407305 at AFLP locus U3944 showed an average 9.4% seed yield increase across years and locations. BC<sub>2</sub> lines of Asgrow germplasm were then developed in order to test the PI 407305 allele in other backgrounds. The following lines were used as recurrent parents: AG4501, AG2401, QP4459, QR4459, QR4544, and QR4604. Three of the populations showed a positive yield response: AG4501 ( $P < 0.0006$ ), AG2401 ( $P < 0.1057$ ), and QP4459 ( $P < 0.0119$ ). QR4459 ( $P < 0.7148$ ), QR4544 ( $P < 0.6541$ ) and QR4606 ( $P < 0.5210$ ) each showed a negative seed yield response.

A study published in 2004 by Kabelka et al. involved the identification of seed yield QTL in a population containing exotic alleles from PI 68508 and FC 04007B. 'BSR101' was used as the elite parent in the population. One hundred and sixty seven F<sub>5</sub>-derived lines were developed with the single-seed descent method. The lines were grouped into sets by maturity and evaluated

in six locations each in 2000 and 2001. A total of 145 polymorphic SSR markers were used to screen the lines for allelic profiles. A total of 15 seed yield QTL were reported. For nine of the 15 QTL, the yield increasing allele was from PI 68508 or FC 04007B. These QTL were mapped with Satt363 (6, 98.1 cM), Satt358 (10, 5.4 cM), Satt225 (5, 95.2 cM), Satt394 (18, 43.4 cM), Satt544 (9, 43.3 cM), Satt168 (14, 55.2 cM), Satt186 (17, 105.4 cM), Satt142 (12, 86.5 cM), and Satt308 (7, 130.8 cM). The seed yield increasing allele for the remaining QTL were from BSR101 and these QTL were mapped by the markers Satt383 (1, 56.6 cM), Satt191 (18, 96.6 cM), Satt387 (3, 53.3 cM), Satt339 (3, 75.9 cM), Satt547 (1, 67.8 cM), Satt440 (20, 112.7 cM). On average, the exotic alleles accounted for an average increase in seed yield of 2.9%, while the BSR101 alleles accounted for an average increase in seed yield of 3.3%.

In 2004, Wang et al. reported data from a seed yield QTL study in BC<sub>2</sub>F<sub>4</sub>-derived lines where 'IA2008' was used as the recurrent parent and *G. soja* 'PI 468916' was used as an exotic donor parent. To produce the lines for study, five random F<sub>2:3</sub> lines were crossed with IA2008 to produce BC<sub>1</sub>F<sub>1</sub> plants. These were again crossed with IA2008 to obtain BC<sub>2</sub>F<sub>1</sub> seed. Five BC<sub>2</sub>F<sub>1</sub> plants that traced to different F<sub>2</sub> plants were inbred to F<sub>4</sub> using the single-seed descent method. Each F<sub>4</sub> plant was then threshed to form BC<sub>2</sub>F<sub>4</sub> lines. The BC<sub>2</sub>F<sub>4</sub> lines and the two parents of the populations were grown in tests with two replicates in Lincoln, NE and Urbana, IL in both 1999 and 2000. Each line was screened for 302 polymorphic SSR markers and QTL were mapped by composite interval mapping (CIM). Four seed yield QTL regions were reported with the CIM method: Satt137 (9, 37.0 cM), Satt134 (6, 112.8 cM), Satt575 (15, 3.3), and Satt567 (7, 33.5 cM). Each of these QTL were derived from IA2008. The authors noted that the CIM method was unable to detect significant seed yield QTL from PI 468916. This may have been due to the

lack of QTL alleles from PI 468916 that could increase the yield of IA2008 or their inability to detect such positive QTL.

A study published by Smalley et al. (2004) took a slightly different approach to the detection of seed yield QTL. The authors identified seed yield QTL by studying allele frequency changes for markers in three recurrent selection populations. The populations were designed as follows: population AP10 originated from 40 PI parents, AP12 originated with 40 elite and 40 PI parents, and AP14 was formed with 40 elite parents. All parents were selected based upon their seed yield performance in maturity groups I to IV. Recurrent selection for seed yield was performed among  $F_4$  lines for four cycles. The cycle four populations and each set of founders were screened with 184 SSR markers that were polymorphic among the founders. Alleles with no selection pressure applied to them should have the same frequency in both the cycle four lines and the founders. If the allele frequency is different in the cycle four lines compared to the founder lines, then it is hypothesized that the alleles were selected because of their beneficial influence on seed yield over time. This approach was first described by Sebastian et al. (1995) as “Breeding Bias”. Smalley et al. (2004) assumed migration was not a concern due to the attention to detail when intermating lines for the next cycle. In addition, the self-pollinated nature of soybean helps reduce undesirable out-crossing. The effect of mutation was also considered negligible because there were only four cycles of recurrent selection. The authors did have concerns about genetic drift and to account for drift, they devised a simulation to create a probability density function (PDF). The simulation was repeated 10,000 times to provide an estimated PDF for allele frequencies of cycle four lines under the null hypothesis of no selection. A *P*-value of 0.05 was used as the significance level to declare that allele changes were not due to drift over time, but were due to selection. With this methodology, the authors identified a

number of seed yield QTL in their populations. Among the three populations, the authors identified a total of 60 SSR markers that significantly deviated from the expected ratio. Of the deviated markers, 15 were unique to the PI parents, 9 were unique to the elite parents, and 36 were common between both the PI and elite parents. This list of significantly deviated markers is too extensive to fully give here. However, nine of the deviated markers map to regions where yield QTL were previously mapped. The large number of identified QTL in this study is likely due to the large number of parents used to form the populations. The large number of parents would have contributed more alleles that are segregating and can be detected.

In 2007, Guzman et al. reported mapping exotic seed yield QTL alleles in three backcross populations. In the development of the three backcross populations, lines were developed and tested for seed yield and those lines with the greatest seed yield were crossed back to the recurrent parent. BC<sub>3</sub>F<sub>2</sub>-derived lines were developed for study from a population developed using PI 68658 as a donor parent and 'Lawrence' as a recurrent parent. In addition, BC<sub>2</sub>F<sub>5</sub>-derived lines were also evaluated in the study from a population developed using PI 407720 as a donor parent and 'Beeson 80' as a recurrent parent. A third population used a selected line from the cross PI 391583 and PI 297544 as the donor parent and 'Kenwood' as the recurrent parent. Lines in the yield tests were BC<sub>1</sub>F<sub>5</sub> derived. The Beeson 80 and Kenwood populations were evaluated at three locations in 2003 and four locations in 2004. The Lawrence population was evaluated at four locations in 2003 and three locations in 2004. Lines from each population were evaluated with polymorphic SSR markers: 45 in the Beeson 80 population, 84 in the Kenwood population, and 30 in the Lawrence population. A total of 13 QTL significant for seed yield were detected: three in the Beeson 80 population and five each in the other two. Quantitative trait loci for seed yield were mapped with the markers Satt046 (9, 45.6cM), Satt215 (16, 44.8 cM), and

Satt547 (16, 67.7 cM) in the Beeson 80 population. Quantitative trait loci for seed yield were mapped with Satt399 (4, 76.2 cM), Satt557 (6, 112.2 cM), Satt405 (16, 11.7 cM), Satt313 (19, 34.5 cM), and Satt477 (10, 82.1 cM) in the Kenwood population. In the Lawrence population, QTL were mapped with Satt300 (5, 30.9 cM), Satt474 (14, 75.3 cM), Satt640 (6, 30.5 cM), Satt622 (16, 42.4 cM), and Satt445 (10, 20.4 cM). The alleles beneficial to seed yield were derived from the PI parent in eight of the 13 QTL identified. The authors noted that all of these QTL mapped to regions that seed yield QTL were previously identified.

Most of the published yield QTL mapping studies are from populations segregating for exotic alleles. This is in contrast to initial studies focused on identifying the QTL in elite populations. One major drawback of mapping QTL in elite populations is the limited allelic diversity available in the modern elite germplasm pool (Concibido et al., 2003; Carter et al., 2004; Kabelka et al., 2004; Smalley et al., 2004; Guzman et al., 2007) resulting in many regions of the genome being fixed among elite parents. This reduces the number of polymorphisms which, in turn, reduces the ability to detect allelic effects. Studies were more successful in identifying seed yield QTL in populations segregating for exotic alleles. With the exception of studies published by Reyna and Sneller (2001) and Concibido et al., (2003), little work has been published on confirming exotic alleles beneficial for seed yield among populations.

The primary source of germplasm in modern breeding programs of North America are elite, commercial cultivars (Kabelka et al., 2004). Limited progress has been made in improving elite populations through the use of mapped QTL controlling seed yield (Reyna and Sneller, 2001; Sebastian et al., 2010). Previous work in elite populations showed that seed yield QTL can be detected, but it was difficult to confirm these QTL in other populations (Reyna and Sneller, 2001). With this in mind, Sebastian et al. (2010) describes a method for implementing the use of

seed yield QTL within elite populations. The authors used a method known as Context-Specific Marker Assisted Selection, which is designed to detect QTL alleles beneficial to seed yield within a set of highly inbred  $F_{7:8}$  lines that were derived from elite cultivars. The  $F_{7:8}$  lines are derived from an elite cultivar and grown in plant row yield trials to collect seed yield data. The  $F_{7:8}$  lines were screened with SSR markers at loci that were polymorphic in the parents of the original population. *F*-tests were then performed to determine which loci were significant for seed yield. The allele with the greatest seed yield mean at each significant locus was considered to be beneficial to seed yield. Lines with the most alleles beneficial to seed yield were then selected to resynthesize a new version of the original elite cultivar with a greater seed yield. In their study, the seed yields of the reselected lines were greater than the original five elite cultivars by an average of 3.1%. While all the resynthesized cultivars had a higher seed yield mean than the original cultivar, only three of the five differences were significant at the 0.05 probability level.

The purpose of our study was to determine whether the methodology explained by Sebastian et al. (2010) could be used in addition to phenotypic selection for seed yield in plant row yield trials of lines developed from biparental crosses. While Sebastian et al. (2010) made selections within highly inbred cultivars that were segregating for only a small proportion of their genomes, in this study the yield QTL will be mapped and selections will be made among  $F_4$ -derived lines in populations developed from biparental crosses.

Plant row yield trials were grown in 2008. Selections were made from these and grown in an eight location confirmation study in 2009. The data from the confirmation study indicated that we were able to use genotypic selection to mitigate Type II errors in the plant row yield trial selections.

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# **MARKER ASSISTED SELECTION FOR SEED YIELD IN SOYBEAN [*Glycine max* (L.)**

## **Merr.] PLANT ROW YIELD TRIALS**

### **INTRODUCTION**

Plant row yield trials (PRYTs) are commonly used by soybean breeders to select lines superior for seed yield early in the product development cycle (Hegstad et al., 1999; Streit et al., 2001). Plant row yield trials are typically grown as single replicate experiments with a relatively small plot size (Hegstad et al., 1999; Streit et al., 2001; Orf et al., 2004). The lack of replicates and small plot size is due to the limited number of seed that can be obtained from a soybean plant.

Unfortunately, the lack of multiple replicates not only results in large errors in the estimates of seed yield but also the confounding of genotype and environment. This can limit the predictability of seed yield data obtained from the experiment and thus reduce the effectiveness of selection from such experiments. This may result in the genotypes with the best potential seed yield not being selected due to poor performance in the PRYT.

### **Usefulness of Quantitative Trait Loci for Predicting Seed Yield**

Several studies identifying quantitative trait loci (QTL) controlling seed yield in soybean have been published. Many involve the identification of seed yield QTL from plant introductions (Concibido et al., 2003; Kabelka et al., 2004; Smalley et al., 2004; Wang et al., 2004; Guzman et al., 2007). Concibido et al., (2003) reported identifying and introgressing an exotic allele into six elite cultivars, three of which resulted in an increase in seed yield ( $P < 0.0006$ ,  $P < 0.1057$ , and  $P < 0.0119$ ). A third resulted in an increase in seed yield, but was not statistically significant. Guzman, et al., (2007) reported QTLs from exotic germplasm in

previously identified regions. This indicates exotic germplasm may contribute alleles beneficial to seed yield in some elite backgrounds.

Unfortunately, confirmation of seed yield QTL among elite populations has been difficult (Reyna, et al., 2001; Sebastian, et al., 2010). The apparent lack of major QTLs for seed yield among elite soybean populations may be due to the limited number of founders of modern elite germplasm. It has been reported that 26 ancestors account for nearly 90% of the alleles found in the modern soybean germplasm pool (Carter, et al., 2004). This may have resulted in the fixation of any major seed yield QTL during early cycles of plant breeding (Bernardo, 2008).

### **Context Specific Marker Assisted Selection**

A method for detecting and leveraging seed yield QTL that are context specific in elite populations was introduced by Sebastian, et al. (2010). With context specific QTL, it is expected that the QTL will be detected in some populations and not others. One possible benefit of this method may be the ability to increase the effectiveness of selection in PRYTs by including beneficial allelic combinations, in addition to seed yield phenotype, as selection criteria.

Sebastian et al. (2010) describes a method to reselect lines with an allelic combination beneficial for seed yield from highly inbred F<sub>7</sub> lines. Their study involved deriving sub-lines from nine elite cultivars and determining seed yield of the sub-lines in PRYTs. Simple sequence repeat markers were analyzed to determine alleles beneficial to seed yield. Sub-lines were then selected that had an allelic profile that was predicted to have a greater seed yield than the mother line. Seed yield gains up to 5.8% were achieved with this procedure.

### **Study Objective**

A marker assisted selection (MAS) program for seed yield would be of great benefit to plant breeders. If realized, it would increase the efficiency of breeding programs by allowing the breeder to select lines with the greatest genetic potential for seed yield. This would increase the heritability of the trait by decreasing the number of lines with limited genetic potential in a breeding program.

The purpose of my study was to determine whether the methodology described by Sebastian et al. (2010) can be used to increase the effectiveness of selection in PRYTs by allowing us to select lines superior in seed yield based upon predicted beneficial allelic combinations in addition to seed yield phenotype.

## **MATERIALS AND METHODS**

### **Population Development**

Two single cross populations were developed in 2006 by Pioneer Hi-Bred International, Inc. (Pioneer) by crossing pairs of elite Pioneer cultivars possessing desirable agronomic characteristics. Population 34 was developed at Dallas Center, IA and population 35 was developed at Ivesdale, IL. All parents were of Maturity Group III.

The F<sub>1</sub> seeds of the single crosses were planted in November 2006 at the Pioneer nursery in Salinas, Puerto Rico. The F<sub>1</sub> plants were confirmed as hybrids for Population 34 by flower color and those of Population 35 by genetic marker analysis. The F<sub>2</sub> seeds of each population were advanced using the bulk method and planted in February 2007 at the Pioneer nursery in Salinas. The F<sub>3</sub> seed of each population was advanced using the bulk method and planted in May 2007 at the Pioneer Research Center in Ivesdale, IL. Each plant was harvested individually but not threshed. This resulted in harvesting 531 F<sub>3</sub> plants in Population 34 and 654 F<sub>3</sub> plants in Population 35. This was not considered to be an adequate number of plants to detect seed yield QTL with small effects in PRYTs. To increase the number of plants for PRYTs, one three seeded pod was collected from each plant and used to form a resynthesized bulk of each population. The F<sub>4</sub> seeds were planted in December 2007 at the Pioneer nursery in Pergamino, Argentina. Leaf tissue was collected from each F<sub>4</sub> plant for QTL marker analysis. The F<sub>4</sub> plants of each population were harvested and threshed individually.

### **Plant Row Yield Trials**

In the spring of 2008, the F<sub>4.5</sub> lines with sufficient seed from each population were divided and planted in PRYTs at one of three diverse locations for seed yield and maturity

classification. Each population was grown in a separate experiment and the lines within each population were divided into experiments of 63 entries for Population 34 and 69 entries for Population 35. Each experiment was planted as an augmented design. The two parents of each population were replicated three times within each experiment to form an error estimate for each experiment. The seed from each plant in Argentina was returned to Ivesdale, IL where they were packaged for planting in the PRYT experiments. Each population was divided into three equal sets for the three locations to be planted. As the experiments were packaged, lines with insufficient seed were not included for planting. This resulted in differing numbers of lines for each location. For Population 34, 486 lines were planted at Hedrick, IA, 384 lines were planted at Ivesdale, IL, and 502 lines were planted at Napoleon, OH. For Population 35, 552 lines were planted at Hedrick, IA, 506 lines were planted at Ivesdale, IL, and 552 lines were planted at Napoleon, OH. Hedrick, IA was planted on May 19, Ivesdale, IL was planted on June 14, and Napoleon, OH was planted on May 22. Each plot was a single row 1.5 m long with a row spacing of 0.9 m. The seeding rate was 30 seeds  $m^{-1}$  of row.

During the growing season, the Napoleon, OH location experienced flooding. As a result, the lines planted there were harvested but the data were not included in any seed yield analysis. Maturity was recorded as the date when 95% of the pods in each plot reached mature color (R8) (Fehr and Caviness, 1977). Lines were harvested individually with a self propelled combine. Seed yield data were collected and expressed on a 13% moisture basis. Least squares (LS) means for seed yield were calculated using the MIXED procedure of the SAS statistical software package (release 9.2) (SAS Institute, 2008). Line was considered to be a fixed effect. Days from planting to maturity was used a covariate in the analysis.

## Single Nucleotide Polymorphism Marker Analysis

During the summer of 2008, marker analysis was performed at the Pioneer Research Center at Dallas Center, IA on the F<sub>4</sub> leaf tissue samples obtained from each plant of both populations grown in Argentina. This analysis was done to identify markers associated with seed yield. Markers used in this analysis were selected by first testing the parents of the populations with 982 Pioneer single nucleotide polymorphism (SNP) markers. All markers identified as polymorphic in this screen were then used to test the lines in the populations. This resulted in the selection of 53 polymorphic markers for Population 34 and 26 polymorphic markers for Population 35. Assuming that each marker is able to detect a QTL within 15 cM, it is estimated that these markers provided 45% detection coverage of the genome for Population 34 and 21% detection coverage in Population 35. The marker analyses were completed using a 5 $\mu$ L reaction format TaqMan SNP marker detection assay (Applied Biosystems, 2004).

Significance of each locus for seed yield was determined with *F*-tests using the MIXED procedure of the SAS statistical software package (release 9.2) (SAS Institute, 2008). The analysis was performed as described by Sebastian et al. (2010) except that days to maturity was used a covariate in the analysis. Associations between markers and seed yield were declared significant using a *P*-value of 0.25. This threshold was chosen because previous reports suggested that a relatively low stringency should be used in identifying loci significant for seed yield (Bernardo, 2008). Allelic effects were determined for each significant locus with the LSMEANS option of the MIXED procedure of the SAS statistical software package (release 9.2) (SAS Institute, 2008). For significant loci, the allele in the homozygous state with the greater mean was considered to be the allele most beneficial to seed yield. The allele in the homozygous state with the lesser mean was considered to be the allele that was either neutral or deleterious to



seed yield. Lines homozygous for the beneficial allele at all or most of the significant loci are considered to have the most desirable genotype for seed yield.

In order to identify whether our QTL were in regions where yield QTL were previously identified, we estimated the positions of our QTL on the soybean map from Choi et al (2007). This was done with markers that have known positions on both the Pioneer and consensus maps. The positions of the Pioneer QTL on the soybean consensus map were then estimated by using the FORECAST function in Microsoft Office Excel 2007 (Microsoft Corporation, 2006).

### **Selection of Lines for Further Study**

Five groups of 11 lines each were formed from each population for further study. The five groups formed were phenotypic high, phenotypic low, genotypic high, genotypic low, and random. The phenotypic high group is composed of lines that had the greatest seed yield in the PRYT. The phenotypic low group is composed of lines with the least seed yield in the PRYT. The genotypic high group is composed of lines that were homozygous for the beneficial allele at all or most of the significant loci identified. The genotypic low group is composed of lines that were homozygous for the neutral/deleterious allele at all or most of the significant loci identified. Lines heterozygous at any significant locus were not considered for selection in either the genotypic high or low group. The random group represents lines that were selected by assigning random numbers to each line and selecting the 11 lines with the lowest numbers. Lines were allowed to reside in multiple groups. For example, a line may exist in both the phenotypic high and genotypic high groups. Although the PRYT seed yield data from Napoleon, OH were not used in the phenotypic selections, lines harvested from this environment were considered for inclusion in the genotypic high, genotypic low, and random groups. In Population 34, five of the

11 lines in the genotypic high group possessed the beneficial allele for all nine markers significant for yield. The remaining lines possessed favorable alleles for eight of the nine markers. Five of the 11 lines in the genotypic low group lacked the beneficial allele for at all nine markers. The remaining lines lacked the beneficial allele for eight of the nine markers. In Population 35, all lines of the genotypic high group possessed the beneficial allele for all significant markers and all lines of the genotypic low group lacked the beneficial allele for all markers. After forming the five groups for Population 35, five lines existed in two groups. As a result, two additional genotypic high lines were selected to fill the extra space in the experiment. This resulted in 13 lines in the genotypic high group.

The selected  $F_{4:6}$  lines were sent to the Pioneer nursery in Viluco, Chile for seed increase. This was done to ensure an adequate seed supply for wide area testing in 2009 and to obtain seed from the same source environment for each line. Each line was harvested individually and returned to Ivesdale, IL where it was packaged for testing in 2009.

### **Germination Testing of the Selected Genotypes**

A warm germination test was performed on each of the selected  $F_{4:7}$  lines by the Pioneer germination laboratory in Johnston, IA. The testing was done by growing each selected line in a randomized complete block in four replications of 50 seeds each as described by Meis et al. (2003) except that the 2008 editions of Rules for Testing Seeds (AOSA, 2008) and 2006 edition of the Seedling Evaluation Handbook (AOSA, 2006) were used as references. Germination percentage was determined by counting the number of seeds that germinated, dividing by the 50 seeds tested, and multiplying by 100.

Data were analyzed using the MIXED procedure of the SAS statistical software package (release 9.2) (SAS Institute, 2008). Groups and lines nested in groups were considered fixed effects. Replication was considered a random effect. *F*-tests were used to evaluate the significance of fixed effects. The LSMEANS option of the MIXED procedure of the SAS statistical software package (release 9.2) (SAS Institute, 2008) was used to calculate the mean for each group. Single degree of freedom linear contrasts were used to determine significant differences between the genotypic high group and the other groups for each trait.

### **Field Testing of Selected Genotypes**

The selected  $F_{4:7}$  lines from each population were grown in separate experiments at eight diverse locations in 2009. Each experiment was a randomized complete block design in a 5 x 12 row-column configuration. The row-column configurations were obtained from the OPTEX procedure of the SAS statistical software package (release 9.2) (SAS Institute, 2008). Elite checks were used to bring the number of entries in each experiment to 60. Each location was grown as a single replication. Population 34 was grown at locations near Metamora, IL; Gilman, IL; Pesotum, IL; Anna, OH; Napoleon, OH; Atlantic, IA; Abington, IA; and Washington, IA. Metamora, IL, was planted on May 30; Gilman, IL was planted on May 27; Pesotum, IL, was planted on June 1; Anna, OH was planted on May 22; Napoleon, OH, was planted on May 25; Atlantic, IA, was planted on May 20; Abington, IA, was planted on June 1; and Washington, IA was planted on May 23. Population 35 was grown at the same locations as Population 34 except that Emery, IL, which was planted on June 7, replaced Gilman, IL. The plots were two rows 3.7 m in length spaced 0.76 m apart within the plot and between adjacent plots. The seeding rate was 30 seeds  $m^{-1}$  of row.

Maturity was recorded as the date when 95% of the pods in each plot reached mature color (R8) (Fehr and Caviness, 1977). Plant height was measured at maturity in centimeters from the soil surface to the terminal node. Lodging was scored at maturity using a scale of 1 (all plants erect) to 5 (all plants prostrate). The plots were harvested with a two-row self-propelled combine. The seed yield was expressed in  $\text{kg ha}^{-1}$  on a 13% moisture basis. Seed size was recorded from the harvested seed and expressed as weight in grams of 100 seeds.

Data were analyzed using the MIXED procedure of the SAS statistical software package (release 9.2) (SAS Institute, 2008). Groups and lines nested in groups were considered fixed effects. Environments, rows nested in environments, columns nested in environments, and the interaction between environment and group were considered random effects. *F*-tests were used to evaluate the significance of fixed effects. The LSMEANS option of the MIXED procedure of the SAS statistical software package (release 9.2) (SAS Institute, 2008) was used to calculate the mean of each trait for each group. Single degree of freedom linear contrasts were used to determine significant differences between the genotypic high group and the other groups for each trait. Phenotypic correlations among the traits measured in the study were computed based on the line means averaged across environments with the CORR procedure of the SAS statistical software package (release 9.2) (SAS Institute, 2008).

## RESULTS AND DISCUSSION

The QTL analysis of the PRYTs resulted in nine SNP markers identified as significant ( $P < 0.25$ ) in Population 34 and three SNP markers in Population 35 (Tables 1 and 2). In Population 34, two significant markers on chromosome 17 (77.2 cM and 77.7 cM) and two on chromosome 18 (102.1 cM and 103.5 cM) are closely linked and likely map the same QTL. In addition, the two significant markers on chromosome 16 are within 21.6 cM of each other and may also map the same QTL. Therefore, we likely mapped six or seven unique QTL in this population. For Population 35, the location of the two significant SNP markers on chromosome 16 are in close proximity (42.8 cM and 49.4 cM). Therefore, we likely mapped two unique QTL controlling seed yield in this population. The frequency distributions of seed yield for lines homozygous for either the paternal or maternal parent allele for the significant QTL show bimodal distributions in Population 34 (Figures 1-9). The paternal alleles for the significant QTL in Population 35 show bimodal distributions (Figures 10-13) but only QTL S14 shows a bimodal distribution of the maternal allele (Figure 10). These bimodal peaks are due to the two environments in the study. None of the QTL were significant for a QTL by location interaction at the 0.05 significance level.

QTL significant for seed yield were identified in each population in regions that had been previously associated with seed yield. In Population 34, S39 and S40 on chromosome 16 map within 8.3 cM and 6.9 cM, respectively, of the marker G815\_1, which Specht et al. (2001) used to map a seed yield QTL. S39 maps to within 2.6 cM of Satt406 that Sebastian et al. (2010) used to map a seed yield QTL. In addition, a seed yield QTL was identified by Cregan et al. (1999) with Satt191 on chromosome 18 and this marker maps to within 9.8 cM of marker S47 and 10.6 cM of marker S46, which we used to map a seed yield QTL in Population 34. A seed yield QTL

was mapped in Population 35 with the marker S14 and this maps 9.4 cM from Satt339, which was used to map a seed yield QTL by Kabelka et al. (2004).

There were significant ( $P<0.01$ ) seed yield differences among the groups of selected lines in both populations based on the 2009 seed yield tests (Table 3). In both populations, the phenotypic high group had the greatest average seed yield followed by the genotypic high group. The difference between the phenotypic high and genotypic high groups was significant ( $P<0.05$ ) for Population 35 but not for Population 34 (Table 4). In Population 35, the phenotypic high group mean was  $90 \text{ kg/ha}^{-1}$  greater in seed yield than the mean of the genotypic high group (Table 3). The genotypic high group in Population 34 was not significantly different from the random group which is at least partially the result of the line with the highest seed yield in the confirmation study of Population 34 being a random selection (Tables 4 and 5). The genotypic high group in Population 35 had significantly ( $P<0.01$ ) greater seed yield than the random group (Table 4). The line with the greatest seed yield in the 2009 test of Population 35 was a line in the genotypic high group (Table 5). This indicates that selection based on genotype alone was successful in identifying lines with superior seed yield in this population.

Single degree of freedom contrasts showed the genotypic high group in both populations was significantly ( $P<0.01$ ) greater in seed yield than both the phenotypic low and genotypic low groups (Table 4). There was a difference of  $242 \text{ kg/ha}^{-1}$  in Population 34 and  $163 \text{ kg/ha}^{-1}$  in Population 35 between the genotypic high and genotypic low groups (Table 3). This indicates we were able to induce a seed yield shift with very few QTL in each population. Seed yield was significantly ( $P<0.01$ ) correlated with seed size and height in both populations and with maturity, emergence and lodging in Population 35 (Tables 6 and 7). Correlation of seed yield between 2008 and 2009 for all lines was significant ( $P<0.05$ ) only in population 35, which had a

coefficient of 0.28, indicating the PRYT seed yield data of 2008 had a low predictive value of seed yield in 2009.

There were significant ( $P<0.01$ ) differences among the groups for maturity in both populations in the 2009 tests (Table 3). Population 34 shows the largest differences with the genotypic high group being the latest in maturity at 2 days later than the random group (Table 3). However, the correlation between maturity and seed yield in Population 34 was not significant (Table 6). The maturity differences among the groups in Population 35 are less pronounced with the phenotypic high group having the latest maturity and the genotypic high group having a maturity equal to the random group (Table 3). Analysis of data from the PRYTs indicated that none of the markers associated with seed yield were significant for maturity in 2008. Maturity was significantly ( $P<0.01$ ) correlated with seed size, height, and lodging in both populations and with seed yield and emergence in Population 35 (Tables 6 and 7).

There were significant ( $P<0.01$ ) differences among the groups of Population 34 for lodging (Table 3). Although statistically significant, the differences in lodging among the groups have little practical value. There was no significant difference among groups in Population 35 for lodging (Table 3). Lodging was significantly ( $P<0.01$ ) correlated with maturity and height in both populations and with seed yield and seed size in Population 35 (Tables 6 and 7).

There were significant ( $P<0.01$ ) differences among the groups of both populations for height (Table 3). The mean heights of the phenotypic low and genotypic low groups were shorter than the other three groups in both populations (Table 3). The Genotypic High group was significantly ( $P<0.01$ ) greater in height than the Random group with a 6 cm difference in Population 34 (Tables 3 and 4). The Genotypic High group was 1 cm greater in height than the Phenotypic High group in Population 35, but this difference was not significant at the 0.05 level

(Tables 3 and 4). The genotypic high group in Population 35 was not significantly different in height when compared to the Phenotypic High and Random groups (Table 4). Height was significantly ( $P<0.01$ ) correlated with seed yield, maturity, and lodging in both populations and with germination in Population 35 (Tables 6 and 7). Height was negatively and significantly ( $P<0.01$ ) correlated with seed size both populations (Tables 6 and 7).

There were significant ( $P<0.05$  and  $P<0.01$ , respectively) differences among the groups of both populations for germination (Table 3). Even though there are significant differences among the groups, there is no clear pattern between the populations. There also are no clear patterns between the populations for correlations of germination with the other measured traits (Tables 6 and 7). There are significant differences ( $P<0.01$ ) among the groups in Population 34 for seed size but not for Population 35 (Table 5). The means of the groups are very similar indicating little meaningful difference among the groups. Seed size was significantly ( $P<0.01$ ) correlated with seed yield, and height in both populations and with lodging in Population 35 (Tables 6 and 7). Seed size was negatively and significantly ( $P<0.01$ ) correlated with maturity in both populations (Tables 6 and 7).

The fact that the phenotypic high group had a greater mean seed yield in both populations indicates that we were unable to account for all the components of seed yield in these populations with the QTL identified. This is likely either the result of our inability to detect all loci controlling seed yield or a QTL by environment effect where the identified QTL were not as important to seed yield in the 2009 environments as in 2008. Detecting seed yield QTL is especially difficult with non-replicated PRYT tests. More QTL controlling seed yield could have potentially been detected with a lower stringency level in our QTL analysis of seed yield data from PRYTs. Bernardo (2008) recommended using alpha levels from 0.20 to 0.40 when



selecting for QTL involved in complex traits because highly stringent significance levels may lead to an unrealistic expectation of QTL effects. An alpha level of 0.25 was used in this study. Using an alpha level of 0.40 would have added seven more loci in Population 34 and an additional three loci in Population 35. At this low stringency level, some of these additional QTL will likely be false positives. However, according to Bernardo (2008), the benefits of identifying additional QTL with minor effects is greater than the drawbacks associated with a higher incidence of false positives. Identifying plants that carry the positive alleles for all of these QTL will also be difficult. In Population 34, if the six unique QTL originally detected were added to the seven additional QTL, only one out of every 46,482 plants in a single seed descent F<sub>4</sub> population would carry all 13 positive alleles in a beneficial homozygous state. A marker based recurrent selection program would be needed to combine all of these positive alleles into a single plant. As with any population, the breeder must choose between testing more plants of fewer populations or fewer plants of more populations.

Despite extremely limited genomic coverage, the methods implemented in this study resulted in selection of superior lines based upon genotype alone for seed yield. In both populations, the genotypic high group had a greater seed yield mean than the random group. In Population 34, the genotypic high group was not statistically different from the phenotypic high group at the 0.05 significance level. In Population 35, the genotypic high group was within 90 kg/ha<sup>-1</sup> of the phenotypic high group. Two of the top ten lines for seed yield in 2009 were selected based on genotype alone, including the line with the greatest seed yield in Population 35 (Table 5). It is unlikely that these lines would have been selected because of their poor seed yield phenotype in PRYTs. Knowledge of beneficial allelic combinations gives us a greater level

of information for selecting lines that may be superior. Using the methodology in this study, we increased our selection effectiveness from PRYTs by including genotype as a selection criteria.

## TABLES AND FIGURES

Table 1. Position and seed yield means of selected loci from Population 34 grown in plant row yield trials in 2008.

Marker	Chromosome	cM	Locus <i>P</i> -value	Beneficial Allele Mean (kg ha <sup>-1</sup> )	Neutral/Deleterious Allele Mean (kg ha <sup>-1</sup> )
S1	Unmapped†		0.1540	3265	3239
S39	16	77.4	0.0545	3326	3171
S40	16	99.0	0.2252	3272	3198
S41	17	16.2	0.2220	3265	3192
S42	17	77.2	0.0019	3279	3178
S43	17	77.7	0.0004	3286	3171
S46	18	102.1	0.0851	3292	3165
S47	18	103.5	0.0955	3292	3178
S48	19	13.7	0.1622	3272	3192

†Chromosome and position based on Pioneer soybean map.

Table 2. Position and seed yield means of selected loci from Population 35 grown in plan row yield trials in 2008.

Marker	Chromosome	cM	Locus <i>P</i> -value	Beneficial Allele Mean (kg ha <sup>-1</sup> )	Neutral/Deleterious Allele Mean (kg ha <sup>-1</sup> )
S14	3†	92.8	0.2412	3328	3295
S69	16	42.9	0.0115	3343	3281
S70	16	49.5	0.0092	3336	3271

†Chromosome and position based on Pioneer soybean map.

Table 3. Agronomic performance of the five groups studied from two populations grown at eight diverse locations in 2009.

Trait	Group	Population			
		34		35	
		Mean	Range	Mean	Range
Seed Yield (kg ha <sup>-1</sup> )	genotypic high	4001**	3801 - 4181	4150**	4046 – 4388
	phenotypic high	4029	3895 - 4257	4240	4132 – 4363
	genotypic low	3759	3236 - 4052**	3987	3878 – 4228*
	phenotypic low	3326	3034 - 3559**	3887	3423 – 4302**
	random	3944	3218 - 4285**	4013	3635 – 4246**
Maturity (d) †	genotypic high	130**	129 - 133**	126**	123 - 129**
	phenotypic high	129	125 - 132**	127	123- 131**
	genotypic low	127	125 - 128	125	124 - 128
	phenotypic low	127	123 - 130**	124	121 - 129**
	random	128	122 - 133**	126	122 - 129**
Lodging (score) ‡	genotypic high	2.0**	1.0 - 2.8**	1.5	1.1 – 1.9**
	phenotypic high	2.0	1.8 - 2.4	1.7	1.3 – 2.0
	genotypic low	1.6	1.3 – 2.0**	1.5	1.2 – 1.8**
	phenotypic low	1.5	1.3 - 2.1**	1.5	1.3 – 1.8*
	random	2.0	1.3 - 2.4**	1.6	1.3 – 1.8

Table 3. Continued.

Trait	Group	Population			
		34		35	
		Mean	Range	Mean	Range
Height (cm)	genotypic high	103**	98 – 108**	91**	87 – 98**
	phenotypic high	102	96 – 107**	92	88 – 95**
	genotypic low	96	88 – 103**	90	84 – 98**
	phenotypic low	93	87 – 102**	87	83 – 94**
	random	97	81 – 106**	92	84 – 100**
Germination (%)	genotypic high	88*	77 – 91**	83**	77 – 88
	phenotypic high	87	75 – 91**	84	80 – 88
	genotypic low	86	81 – 93**	83	74 – 89**
	phenotypic low	87	82 – 92*	79	71 – 89**
	random	89	82 – 95**	81	69 – 86**
Seed Size (g per 100)	genotypic high	17.3**	16.2 – 18.3**	18.1	16.8 – 19.2**
	phenotypic high	17.9	16.2 – 18.5**	18.3	17.1 – 19.6**
	genotypic low	17.8	16.2 – 18.9**	18.2	17.3 – 19.0**
	phenotypic low	17.0	16.0 – 18.0**	18.0	16.2 – 19.3**
	random	18.2	16.7 – 20.4**	18.1	16.9 – 19.1**

\*,\*\* Differences among the means of the groups or means of the lines within each group were significant at the 0.05 and 0.01 probability levels, respectively.

† Days after planting.

‡ Lodging score = 1.0 (all plants erect) to 5.0 (all plants prostrate).

Table 4. Single degree of freedom linear contrasts of genotypic high group versus other groups.

Trait	Population							
	34				35			
	PH†	GL	PL	RA	PH	GL	PL	RA
Seed Yield	ns‡	**	**	ns	*	**	**	**
Maturity	**	**	**	**	**	*	**	ns
Lodging	ns	**	**	ns	-§	-	-	-
Height	ns	**	**	**	ns	**	**	ns
Germination	ns	*	ns	ns	ns	ns	**	ns
Seed Size	**	*	ns	**	-	-	-	-

\*,\*\* Differences among the means of the groups or means of the lines within each group were significant at the 0.05 and 0.01 probability levels, respectively.

† PH=phenotypic high, GL=genotypic low, PL=phenotypic low, RA=random.

‡ Differences were not significant at the 0.05 probability level.

§ A dash sign indicates the contrast was not performed due to lack of significance among the groups for the trait.

Table 5. Seed yield of selected lines grown in plant row yield trials (PRYT<sub>s</sub>) in 2008 and eight diverse environments in 2009.

Population	2009 Seed Yield Rank	Line	Group <sup>†</sup>	2008 PRYT <sub>s</sub> (kg ha <sup>-1</sup> ) <sup>‡</sup>	2009 Yield Test (kg ha <sup>-1</sup> ) <sup>§</sup>
34					
	1	34-1140	RA	1775	4285 <sup>¶</sup>
	2	34-1157	PH	4340	4257
	3	34-1730	GH	1211	4181
	4	34-0692	GH	2005	4181
	5	34-0679	RA	1444	4131
	6	34-0724	GH/PH	3701	4129
	7	34-0754	PH	3792	4124
	8	34-1340	PH	4698	4097
	9	34-1118	PH	3305	4073
	10	34-0467	RA	3045	4055
	11	34-0873	GL	2889	4052
	12	34-1029	RA	2562	4052
	13	34-1847	RA	709	4045
	14	34-1058	GH	2115	4038
	15	34-1716	GL	1340	4019
	16	34-0947	PH	3483	4018
	17	34-1757	GH	3059	4008
	18	34-1779	RA	2729	4008
	19	34-1046	GH	3147	3999
	20	34-1288	GL	2472	3979
	21	34-0662	PH	4538	3967
	22	34-1055	GH	2255	3960
	23	34-1333	RA	1171	3957
	24	34-1242	GL	3071	3953
	25	34-0626	PH	3789	3949
	26	34-1806	GL	2622	3947
	27	34-1627	GH	3457	3944
	28	34-1032	RA	1011	3926
	29	34-0518	PH	4543	3911
	30	34-1440	PH	4324	3902
	31	34-0761	RA	1779	3899

Table 5. Continued.

Population	2009 Seed Yield Rank	Line	Group†	2008 PRYTs (kg ha <sup>-1</sup> )‡	2009 Yield Test (kg ha <sup>-1</sup> )§
	32	34-0506	PH	4569	3895
	33	34-1553	GH	3652	3895
	34	34-0777	GH	1158	3879
	35	34-1625	RA	3551	3810
	36	34-0414	GH	1773	3801
	37	34-1897	GL	1118	3775
	38	34-0450	GL	2614	3758
	39	34-0485	GL	3886	3702
	40	34-0503	GL	2991	3626
	41	34-1426	PL	2817	3559
	42	34-0778	PL	2192	3472
	43	34-1256	PL	1932	3471
	44	34-0378	PL	2400	3466
	45	34-1486	PL	2339	3425
	46	34-0351	GL	1512	3307
	47	34-1654	PL	2665	3298
	48	34-0538	PL	2406	3268
	49	34-1808	PL	1619	3252
	50	34-0760	GL	1067	3236
	51	34-1666	RA	3400	3218
	52	34-0405	PL	1896	3198
	53	34-1609	PL	2507	3138
	54	34-0297	PL	2742	3034
35	1	35-2455	GH	1658	4388#
	2	35-3018	PH	5726	4371
	3	35-2701	PH	5692	4363
	4	35-2684	PH	6079	4345
	5	35-2972	GH	3999	4312
	6	35-3545	PH	3716	4311
	7	35-2328	PL	1443	4302
	8	35-2323	PH	3639	4287



Table 5. Continued.

Population	2009 Seed Yield Rank	Line	Group <sup>†</sup>	2008 PRYTs (kg ha <sup>-1</sup> ) <sup>‡</sup>	2009 Yield Test (kg ha <sup>-1</sup> ) <sup>§</sup>
	9	35-2589	RA	1370	4246
	10	35-2808	RA	3461	4238
	11	35-3388	GL	3873	4228
	12	35-2366	PL	1117	4219
	13	35-2235	PH	4647	4218
	14	35-2485	GL	3091	4196
	15	35-2690	PH	4255	4184
	16	35-2343	GH/RA	1491	4180
	17	35-2152	GH	2989	4179
	18	35-3049	GH	3763	4162
	19	35-3927	PH	4615	4146
	20	35-3544	PH	3420	4141
	21	35-3993	PH	2721	4136
	22	35-3707	RA	2890	4134
	23	35-2592	GH/PH	3662	4132
	24	35-2213	PL	2460	4127
	25	35-3787	GH	2882	4125
	26	35-2344	PL	1508	4122
	27	35-2820	GH	3775	4110
	28	35-3190	GH	2815	4100
	29	35-2039	GH	2439	4096
	30	35-2646	GH	2269	4063
	31	35-3071	GH/RA	2781	4053
	32	35-2671	GH	2835	4046
	33	35-3980	RA	2001	4046
	34	35-1983	GL	2339	4038
	35	35-3731	GL	3733	3996
	36	35-2450	GL	1786	3995
	37	35-3536	RA	1734	3977
	38	35-3497	RA	2127	3964
	39	35-2184	GL	1232	3941
	40	35-3478	GL	3055	3932
	41	35-3801	GL	2992	3889

Table 5. Continued.

Population	2009 Seed Yield Rank	Line	Group†	2008 PRYTs (kg ha <sup>-1</sup> )‡	2009 Yield Test (kg ha <sup>-1</sup> )§
	42	35-2062	GL	1913	3883
	43	35-2244	GL	2134	3881
	44	35-3253	GL	2353	3878
	45	35-2474	RA	3015	3861
	46	35-2874	PL	3281	3851
	47	35-2951	PL	3209	3831
	48	35-3726	PL	2815	3814
	49	35-2728	RA	4145	3812
	50	35-3757	PL	2307	3736
	51	35-2270	PL	2902	3695
	52	35-2845	PL/RA	3142	3635
	53	35-2666	PL	2003	3423

†GH=genotypic high, PH=phenotypic high, GL=genotypic low, PL=phenotypic low, RA=random.

‡LS Mean calculated on single replication PRYT.

§LS Mean calculated using eight replications at diverse locations.

¶ LSD ( $\alpha=0.05$ ) Of Population 34 in 2009 = 232.0 kg/ha<sup>-1</sup>

#LSD ( $\alpha=0.05$ ) Of Population 35 in 2009 = 197.2 kg/ha<sup>-1</sup>

Table 6. Phenotypic correlation coefficients of Population 34 grown at eight diverse locations in 2009.

Trait	Maturity	Seed Size	Emergence	Height	Lodging (score)†
Seed Yield (kg ha <sup>-1</sup> )	0.06	0.47**	-0.02	0.16**	0.10
Maturity (d)‡		-0.28**	-0.05	0.35**	0.40**
Seed Size (g per 100)			0.01	-0.41**	0.00
Germination (%)				0.02	0.02
Height (cm)					0.67**

\*,\*\* Significant at the 0.05 and 0.01 probability levels, respectively.

† Lodging score = 1.0 (all plants erect) to 5.0 (all plants prostrate).

‡ Days after planting.

Table 7. Phenotypic correlation coefficients of Population 35 grown at eight diverse locations in 2009.

Trait	Maturity	Seed Size	Emergence	Height	Lodging (score)†
Seed Yield (kg ha <sup>-1</sup> )	0.32**	0.38**	0.19**	0.34**	0.26**
Maturity (d)‡		-0.21**	0.14*	0.38**	0.46**
Seed Size (g per 100)			-0.09	-0.40**	0.16*
Germination (%)				0.17**	-0.09
Height (cm)					0.38**

\*,\*\* Significant at the 0.05 and 0.01 probability levels, respectively.

† Lodging score = 1.0 (all plants erect) to 5.0 (all plants prostrate).

‡ Days after planting.

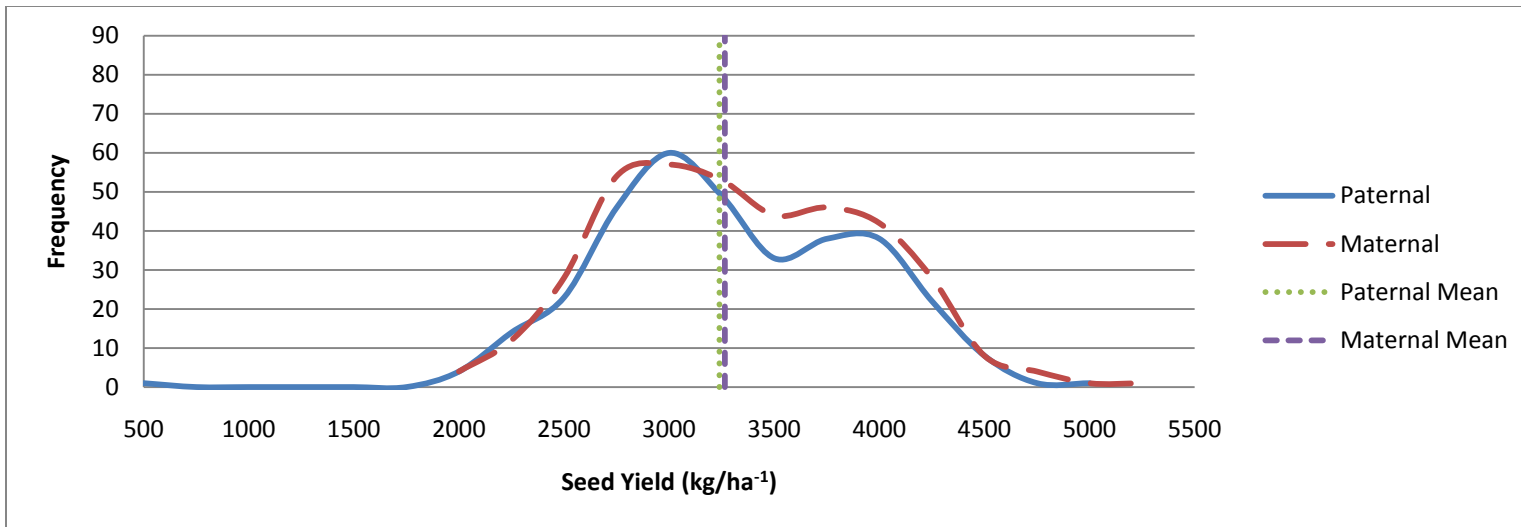


Figure 1. Frequency distribution of seed yields for lines homozygous for either the maternal or paternal allele at the quantitative trait locus S1 in Population 34 at two plant row yield trial locations in 2008.

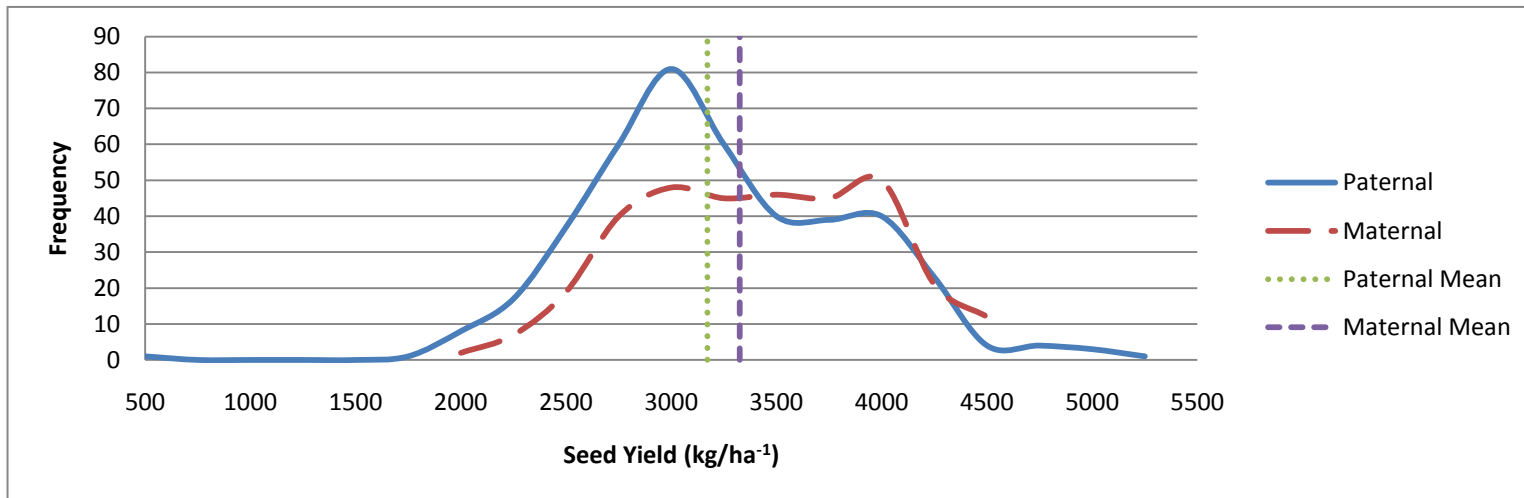


Figure 2. Frequency distribution of seed yields for lines homozygous for either the maternal or paternal allele at the quantitative trait locus S39 in Population 34 at two plant row yield trial locations in 2008.

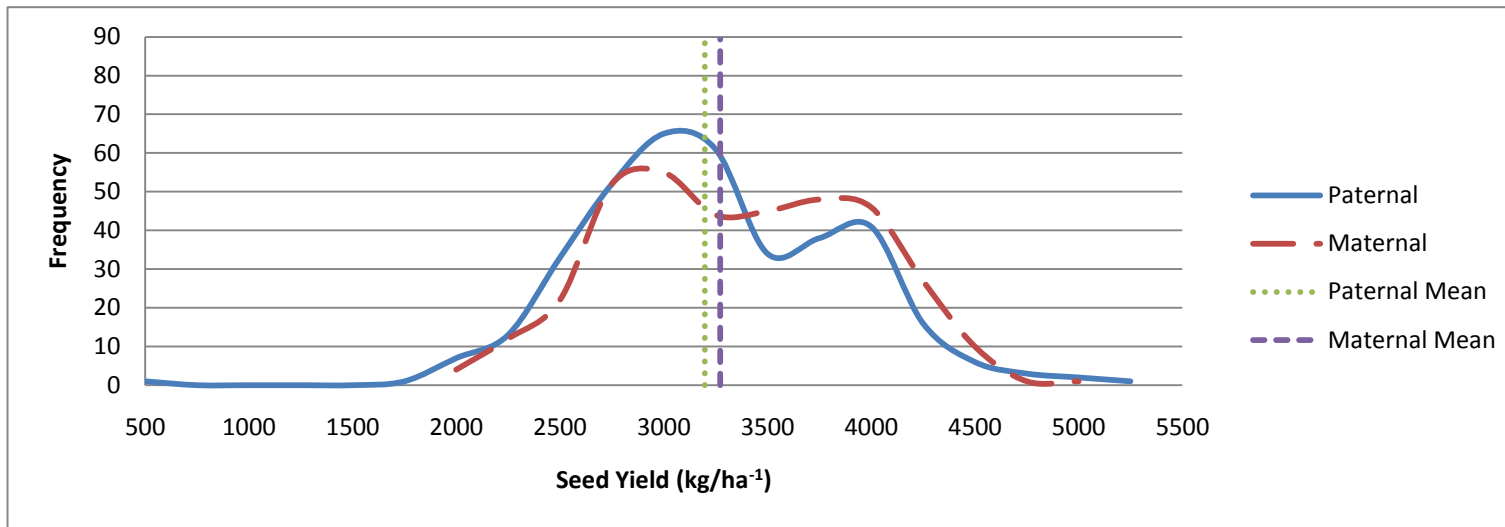


Figure 3. Frequency distribution of seed yields for lines homozygous for either the maternal or paternal allele at the quantitative trait locus S40 in Population 34 at two plant row yield trial locations in 2008.

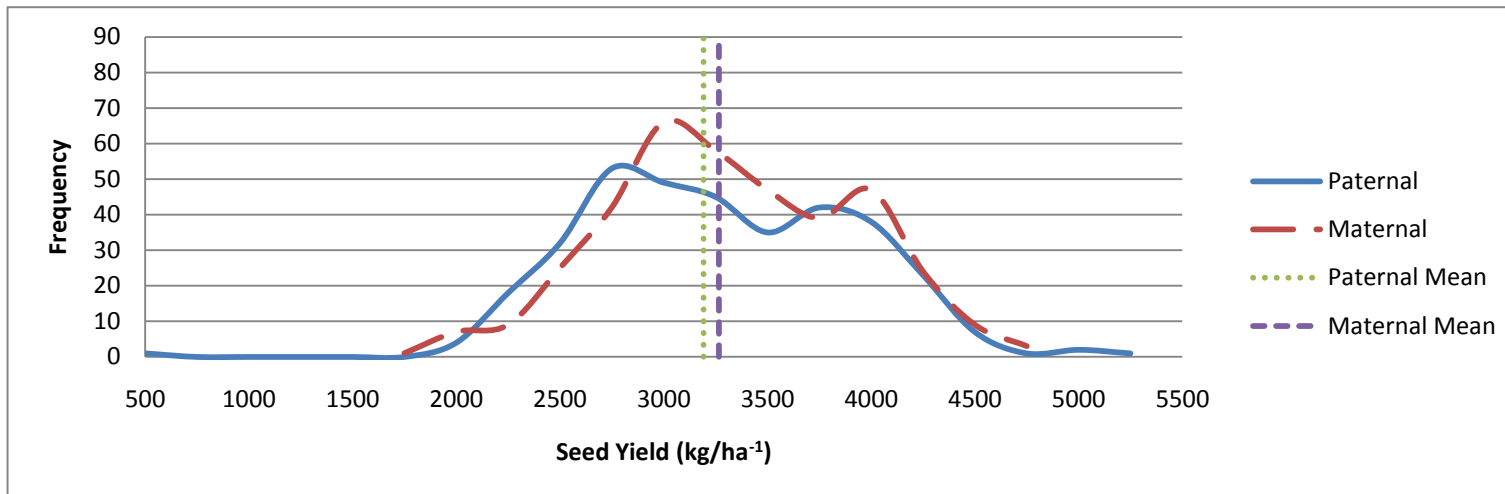


Figure 4. Frequency distribution of seed yields for lines homozygous for either the maternal or paternal allele at the quantitative trait locus S41 in Population 34 at two plant row yield trial locations in 2008.

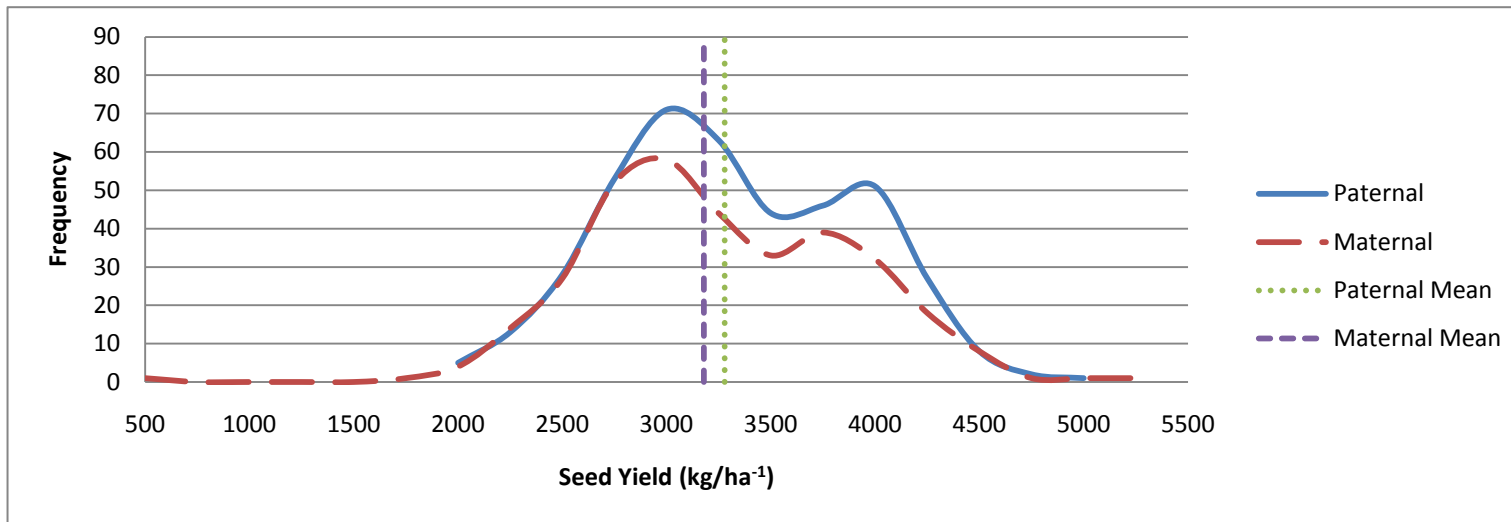


Figure 5. Frequency distribution of seed yields for lines homozygous for either the maternal or paternal allele at the quantitative trait locus S42 in Population 34 at two plant row yield trial locations in 2008.

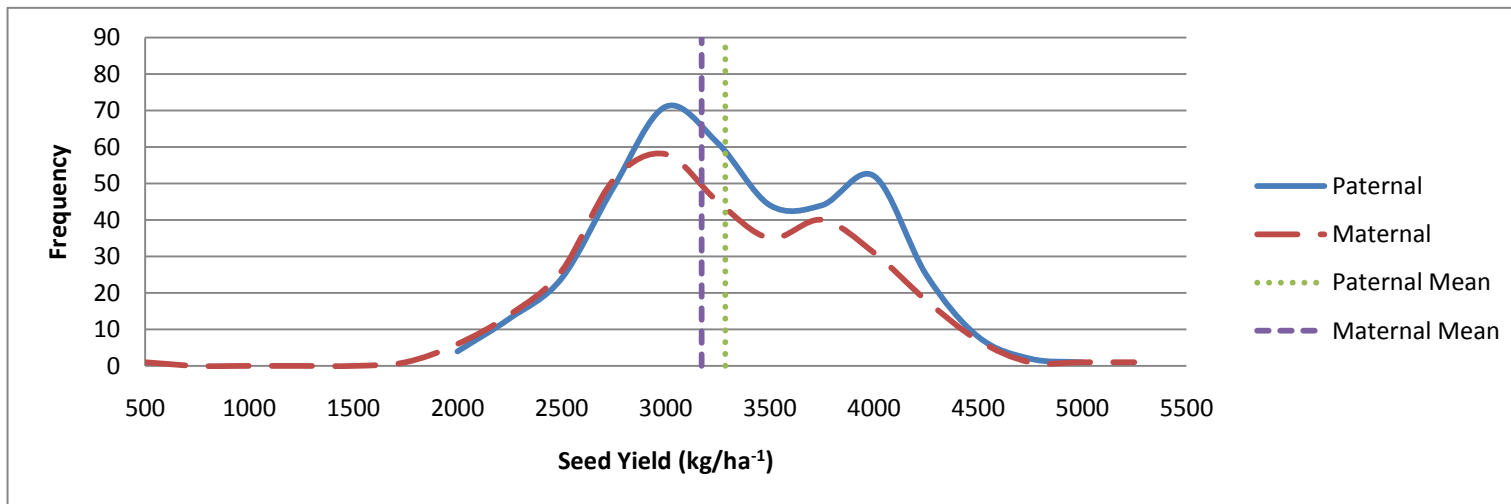


Figure 6. Frequency distribution of seed yields for lines homozygous for either the maternal or paternal allele at the quantitative trait locus S43 in Population 34 at two plant row yield trial locations in 2008.

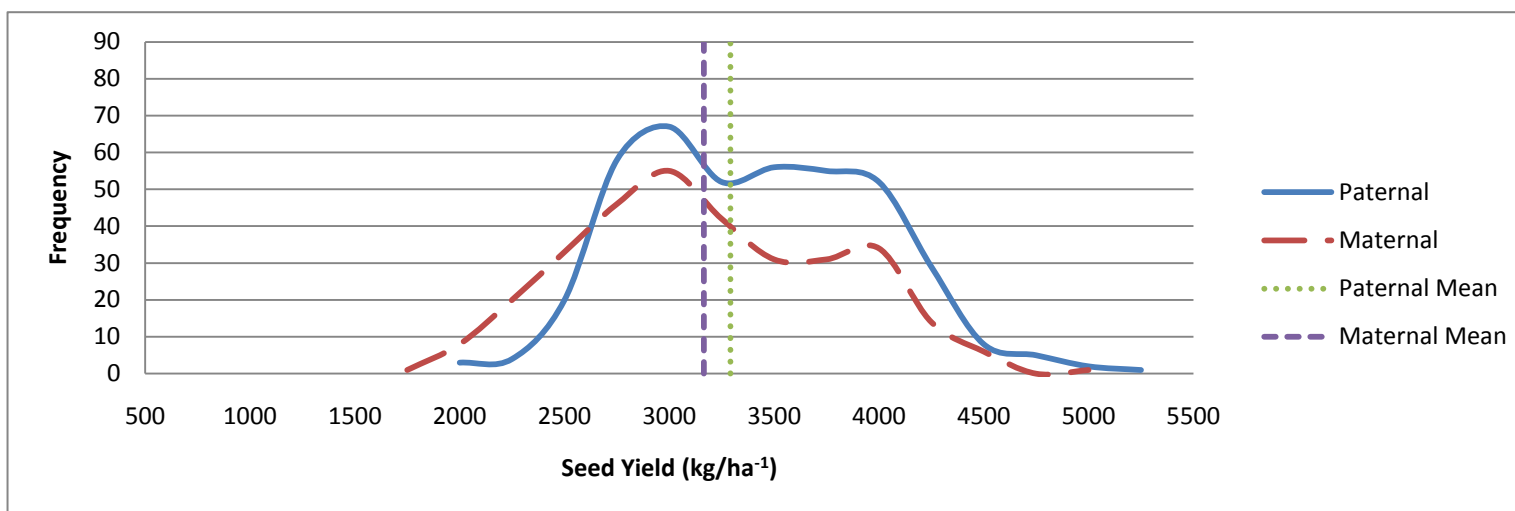


Figure 7. Frequency distribution of seed yields for lines homozygous for either the maternal or paternal allele at the quantitative trait locus S46 in Population 34 at two plant row yield trial locations in 2008.

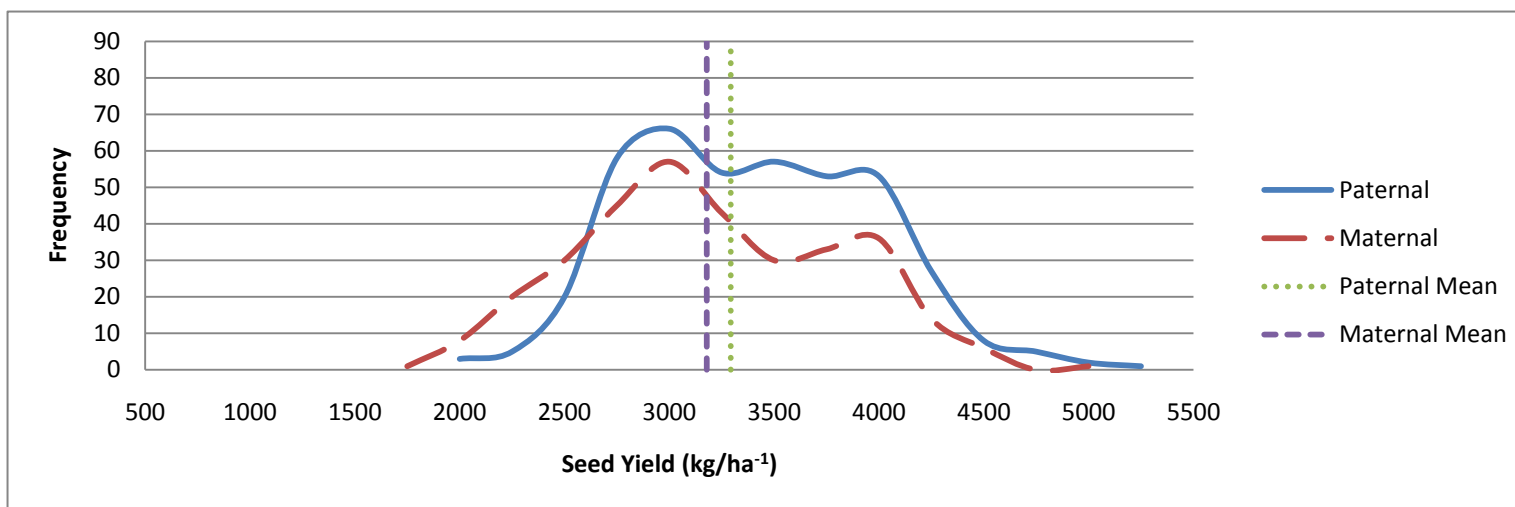


Figure 8. Frequency distribution of seed yields for lines homozygous for either the maternal or paternal allele at the quantitative trait locus S47 in Population 34 at two plant row yield trial locations in 2008.

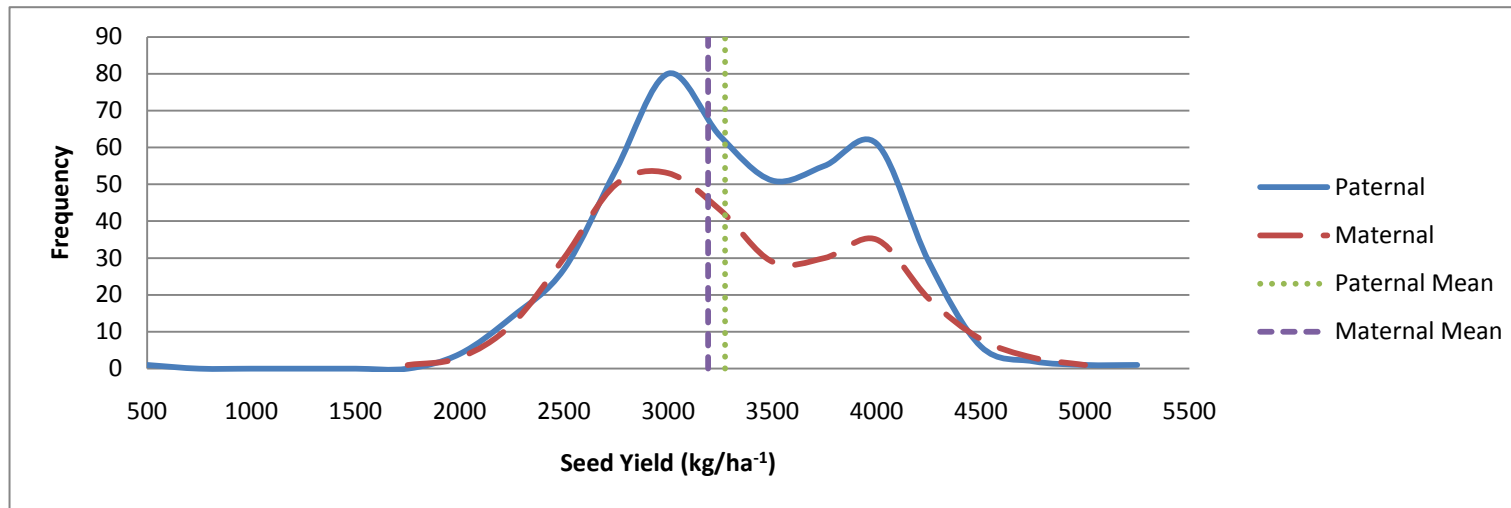


Figure 9. Frequency distribution of seed yields for lines homozygous for either the maternal or paternal allele at the quantitative trait locus S48 in Population 34 at two plant row yield trial locations in 2008.



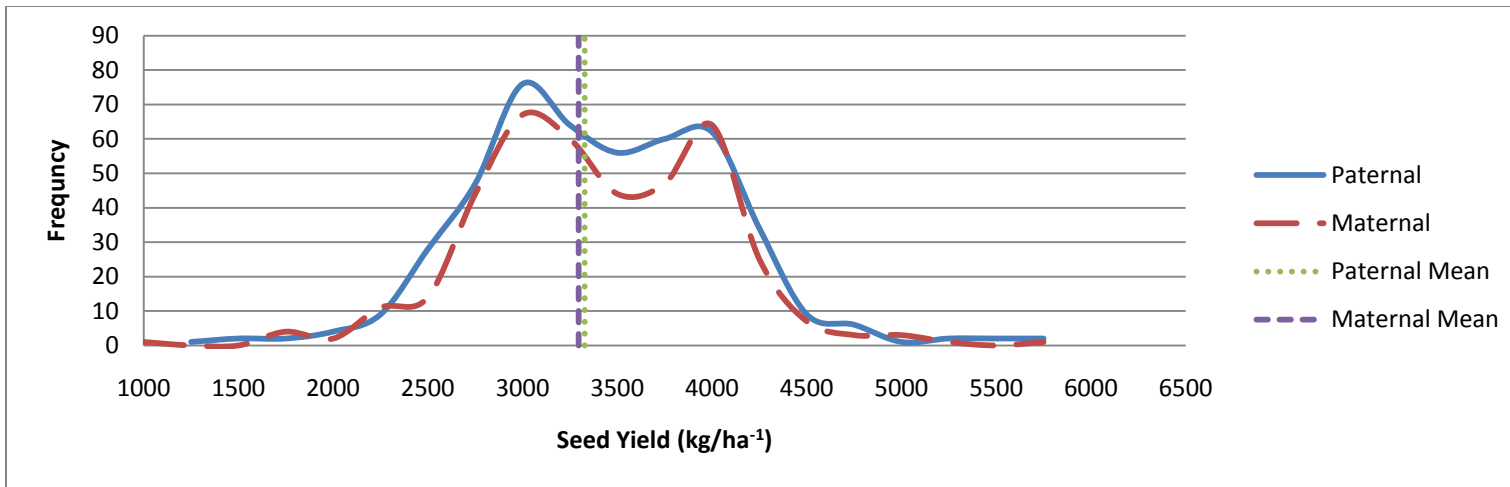


Figure 10. Frequency distribution of seed yields for lines homozygous for either the maternal or paternal allele at the quantitative trait locus S14 in Population 35 at two plant row yield trial locations in 2008.

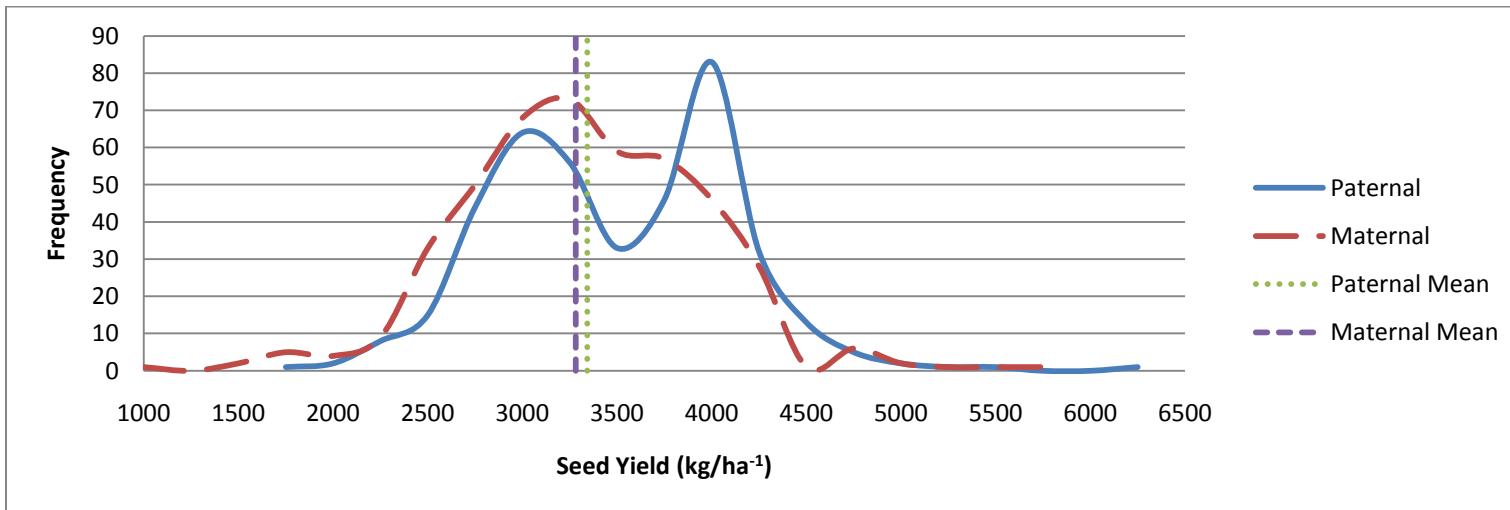


Figure 11. Frequency distribution of seed yields for lines homozygous for either the maternal or paternal allele at the quantitative trait locus S69 in Population 35 at two plant row yield trial locations in 2008.

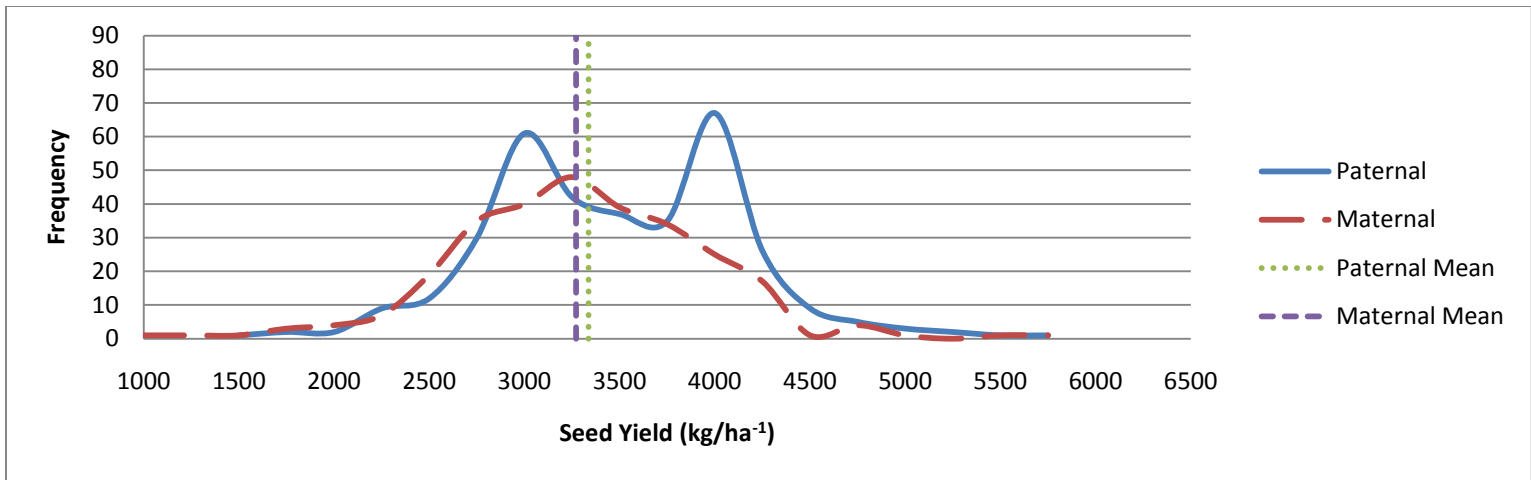


Figure 12. Frequency distribution of seed yields for lines homozygous for either the maternal or paternal allele at the quantitative trait locus S70 in Population 35 at two plant row yield trial locations in 2008.

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## **APPENDIX A**

### **ANALYSIS OF VARIANCE FOR AGRONOMIC TRAITS AMONG ENVIRONMENTS**

The additive model for all traits for the combined analysis across environments in 2009

was:

$$Y_{ijklm} = \mu + E_i + R_{(i)j} + C_{(i)k} + G_l + L_{(l)m} + EG_{im} + \beta(M_{ijklm} - M_{\dots}) + e_{(ijklm)}$$

where:

$Y_{ijklm}$  = observed value of the  $m^{\text{th}}$  line in the  $l^{\text{th}}$  group in the  $k^{\text{th}}$  column of the  $j^{\text{th}}$  row of the  $i^{\text{th}}$  environment

$\mu$  = overall mean

$E_i$  = effect of the  $i^{\text{th}}$  environment,  $NID(0, \sigma^2_i)$

$R_{(i)j}$  = effect of the  $j^{\text{th}}$  row within the  $i^{\text{th}}$  environment,  $NID(0, \sigma^2_j)$

$C_{(i)k}$  = effect of the  $k^{\text{th}}$  column within the  $i^{\text{th}}$  environment,  $NID(0, \sigma^2_k)$

$G_l$  = effect of the  $l^{\text{th}}$  group

$L_{(l)m}$  = effect of the  $m^{\text{th}}$  line within the  $l^{\text{th}}$  group

$EG_{im}$  = effect of the interaction between the  $i^{\text{th}}$  environment and the  $m^{\text{th}}$  group,  
 $NID(0, \sigma^2_{im})$

$\beta(M_{ijklm} - M_{\dots})$  = maturity covariate

$e_{(ijklm)}$  = residual of the  $m^{\text{th}}$  line in the  $l^{\text{th}}$  group in the  $k^{\text{th}}$  column of the  $j^{\text{th}}$  row of the  $i^{\text{th}}$  environment

Table A1. Analysis of variance and expected mean squares for combined environments for all traits

Source of Variation	Expected Mean Squares
Environment	$\sigma_e^2 + rgl\sigma_{C(E)}^2 + cgl\sigma_{R(E)}^2 + rcgl\sigma_E^2$
Row/Environment	$\sigma_e^2 + cgl\sigma_{R(E)}^2$
Column/Environment	$\sigma_e^2 + rgl\sigma_{C(E)}^2$
Group	$\sigma_e^2 + rcl\sigma_{EG}^2 + ercl\Phi_G$
Line/Group	$\sigma_e^2 + erc\Phi_{L(G)}$
Genotypic High/Group	$\sigma_e^2 + ercGI\text{PhPIRa}\Phi_{Gh(G)}$
Genotypic Low/Group	$\sigma_e^2 + ercGh\text{PhPIRa}\Phi_{Gl(G)}$
Phenotypic High/Group	$\sigma_e^2 + ercGhGI\text{PIRa}\Phi_{Ph(G)}$
Phenotypic Low/Group	$\sigma_e^2 + ercGhGI\text{PIRa}\Phi_{Pl(G)}$
Random/Group	$\sigma_e^2 + ercGhGI\text{PhPI}\Phi_{Ra(G)}$
Environment x Group	$\sigma_e^2 + rcl\sigma_{EG}^2$
Maturity Covariate	$\sigma_e^2 + \Phi_{MAT}$
Error	$\sigma_e^2$



Table A2. Analysis of variance for yield across eight diverse environments in 2009 for Population 34 and 35.

Source of Variation	Population			
	34		35	
	df	Mean Square	df	Mean Square
Environment	7	1271.22**	7	1023.81**
Row/Environment	32	35.44**	32	20.90**
Column/Environment	88	38.18**	88	29.31**
Group	4	745.16**	4	150.48**
Line/Group	50	59.51**	52	20.29**
Genotypic High/Group	10	13.37	12	8.19
Genotypic Low/Group	10	111.54**	10	18.78*
Phenotypic High/Group	10	15.83	10	12.20
Phenotypic Low/Group	10	33.88**	10	34.79**
Random/Group	10	110.25**	10	44.43**
Environment x Group	28	17.67	28	14.07
Maturity Covariate	1	12.45	1	10.56
Error	224	12.27	242	9.22
C.V.		6.2		5.0

\*, \*\* Significant at the 0.05 and 0.01 probability levels, respectively

Table A3. Analysis of variance for lodging across seven diverse environments in 2009 for Population 34 and 35.

Source of Variation	Population			
	34		35	
	df	Mean Square	df	Mean Square
Environment	6	72.30**	6	23.16**
Row/Environment	28	1.10**	28	0.58
Column/Environment	77	0.83**	77	0.67*
Group	4	8.91**	4	1.21
Line/Group	50	1.72**	52	0.84**
Genotypic High/Group	10	3.71**	12	1.05*
Genotypic Low/Group	10	1.00*	10	0.79
Phenotypic High/Group	10	0.58	10	0.81
Phenotypic Low/Group	10	1.80**	10	0.73
Random/Group	10	1.85**	10	0.74
Environment x Group	24	0.71*	24	0.50
Error	194	0.43	207	0.47
C.V.		8.9		8.6

\*, \*\* Significant at the 0.05 and 0.01 probability levels, respectively

Table A4. Analysis of variance for height across seven diverse environments in 2009 for Population 34 and 35.

Source of Variation	Population			
	34		35	
	df	Mean Square	df	Mean Square
Environment	6	863.91**	6	614.35**
Row/Environment	28	4.61*	28	4.24**
Column/Environment	77	6.30**	77	3.87**
Group	4	105.21**	4	30.93**
Line/Group	50	16.45**	52	11.06**
Genotypic High/Group	10	8.57**	12	9.00**
Genotypic Low/Group	10	9.93**	10	16.73**
Phenotypic High/Group	10	10.78**	10	3.56*
Phenotypic Low/Group	10	13.87**	10	10.67**
Random/Group	10	41.20**	10	13.20**
Environment x Group	24	7.09**	24	1.47
Error	194	2.94	207	1.58
C.V.		4.4		3.5

\*, \*\* Significant at the 0.05 and 0.01 probability levels, respectively

Table A5. Analysis of variance for seed size across four diverse environments in 2009 for Population 34 and 35.

Source of Variation	Population			
	34		35	
	df	Mean Square	df	Mean Square
Environment	3	48.24**	3	75.92**
Row/Environment	16	0.27	16	0.35
Column/Environment	44	0.37	44	0.66*
Group	4	6.08**	4	0.42
Line/Group	50	1.52**	52	1.32**
Genotypic High/Group	10	1.15**	12	1.76**
Genotypic Low/Group	10	0.87**	10	0.73
Phenotypic High/Group	10	1.58**	10	1.37**
Phenotypic Low/Group	10	1.13**	10	1.33**
Random/Group	10	3.10**	10	1.29**
Environment x Group	12	0.53	12	0.28
Error	84	0.32	94	0.42
C.V.		3.2		3.4

\*, \*\* Significant at the 0.05 and 0.01 probability levels, respectively

Table A6. Analysis of variance for maturity across six diverse environments in 2009 for Population 34 and 35.

Source of Variation	Population			
	34		35	
	df	Mean Square	df	Mean Square
Environment	5	2084.26**	5	1677.12**
Row/Environment	24	3.97**	24	4.27**
Column/Environment	66	2.47*	66	5.21**
Group	4	87.84**	4	37.31**
Line/Group	50	24.82**	52	17.61**
Genotypic High/Group	10	19.81**	12	13.78**
Genotypic Low/Group	10	3.35*	10	5.09**
Phenotypic High/Group	10	17.97**	10	14.90**
Phenotypic Low/Group	10	19.73**	10	38.90**
Random/Group	10	47.28**	10	17.59**
Environment x Group	20	3.89*	20	1.99
Error	160	1.63	170	1.46
C.V.		1.0		1.0

\*, \*\* Significant at the 0.05 and 0.01 probability levels, respectively

**APPENDIX B**

**ANALYSIS OF VARIANCE FOR GERMINATION**

The additive model for germination was:

$$Y_{ijk} = \mu + R_i + G_j + L_{(j)k} + e_{(ijk)}$$

where:

$Y_{ijk}$  = observed value of the  $k^{\text{th}}$  line in the  $j^{\text{th}}$  group in the  $i^{\text{th}}$  replication

$\mu$  = overall mean

$R_i$  = effect of the  $i^{\text{th}}$  replication,  $\text{NID}(0, \sigma^2_i)$

$G_j$  = effect of the  $j^{\text{th}}$  group

$L_{(j)k}$  = effect of the  $k^{\text{th}}$  line within the  $j^{\text{th}}$  group

$e_{(ijk)}$  = residual of the  $k^{\text{th}}$  line in the  $j^{\text{th}}$  group in the  $i^{\text{th}}$  replication

Table B1. Analysis of variance and expected mean squares for germination

Source of Variation	Expected Mean Squares
Replications	$\sigma_e^2 + gl\sigma_R^2$
Group	$\sigma_e^2 + rl\Phi_G$
Line/Group	$\sigma_e^2 + r\Phi_{L(G)}$
Genotypic High/Group	$\sigma_e^2 + rGI\text{PhPIRa}\Phi_{Gh(G)}$
Genotypic Low/Group	$\sigma_e^2 + rGh\text{PhPIRa}\Phi_{Gl(G)}$
Phenotypic High/Group	$\sigma_e^2 + rGhGI\text{PIRa}\Phi_{Ph(G)}$
Phenotypic Low/Group	$\sigma_e^2 + rGhGI\text{PIRa}\Phi_{Pl(G)}$
Random/Group	$\sigma_e^2 + rGhGI\text{PhPI}\Phi_{Ra(G)}$
Error	$\sigma_e^2$



Table B2. Analysis of variance for germination for Population 34 and 35.

Source of Variation	Population			
	34		35	
	df	Mean Square	df	Mean Square
Replications	3	13.3	3	279.42**
Group	4	58.30*	4	158.58**
Line/Group	50	61.24**	52	81.93**
Genotypic High/Group	10	74.80**	12	30.23
Genotypic Low/Group	10	58.29**	10	91.00**
Phenotypic High/Group	10	85.45**	10	26.96
Phenotypic Low/Group	10	40.67*	10	133.76**
Random/Group	10	47.00**	10	138.02**
Error	162	18.59	168	33.19
C.V.		4.9		7.0

\*, \*\* Significant at the 0.05 and 0.01 probability levels, respectively

**APPENDIX C**

**ANALYSIS OF VARIANCE FOR AGRONOMIC TRAITS AT INDIVIDUAL  
ENVIRONMENTS**

The additive model for all traits at individual environments in 2009 was:

$$Y_{ij} = \mu + G_i + L_{(i)j} + e_{(ij)}$$

where:

$Y_{ij}$  = observed value of the  $j^{\text{th}}$  line of the  $i^{\text{th}}$  group

$\mu$  = overall mean

$G_i$  = effect of the  $i^{\text{th}}$  group

$L_{(i)j}$  = effect of the  $j^{\text{th}}$  line within the  $i^{\text{th}}$  group

$e_{(ij)}$  = residual of the  $j^{\text{th}}$  line in the  $i^{\text{th}}$  group

Table C1. Analysis of variance and expected mean squares for individual environments for all traits.

Source of Variation	Expected Mean Squares
Group	$\sigma_e^2 + I\Phi_G$
Line/Group	$\sigma_e^2 + \Phi_{L(G)}$
Check/Group	$\sigma_e^2 + GhGlPhPIRa\Phi_{Gh(G)}$
Genotypic High/Group	$\sigma_e^2 + ChGlPhPIRa\Phi_{Gh(G)}$
Genotypic Low/Group	$\sigma_e^2 + ChGhPhPIRa\Phi_{Gl(G)}$
Phenotypic High/Group	$\sigma_e^2 + ChGhGlPIRa\Phi_{Ph(G)}$
Phenotypic Low/Group	$\sigma_e^2 + ChGhGlPIRa\Phi_{Pl(G)}$
Random/Group	$\sigma_e^2 + ChGhGlPhPI\Phi_{Ra(G)}$
Error	$\sigma_e^2$

Table C2. Analysis of variance mean squares for seed yield at individual locations in 2009 for Population 34.

Source of Variation	df	Location							
		Metamora, IL	Anna, OH	Napoleon, OH	Atlantic, IA	Hedrick, IA	Washington, IA	Gillman, IL	Pesotum, IL
Group	5	83.77	69.69	236.22	259.28	309.76*	369.73	242.91**	154.15**
Line/Group	53	34.50	13.22	18.83	30.86	38.72	48.77	17.93	14.81
Check/Group	3	11.08	14.17	8.47	44.92	2.68	78.04	7.79	38.61*
Genotypic High/Group	10	42.99	10.75	8.54	23.54	16.12	117.81	15.88	9.03
Genotypic Low/Group	10	72.59	15.52	25.97	34.16	54.43	22.21	27.39	16.64
Phenotypic High/Group	10	38.06	10.85	16.50	28.45	64.53	6.45	13.15	13.22
Phenotypic Low/Group	10	9.81	11.21	15.23	33.49	7.44	84.30	14.21	15.64
Random/Group	10	16.09	17.27	30.88	30.44	58.77	18.66	22.05	12.40
Error	2	21.22	17.33	24.91	16.21	5.05	24.20	2.21	1.12
C.V.		9.4	7.7	8.5	5.8	4.2	8.1	2.6	1.6

\*, \*\* Significant at the 0.05 and 0.01 probability levels, respectively

Table C3. Analysis of variance mean squares for lodging at individual locations in 2009 for Population 34.

Source of Variation	df	Location					
		Metamora, IL	Atlantic, IA	Hedrick, IA	Washington, IA	Gillman, IL	Pesotum, IL
Group	5	1.10	8.00	4.68	2.51	0.04	1.27
Line/Group	53	0.81	2.77	1.14	0.76	0.08	0.35
Check/Group	3	1.78	14.74	2.78	4.00	0.00	0.78
Genotypic High/Group	10	0.80	2.67	1.62	0.69	0.16	0.46
Genotypic Low/Group	10	0.89	1.07	1.25	0.60	0.09	0.09
Phenotypic High/Group	10	0.85	1.07	0.47	0.27	0.09	0.22
Phenotypic Low/Group	10	0.47	2.80	0.85	0.56	0.00	0.27
Random/Group	10	0.76	2.62	0.96	0.67	0.09	0.56
Error	2	0.00	0.00	0.25	0.00	0.00	0.50
C.V.		0.0	0.0	6.9	0.0	0.0	8.8

Table C4. Analysis of variance mean squares for height at individual locations in 2009 for Population 34.

Source of Variation	df	Location						
		Metamora, IL	Napoleon, OH	Atlantic, IA	Hedrick, IA	Washington, IA	Gillman, IL	Pesotum, IL
Group	5	29.35*	80.75*	19.75	36.43	30.85**	33.05	32.63
Line/Group	53	6.71	12.59	9.45	9.30	6.64*	6.34	7.50
Check/Group	3	33.61**	31.44*	13.44	36.00	24.78*	12.78	36.11
Genotypic High/Group	10	2.36	10.49	5.36	2.26	4.22	4.89	3.29
Genotypic Low/Group	10	3.09	9.82	5.47	2.56	2.16	3.96	4.85
Phenotypic High/Group	10	3.87	8.27	7.89	1.76	2.26	11.42	4.00
Phenotypic Low/Group	10	4.29	5.65	8.05	12.84	5.02*	3.89	2.87
Random/Group	10	11.87	23.05*	19.27	19.42	14.09*	5.62	13.89
Error	2	1.25	2.25	2.5	5.0	0.25	8.50	13.25
C.V.		2.9	3.4	3.6	6.2	1.4	8.4	10.2

\*, \*\* Significant at the 0.05 and 0.01 probability levels, respectively

Table C5. Analysis of variance mean squares for seed size at individual locations in 2009 for Population 34.

Source of Variation	df	Location			
		Metamora, IL	Napoleon, OH	Gillman, IL	Pesotum, IL
Group	5	2.22	2.14	6.89	1.86
Line/Group	53	0.75	0.77	1.96	1.31
Check/Group	3	3.60	1.34	18.07	3.27
Genotypic High/Group	10	0.38	0.50	0.70	1.71
Genotypic Low/Group	10	0.56	1.46	0.68	0.82
Phenotypic High/Group	10	0.65	0.45	0.81	1.17
Phenotypic Low/Group	10	0.56	0.66	0.86	0.45
Random/Group	10	1.00	0.96	1.79	1.79
Error	2	0.20	2.41	17.64	0.43
C.V.		2.6	9.3	23.46	3.5



Table C6. Analysis of variance mean squares for maturity at individual locations in 2009 for Population 34.

Source of Variation	df	Location					
		Atlantic, IA	Hedrick, IA	Washington, IA	Gillman, IL	Metamora, IL	Pesotum, IL
Group	5	21.37	20.18	51.61	33.63**	23.46*	25.51
Line/Group	53	9.48	6.70	13.64	10.39*	7.62*	10.07
Check/Group	3	46.28	26.94	79.11	64.78**	48.94**	40.50
Genotypic High/Group	10	7.42	1.20	11.56	6.07*	2.02	4.47
Genotypic Low/Group	10	2.22	3.67	3.22	2.13	3.27	2.09
Phenotypic High/Group	10	8.27	6.26	6.66	4.06	3.02	5.69
Phenotypic Low/Group	10	9.05	5.49	8.69	8.27*	5.76*	11.36
Random/Group	10	9.42	10.80	18.46	14.96*	11.62*	17.60
Error	2	0.00	0.00	0.00	0.25	0.25	0.00
C.V.		0.0	0.0	0.0	0.4	0.4	0.0

\*,\*\* Significant at the 0.05 or 0.01 probability levels, respectively

Table C7. Analysis of variance mean squares for yield at individual locations in 2009 for Population 35.

Source of Variation	df	Location							
		Atlantic, IA	Hedrick, IA	Washington, IA	Emery, IL	Metamora, IL	Pesotum, IL	Anna, OH	Napoleon, OH
Group	5	61.78	52.32	58.28	41.08*	10.83	38.63	34.71	65.35
Line/Group	56	20.76	29.82	9.93	22.57*	14.14	22.95	8.74	15.44
Check/Group	4	24.50	38.21	11.82	56.00*	5.45	12.13	11.54	9.91
Genotypic High/Group	12	15.07	19.46	8.18	10.86	12.44	16.35	5.15	19.70
Genotypic Low/Group	10	24.51	27.11	11.46	30.73*	20.98	28.93	8.51	13.60
Phenotypic High/Group	10	14.53	37.23	4.09	20.46*	11.26	29.57	3.45	17.18
Phenotypic Low/Group	10	23.29	36.95	18.42	29.41*	11.56	12.08	7.80	15.93
Random/Group	10	26.04	27.05	7.08	11.67	18.27	33.46	18.40	12.17
Error	2	6.05	36.57	8.90	0.94	6.51	5.93	16.47	31.77
C.V.		3.8	10.2	4.8	1.6	5.1	4.1	7.0	8.8

\* Significant at the 0.05 and 0.01 probability level

Table C8. Analysis of variance mean squares for lodging at individual locations in 2009 for Population 35.

Source of Variation	df	Location					
		Atlantic, IA	Hedrick, IA	Washington, IA	Emery, IL	Metamora, IL	Pesotum, IL
Group	5	2.92	0.35	0.53	0.12	0.18	0.16
Line/Group	56	1.93	0.78	0.54	0.22	16.34	0.28
Check/Group	4	6.59*	2.09	2.09	0.18	0.73	0.68
Genotypic High/Group	12	1.47	0.31	0.58	0.27	0.67	0.23
Genotypic Low/Group	10	2.16	0.29	0.22	0.22	0.27	0.22
Phenotypic High/Group	10	2.29	0.42	0.22	0.22	0.22	0.26
Phenotypic Low/Group	10	1.47	0.29	0.49	0.22	0.26	0.27
Random/Group	10	0.49	0.49	0.56	0.16	0.27	0.26
Error	2	0.25	0.25	0.25	0.50	0.25	0.5
C.V.		7.2	6.3	6.2	8.5	5.9	8.2

\* Significant at the 0.05 probability level

Table C9. Analysis of variance mean squares for height at individual locations in 2009 for Population 35.

Source of Variation	df	Location						
		Atlantic, IA	Hedrick, IA	Washington, IA	Emery, IL	Metamora, IL	Pesotum, IL	Napoleon, OH
Group	5	11.69	5.46	6.99	6.82	2.96	7.10	7.78*
Line/Group	56	6.18	4.52	5.01	3.62	6.35	4.82	4.08
Check/Group	4	18.21	4.43	14.68*	4.80	8.88	24.71*	13.72*
Genotypic High/Group	12	5.19	5.23	2.83	1.97	7.31	5.09	1.73
Genotypic Low/Group	10	6.07	2.07	5.42	5.06	6.62	3.07	5.47*
Phenotypic High/Group	10	5.07	2.27	2.76	2.47	4.07	1.42	1.89
Phenotypic Low/Group	10	5.25	6.62	3.82	4.96	5.67	2.66	5.27*
Random/Group	10	4.67	6.27	6.76	3.47	6.86	5.06	2.66
Error	2	1.00	1.00	0.50	10.25	16.25	1.00	0.25
C.V.		2.4	3.0	2.2	9.8	11.4	3.0	1.3

\* Significant at the 0.05 probability level

Table C10. Analysis of variance mean squares for seed size at individual locations in 2009 for Population 34.

Source of Variation	df	Location			
		Emery, IL	Metamora, IL	Pesotum, IL	Napoleon, OH
Group	5	1.04	0.43	0.84	0.73
Line/Group	56	0.90	0.69	1.17	0.90
Check/Group	4	0.81	1.25	1.81	0.88
Genotypic High/Group	12	1.33	0.75	1.49	1.16
Genotypic Low/Group	10	1.08	0.24	0.62	0.25
Phenotypic High/Group	10	0.73	0.85	0.39	0.71
Phenotypic Low/Group	10	0.68	0.66	1.78	1.65
Random/Group	10	0.60	0.71	1.27	0.70
Error	2	0.59	0.45	0.27	0.43
C.V.		4.0	4.0	2.7	3.8

Table C11. Analysis of variance mean squares for maturity at individual locations in 2009 for Population 34.

Source of Variation	df	Location					
		Atlantic, IA	Hedrick, IA	Washington, IA	Emery, IL	Metamora, IL	Pesotum, IL
Group	5	82.65	8.80	19.80*	11.72	15.35	10.11
Line/Group	56	7.64	7.59	12.66*	7.22	7.64	9.88
Check/Group	4	27.96	20.86	34.75*	41.23*	34.38*	44.23
Genotypic High/Group	12	3.94	6.42	7.97	3.58	3.40	4.69
Genotypic Low/Group	10	5.96	3.36	3.96	4.67	2.85	5.66
Phenotypic High/Group	10	5.87	4.67	14.67*	5.76	3.76	4.82
Phenotypic Low/Group	10	9.22	11.56	19.60*	3.76	13.16	11.49
Random/Group	10	5.82	6.87	9.16	5.47	5.20	10.06
Error	2	2.50	0.00	0.50	1.25	1.25	4.25
C.V.		1.2	0.0	0.5	0.9	0.9	1.8

\*,\*\* Significant at the 0.05 or 0.01 probability levels, respectively

**APPENDIX D**

**LINE PERFORMANCE AT EACH LOCATION**

Table D1. Seed yield by line at individual locations of Population 34 in 2009.

Line	Group	Atlantic, IA	Hedrick, IA	Washington, IA	Gillman, IL	Metamora, IL	Pesotum, IL	Anna, OH	Napoleon, OH
34-0414	GH†	3830‡	3622	2452	4159	3601	4515	4347	3884
34-0692	GH	4569	3924	-§	4650	3494	4710	4172	3642
34-0724	GH/PH	4005	4320	4582	4267	3548	4354	4220	4092
34-0777	GH	3810	4112	4132	3877	3393	4253	4072	3904
34-1046	GH	4562	4125	-	3937	3393	4502	3991	3487
34-1055	GH	3964	3783	4697	4246	3830	4172	3870	3709
34-1058	GH	4220	3924	4697	4347	3400	4891	4448	3272
34-1553	GH	3480	3366	4307	4569	3178	4354	4388	3662
34-1627	GH	4005	3769	4535	3951	2331	4535	3984	3756
34-1730	GH	4011	4139	4663	4361	4038	4535	4085	3648
34-1757	GH	4300	3837	4784	3904	3104	4488	3910	3534
34-0351	GL	3239	2553	3205	3030	3118	4125	3608	3454
34-0450	GL	3622	3931	3944	4065	2970	4408	3454	3400
34-0485	GL	4193	3091	3984	3675	1915	4320	4112	3810
34-0503	GL	3695	3413	3951	3716	3333	3991	3501	3312
34-0760	GL	3366	3333	3682	3575	3239	3924	3218	2970
34-0873	GL	4428	3991	4213	4172	2936	4636	4435	3454
34-1242	GL	4031	3501	4327	4058	2298	4461	3890	3541
34-1288	GL	3857	3769	4139	4193	3151	4656	3796	3742
34-1716	GL	4361	4374	3984	4119	3857	4596	3830	3420



Table D1. Continued.

Line	Group	Atlantic, IA	Hedrick, IA	Washington, IA	Gillman, IL	Metamora, IL	Pesotum, IL	Anna, OH	Napoleon, OH
34-1806	GL	3561	3642	4260	3924	3803	4623	4052	3816
34-1897	GL	3648	3212	3776	3615	3312	4650	3924	3837
34-0506	PH	3575	3460	4596	3910	3615	5019	3810	3460
34-0518	PH	3904	3601	4179	3689	2217	4650	3884	3507
34-0626	PH	4058	4300	4293	3769	3259	4455	4273	3709
34-0662	PH	4186	2473	4273	4179	3548	4670	4643	3924
34-0754	PH	4233	4186	4623	4206	3454	4112	4670	4085
34-0947	PH	4562	4132	4629	3642	3467	4253	4307	3655
34-1118	PH	4609	3608	4596	4206	3353	4468	4112	3568
34-1157	PH	4388	4213	4555	4320	3467	4414	4488	3951
34-1340	PH	4522	3790	4643	4018	3763	4576	4293	3863
34-1440	PH	3622	3910	4374	3870	3084	4670	4388	3756
34-0297	PL	3064	2977	-	3077	2963	3272	3185	3286
34-0378	PL	3675	3165	3588	3514	3144	3635	3292	3514
34-0405	PL	3198	3037	2916	3333	2869	3843	3165	3259
34-0538	PL	3124	2654	3521	3487	3178	3776	3205	3057
34-0778	PL	3870	2916	3588	3259	3091	4159	3044	3669
34-1256	PL	3608	2983	3937	3554	3393	3830	3722	3480
34-1426	PL	3245	-	3937	3124	3077	4072	3507	3588
34-1486	PL	3588	2735	3776	3595	2889	4078	3642	3467

Table D1. Continued.

Line	Group	Atlantic, IA	Hedrick, IA	Washington, IA	Gillman, IL	Metamora, IL	Pesotum, IL	Anna, OH	Napoleon, OH
34-1609	PL	2627	2869	3279	2802	2688	3541	3366	2956
34-1654	PL	2748	3259	3003	3071	2694	4018	3870	3232
34-1808	PL	3138	2849	1901	3104	2990	3910	3521	3192
34-0467	RA	3494	4596	4320	3575	3373	4690	4226	3776
34-0679	RA	4293	4025	4703	4052	3407	4334	4529	3521
34-0761	RA	3863	4018	-	3722	3104	4193	4441	3494
34-1029	RA	3971	4072	4616	3857	3642	4603	4112	3588
34-1032	RA	3662	3608	4267	4394	3286	4623	4374	3561
34-1140	RA	4065	4502	4220	4246	3232	4488	4522	3790
34-1333	RA	4253	4065	4206	3736	3393	4307	4058	4293
34-1625	RA	4401	3292	4085	4112	2950	4354	3561	3884
34-1666	RA	3360	2802	3742	3306	2788	3857	3373	3245
34-1779	RA	4468	4119	3890	4112	3487	4253	4045	3890
34-1847	RA	4199	4031	4260	3924	3648	4482	4038	3521
CV		7	4	8	3	10	4	8	9
s		471	523	587	404	413	342	285	409
SE		64	71	80	55	56	47	39	56

†GH=genotypic high, GL=genotypic low, PH=phenotypic high, PL=phenotypic low, RA=random

‡Seed yield expressed in kg ha<sup>-1</sup>

§A dash sign indicates data not reported

Table D2. Lodging by line at individual locations of Population 34 in 2009.

Line	Group	Atlantic, IA	Hedrick, IA	Washington, IA	Gillman, IL	Metamora, IL	Pesotum, IL
34-0414	GH†	4.5‡	3.0	2.5	1.0	1.5	1.5
34-0692	GH	1.5	1.5	1.5	1.0	1.0	1.5
34-0724	GH/PH	2.5	2.5	1.5	1.0	1.0	1.5
34-0777	GH	4.0	3.0	2.5	1.5	2.0	2.5
34-1046	GH	3.0	1.5	1.5	1.0	1.5	2.0
34-1055	GH	3.0	2.0	2.0	1.0	1.5	1.5
34-1058	GH	3.0	2.0	2.0	1.0	1.5	1.5
34-1553	GH	4.0	3.5	2.5	1.5	2.5	2.0
34-1627	GH	3.5	2.5	2.5	1.0	1.5	1.5
34-1730	GH	3.0	2.0	2.0	1.0	1.5	1.5
34-1757	GH	3.5	2.5	2.0	1.0	1.0	1.5
34-0351	GL	2.5	1.0	1.0	1.0	1.0	1.0
34-0450	GL	3.0	1.5	1.5	1.5	1.5	1.5
34-0485	GL	3.0	1.0	1.5	1.0	2.0	1.5
34-0503	GL	3.0	2.0	2.0	1.0	2.0	1.5
34-0760	GL	2.0	2.0	1.0	1.0	1.0	1.5
34-0873	GL	2.5	2.5	1.5	1.0	2.0	1.5
34-1242	GL	3.5	2.5	2.0	1.0	2.0	1.5
34-1288	GL	3.0	1.5	2.0	1.0	2.0	1.5
34-1716	GL	2.0	1.5	1.5	1.0	1.0	1.5

Table D2. Continued.

Line	Group	Atlantic, IA	Hedrick, IA	Washington, IA	Gillman, IL	Metamora, IL	Pesotum, IL
34-1806	GL	3.5	2.0	1.5	1.0	1.5	1.5
34-1897	GL	2.5	1.0	1.0	1.0	1.0	1.5
34-0506	PH	4.5	2.5	1.5	1.0	2.0	1.5
34-0518	PH	3.0	2.0	1.5	1.5	2.0	1.5
34-0626	PH	3.0	2.0	2.0	1.0	1.5	2.0
34-0662	PH	3.0	2.0	1.5	1.0	2.0	2.0
34-0754	PH	3.5	2.0	2.0	1.0	2.0	1.5
34-0947	PH	3.5	2.0	2.0	1.0	1.5	1.5
34-1118	PH	3.5	2.5	2.0	1.0	2.5	2.0
34-1157	PH	3.0	2.5	2.0	1.0	1.5	1.5
34-1340	PH	3.5	2.0	1.5	1.0	1.5	1.5
34-1440	PH	3.0	3.0	1.5	1.0	1.0	1.5
34-0297	PL	3.0	2.0	1.0	1.0	1.0	1.0
34-0378	PL	2.0	2.0	1.5	1.0	1.5	1.0
34-0405	PL	2.0	1.0	1.5	1.0	1.5	1.0
34-0538	PL	3.0	1.5	1.5	1.0	1.5	1.5
34-0778	PL	1.5	1.5	1.5	1.0	1.0	1.5
34-1256	PL	4.0	2.5	2.0	1.0	2.0	1.5
34-1426	PL	3.5	-.§	2.0	1.0	1.0	1.5
34-1486	PL	2.0	1.5	1.5	1.0	1.0	1.5
34-1609	PL	3.0	1.5	1.0	1.0	1.5	1.0

Table D2. Continued.

Line	Group	Atlantic, IA	Hedrick, IA	Washington, IA	Gillman, IL	Metamora, IL	Pesotum, IL
34-1654	PL	2.0	1.0	1.0	1.0	1.0	1.0
34-1808	PL	1.5	1.5	1.0	1.0	1.0	1.5
34-0467	RA	2.5	2.5	1.5	1.0	1.5	1.5
34-0679	RA	3.0	1.5	1.5	1.0	1.0	1.0
34-0761	RA	3.5	2.5	2.5	1.0	2.0	2.0
34-1029	RA	3.0	1.5	1.5	1.0	1.5	1.5
34-1032	RA	4.5	2.5	2.0	1.0	2.5	2.0
34-1140	RA	3.5	2.0	2.0	1.0	2.0	2.0
34-1333	RA	3.0	2.0	2.0	1.5	1.5	1.5
34-1625	RA	2.5	2.0	1.5	1.0	1.5	1.5
34-1666	RA	2.0	1.0	1.0	1.0	1.0	1.0
34-1779	RA	2.5	2.0	2.0	1.0	1.5	1.5
34-1847	RA	1.5	1.5	1.5	1.0	1.5	2.0
CV		0.0	7.0	0.0	0.0	0.0	8.9
s		1.7	1.2	0.9	0.3	0.9	0.6
SE		0.2	0.2	0.1	0.0	0.1	0.1

†GH=genotypic high, GL=genotypic low, PH=phenotypic high, PL=phenotypic low,  
RA=random

‡1 = all plants erect, 5 = all plant prostrate

§A dash sign indicates data not reported

Table D3. Height by line at individual locations of Population 34 in 2009.

Line	Group	Atlantic, IA	Hedrick, IA	Washington, IA	Gillman, IL	Metamora, IL	Pesotum, IL	Napoleon, OH
34-0414	GH†	117‡	91	89	84	104	89	104
34-0692	GH	112	97	94	86	97	94	117
34-0724	GH/PH	102	97	94	91	102	97	114
34-0777	GH	117	102	104	99	109	102	132
34-1046	GH	122	97	89	91	99	97	109
34-1055	GH	112	102	97	102	107	102	124
34-1058	GH	117	97	99	99	104	97	117
34-1553	GH	122	102	102	94	109	99	124
34-1627	GH	117	97	91	94	104	99	127
34-1730	GH	117	102	94	97	104	102	124
34-1757	GH	109	91	89	99	102	89	117
34-0351	GL	119	81	84	76	94	91	114
34-0450	GL	109	97	84	86	97	89	107
34-0485	GL	109	86	84	86	97	89	114
34-0503	GL	117	86	81	76	99	86	104
34-0760	GL	104	86	81	81	89	81	99
34-0873	GL	117	91	89	86	97	97	109
34-1242	GL	122	91	91	89	94	89	117
34-1288	GL	112	91	91	89	107	102	124
34-1716	GL	114	89	86	76	94	91	104

Table D3. Continued.

Line	Group	Atlantic, IA	Hedrick, IA	Washington, IA	Gillman, IL	Metamora, IL	Pesotum, IL	Napoleon, OH
34-1806	GL	117	86	89	84	99	84	122
34-1897	GL	104	86	84	86	94	89	107
34-0506	PH	109	91	97	84	104	94	127
34-0518	PH	112	94	91	84	99	84	119
34-0626	PH	114	102	99	97	104	97	132
34-0662	PH	107	94	91	102	102	94	119
34-0754	PH	119	91	99	104	107	99	122
34-0947	PH	104	102	91	81	94	89	112
34-1118	PH	114	97	97	91	112	97	130
34-1157	PH	124	97	99	76	102	102	124
34-1340	PH	104	97	91	86	99	89	109
34-1440	PH	117	97	102	89	109	94	117
34-0297	PL	112	102	89	86	97	86	107
34-0378	PL	99	91	79	84	84	79	94
34-0405	PL	104	81	89	89	94	89	99
34-0538	PL	107	71	81	84	91	86	107
34-0778	PL	112	91	86	81	94	89	104
34-1256	PL	124	91	99	91	102	94	117
34-1426	PL	102	-§	81	89	102	86	107
34-1486	PL	99	76	81	84	91	84	104

Table D3. Continued.

Line	Group	Atlantic, IA	Hedrick, IA	Washington, IA	Gillman, IL	Metamora, IL	Pesotum, IL	Napoleon, OH
34-1609	PL	109	81	84	79	91	94	112
34-1654	PL	109	91	89	74	97	89	104
34-1808	PL	107	81	84	84	89	89	109
34-0467	RA	109	102	99	99	102	94	114
34-0679	RA	109	102	97	91	107	94	114
34-0761	RA	117	107	99	89	107	97	127
34-1029	RA	114	86	86	86	89	89	107
34-1032	RA	127	86	91	102	107	97	119
34-1140	RA	114	107	91	94	107	94	117
34-1333	RA	124	102	99	81	99	99	112
34-1625	RA	104	91	89	89	99	84	119
34-1666	RA	89	76	71	89	89	69	89
34-1779	RA	109	91	79	97	84	81	104
34-1847	RA	97	76	79	97	89	79	89
CV		4	6	1	8	3	10	3
s		8	8	7	7	7	7	10
SE		1	1	1	1	1	1	1

†GH=genotypic high, GL=genotypic low, PH=phenotypic high, PL=phenotypic low, RA=random

‡Plant height in cm from ground to terminal node

§A dash sign indicates data not reported



Table D4. Seed size by line at individual locations of Population 34 in 2009.

Line	Group	Gillman, IL	Metamora, IL	Pesotum, IL	Napoleon, OH
34-0414	GH†	17.2‡	17.2	18.8	15.4
34-0692	GH	18.9	17.2	19.7	17.0
34-0724	GH/PH	16.6	15.4	16.4	16.2
34-0777	GH	16.9	16.3	16.9	16.7
34-1046	GH	17.5	16.3	17.9	15.9
34-1055	GH	18.3	17.2	19.7	15.3
34-1058	GH	18.7	17.2	20.7	16.6
34-1553	GH	17.1	16.3	17.5	16.5
34-1627	GH	17.9	17.2	18.8	15.7
34-1730	GH	18.7	17.2	19.1	17.4
34-1757	GH	16.9	17.2	17.8	17.1
34-0351	GL	17.3	16.3	17.9	15.8
34-0450	GL	19.6	17.2	19.5	17.7
34-0485	GL	18.3	17.2	19.6	17.4
34-0503	GL	18.0	17.2	18.5	16.7
34-0760	GL	17.8	17.2	18.7	16.1
34-0873	GL	18.7	16.3	19.3	16.6
34-1242	GL	19.6	18.1	19.9	-§
34-1288	GL	18.7	18.1	19.8	14.2
34-1716	GL	18.6	18.1	20.5	16.6

Table D4. Continued.

Line	Group	Gillman, IL	Metamora, IL	Pesotum, IL	Napoleon, OH
34-1806	GL	17.2	16.3	17.5	14.9
34-1897	GL	17.6	16.3	19.5	16.8
34-0506	PH	18.9	18.1	20.1	16.7
34-0518	PH	17.0	16.3	17.8	16.2
34-0626	PH	18.7	18.1	18.9	17.7
34-0662	PH	19.3	17.2	19.5	-
34-0754	PH	18.5	17.2	19.4	18.0
34-0947	PH	17.7	17.2	19.4	-
34-1118	PH	19.2	17.2	19.6	17.8
34-1157	PH	18.0	17.2	18.4	17.1
34-1340	PH	19.0	17.2	19.8	17.4
34-1440	PH	18.8	18.1	19.6	17.4
34-0297	PL	-	17.2	18.5	17.2
34-0378	PL	18.1	16.3	18.3	16.9
34-0405	PL	18.5	17.2	19.4	16.9
34-0538	PL	15.6	15.4	17.6	15.7
34-0778	PL	17.9	16.3	18.3	16.5
34-1256	PL	17.1	16.3	17.6	16.4
34-1426	PL	15.9	15.4	18.1	15.5
34-1486	PL	17.3	17.2	18.4	16.4
34-1609	PL	16.5	15.4	17.7	14.5

Table D4. Continued.

Line	Group	Gillman, IL	Metamora, IL	Pesotum, IL	Napoleon, OH
34-1654	PL	17.1	17.2	19.6	-
34-1808	PL	17.2	16.3	17.9	15.8
34-0467	RA	17.8	16.3	19.4	16.1
34-0679	RA	18.7	17.2	19.7	-
34-0761	RA	17.6	17.2	18.9	16.6
34-1029	RA	19.0	17.2	18.7	17.8
34-1032	RA	19.8	18.1	19.9	16.7
34-1140	RA	18.6	17.2	18.6	16.5
34-1333	RA	17.2	16.3	17.2	16.3
34-1625	RA	18.3	17.2	18.6	17.3
34-1666	RA	18.4	16.3	18.4	16.6
34-1779	RA	21.2	19.1	20.8	18.6
34-1847	RA	21.2	19.1	22.2	18.9
CV		23.5	2.6	3.5	9.3
s		1.1	0.8	1.1	0.9
SE		0.2	0.1	0.2	0.1

†GH=genotypic high, GL=genotypic low, PH=phenotypic high,  
 PL=phenotypic low, RA=random

‡Seed size expressed in g 100 seed<sup>-1</sup>

§A dash sign indicates data not reported

Table D5. Maturity by line at individual locations of Population 34 in 2009.

Line	Group	Atlantic, IA	Hedrick, IA	Washington, IA	Gillman, IL	Metamora, IL	Pesotum, IL
34-0414	GH†	140‡	131	142	136	128	123
34-0692	GH	134	128	133	130	125	117
34-0724	GH/PH	137	129	134	132	125	121
34-0777	GH	139	130	141	135	127	123
34-1046	GH	137	129	135	133	126	120
34-1055	GH	132	128	135	130	125	121
34-1058	GH	135	129	138	134	127	122
34-1553	GH	137	130	141	135	128	122
34-1627	GH	139	129	140	133	128	119
34-1730	GH	132	127	133	128	124	117
34-1757	GH	137	129	137	132	126	121
34-0351	GL	134	123	131	128	124	117
34-0450	GL	135	129	133	129	124	118
34-0485	GL	134	124	132	128	124	120
34-0503	GL	134	126	133	128	125	118
34-0760	GL	135	127	132	128	125	117
34-0873	GL	132	127	132	130	122	116
34-1242	GL	137	125	134	128	124	117
34-1288	GL	136	124	132	131	124	115
34-1716	GL	132	127	131	127	125	118

Table D5. Continued.

Line	Group	Atlantic, IA	Hedrick, IA	Washington, IA	Gillman, IL	Metamora, IL	Pesotum, IL
34-1806	GL	134	126	136	132	122	117
34-1897	GL	134	123	129	129	119	115
34-0506	PH	139	129	139	135	128	121
34-0518	PH	134	124	132	130	125	116
34-0626	PH	136	129	138	133	127	118
34-0662	PH	130	122	132	128	122	115
34-0754	PH	137	125	138	131	126	118
34-0947	PH	131	128	133	130	124	120
34-1118	PH	138	129	138	132	127	119
34-1157	PH	136	128	135	129	125	117
34-1340	PH	138	129	136	133	127	123
34-1440	PH	135	129	137	132	127	120
34-0297	PL	136	129	136	129	126	121
34-0378	PL	135	129	136	132	126	118
34-0405	PL	130	127	131	128	125	116
34-0538	PL	134	124	133	130	124	117
34-0778	PL	134	124	131	130	124	118
34-1256	PL	137	128	135	132	126	121
34-1426	PL	132	127	133	131	125	119
34-1486	PL	134	123	129	127	125	117

Table D5. Continued.

Line	Group	Atlantic, IA	Hedrick, IA	Washington, IA	Gillman, IL	Metamora, IL	Pesotum, IL
34-1609	PL	128	124	128	124	121	112
34-1654	PL	129	127	131	125	121	116
34-1808	PL	130	123	128	125	119	110
34-0467	RA	134	128	131	128	122	113
34-0679	RA	136	129	136	135	126	122
34-0761	RA	138	130	140	134	127	119
34-1029	RA	131	123	131	128	122	117
34-1032	RA	137	124	135	131	125	117
34-1140	RA	130	122	129	127	119	111
34-1333	RA	138	130	140	136	128	123
34-1625	RA	136	126	134	133	126	118
34-1666	RA	132	123	132	127	124	117
34-1779	RA	135	129	133	129	125	120
34-1847	RA	130	122	126	124	117	110
CV		0	0	0	1	1	0
s		3	3	4	3	3	3
SE		0	0	1	0	0	0

†GH=genotypic high, GL=genotypic low, PH=phenotypic high, PL=phenotypic low,  
RA=random

‡Days after planting

Table D6. Seed yield by line at individual locations of Population 35 in 2009.

Line	Group	Atlantic, IA	Hedrick, IA	Washington, IA	Emery, IL	Metamora, IL	Pesotum, IL	Anna, OH	Napoleon, OH
35-2039	GH†	4381‡	4381	4421	4052	3178	4576	4152	4112
35-2152	GH	4401	4280	4381	4287	3541	3642	3910	4542
35-2343	GH/RA	4099	4125	4159	3964	3494	4508	4179	4555
35-2455	GH	4367	4549	4797	4508	3575	4334	4119	4522
35-2592	GH/PH	4273	4508	4280	4159	3050	4502	4199	4549
35-2646	GH	4092	3998	4435	3689	3050	4105	4031	4515
35-2671	GH	4629	3581	4092	4058	3669	4334	3816	3904
35-2820	GH	4488	3803	4320	4401	3124	3951	3722	4495
35-2972	GH	4770	3971	4569	4300	3527	4623	3884	5153
35-3049	GH	4932	4273	4388	3964	3239	4293	4179	4623
35-3071	GH/RA	4643	3823	4374	4240	3003	4260	3931	4616
35-3190	GH	4603	3957	4435	4031	3212	4213	3984	4267
35-3787	GH	4797	3843	4092	4334	3527	4435	4025	4267
35-1983	GL	4542	3333	4119	-§	3662	3904	3769	4421
35-2062	GL	4112	4052	4099	3138	3346	4058	3447	4448
35-2184	GL	4589	4515	4018	3917	2681	4092	4031	4146
35-2244	GL	4193	4273	4253	3742	3454	4300	3595	3971
35-2450	GL	4347	4354	4730	3924	3635	3144	4065	4421
35-2485	GL	4703	4099	4159	4394	3830	4267	3931	4267
35-3253	GL	4455	3964	4220	3991	3366	3722	3581	4166

Table D6. Continued.

Line	Group	Atlantic, IA	Hedrick, IA	Washington, IA	Emery, IL	Metamora, IL	Pesotum, IL	Anna, OH	Napoleon, OH
35-3388	GL	4576	4031	4186	4226	3218	4515	3910	4226
35-3478	GL	4038	4179	4125	4085	3662	4078	3830	4085
35-3731	GL	3615	3521	4018	4455	3279	3964	3910	4777
35-3801	GL	3957	4260	3796	3897	3413	4220	3749	3910
35-2235	PH	4280	4320	4428	4011	3447	4448	4125	4287
35-2323	PH	4670	4818	4340	4300	3467	4058	3904	4515
35-2684	PH	5059	4025	4502	4468	3373	4455	4085	4031
35-2690	PH	4542	3474	4172	4226	3554	4764	4092	4428
35-2701	PH	4764	4031	4421	4293	3642	4643	3951	4280
35-3018	PH	4253	3682	4589	4367	3776	4650	4045	5086
35-3544	PH	4603	4576	4616	4280	3050	3608	3790	4663
35-3545	PH	4589	3957	4320	4435	3299	4240	4186	4495
35-3927	PH	4885	4529	4374	3360	3507	3816	3984	4414
35-3993	PH	4582	4018	4529	4206	3292	4475	3964	4172
35-2213	PL	4112	4609	4045	4105	3265	4300	3682	4381
35-2270	PL	4159	4025	3601	3702	2923	4031	3742	3910
35-2328	PL	4804	3756	4307	4656	3742	4616	3931	4092
35-2344	PL	4589	4011	4246	4441	3178	4092	3964	4408
35-2366	PL	4320	4146	4643	4253	3554	4193	3924	4388
35-2666	PL	4549	3192	3642	3386	3178	3910	3460	3588



Table D6. Continued.

Line	Group	Atlantic, IA	Hedrick, IA	Washington, IA	Emery, IL	Metamora, IL	Pesotum, IL	Anna, OH	Napoleon, OH
35-2845	PL/RA	3575	3561	4099	3924	3366	4193	3648	4011
35-2874	PL	4038	3944	4092	3850	3252	4146	3716	3736
35-2951	PL	4246	3393	4139	4233	3312	4058	3937	4011
35-3726	PL	4199	3769	4146	3716	3413	3736	3433	4031
35-3757	PL	4186	3373	4005	4105	3037	4367	3648	3870
35-2474	RA	4354	4052	4025	4105	3064	4025	3937	4146
35-2589	RA	4549	4495	4058	4112	3937	4226	3648	4522
35-2728	RA	3937	3736	3830	3816	3218	3749	3837	4562
35-2808	RA	4428	3386	4179	4394	3742	4435	4616	4273
35-3497	RA	4576	3729	4139	3548	3400	3944	3897	4273
35-3536	RA	4031	4025	4186	4052	3601	3890	3843	4220
35-3707	RA	4643	3306	4172	3931	3279	4488	3749	4770
35-3980	RA	4414	4038	3722	4139	3648	3185	3615	4542
CV		4	10	5	2	5	2	7	9
s		324	379	249	320	247	326	222	300
SE		45	52	34	44	34	45	31	41

†GH=genotypic high, GL=genotypic low, PH=phenotypic high, PL=phenotypic low, RA=random

‡Seed yield expressed in kg ha<sup>-1</sup>

§A dash sign indicates data not reported

Table D7. Lodging by line at individual locations of Population 35 in 2009.

Line	Group	Atlantic, IA	Hedrick, IA	Washington, IA	Emery, IL	Metamora, IL	Pesotum, IL
35-2039	GH†	2.0‡	1.5	2.0	1.0	1.5	1.0
35-2152	GH	2.0	1.5	2.0	1.5	1.5	1.0
35-2343	GH/RA	1.5	1.5	1.0	1.0	1.0	1.0
35-2455	GH	3.5	2.0	1.5	1.5	1.5	1.5
35-2592	GH/PH	1.5	1.0	1.5	1.0	1.0	1.0
35-2646	GH	2.5	1.5	1.5	1.0	1.5	1.0
35-2671	GH	2.0	1.5	1.0	1.5	1.0	1.0
35-2820	GH	1.0	1.0	1.0	1.0	1.0	1.0
35-2972	GH	2.0	1.0	1.5	1.0	1.0	1.0
35-3049	GH	1.5	1.5	1.0	1.5	1.0	1.5
35-3071	GH/RA	2.0	1.5	1.5	1.5	1.5	1.5
35-3190	GH	1.5	1.5	1.5	1.5	1.0	1.0
35-3787	GH	2.0	1.5	2.0	1.5	1.5	1.5
35-1983	GL	1.5	1.5	1.0	1.0	1.5	1.0
35-2062	GL	1.5	1.5	1.5	1.5	1.5	1.0
35-2184	GL	1.5	1.5	1.5	1.5	1.5	1.5
35-2244	GL	1.5	1.5	1.5	1.5	1.5	1.5
35-2450	GL	2.0	1.0	1.0	1.0	1.0	1.0
35-2485	GL	1.5	1.0	1.0	1.5	1.0	1.0
35-3253	GL	1.0	1.5	1.5	1.0	1.0	1.0

Table D7. Continued.

Line	Group	Atlantic, IA	Hedrick, IA	Washington, IA	Emery, IL	Metamora, IL	Pesotum, IL
35-3388	GL	3.5	1.5	1.5	1.5	1.0	1.0
35-3478	GL	2.0	1.5	1.5	1.5	1.5	1.5
35-3731	GL	3.0	1.5	1.5	1.5	1.0	1.0
35-3801	GL	2.0	2.0	1.5	1.5	1.5	1.0
35-2235	PH	3.0	1.5	2.0	1.5	1.0	1.0
35-2323	PH	3.5	1.5	1.5	1.5	1.0	1.0
35-2684	PH	2.0	2.0	1.5	1.5	1.0	1.0
35-2690	PH	2.5	1.5	1.5	1.5	1.5	1.0
35-2701	PH	3.5	1.5	1.5	1.5	1.0	1.5
35-3018	PH	3.0	2.0	2.0	1.5	1.5	1.5
35-3544	PH	2.5	2.0	2.0	1.5	1.0	1.0
35-3545	PH	1.5	2.0	1.5	1.0	1.0	1.0
35-3927	PH	2.5	1.5	1.5	1.0	1.5	1.5
35-3993	PH	1.5	1.5	1.5	1.5	1.0	1.5
35-2213	PL	2.5	2.0	2.0	1.5	1.0	1.5
35-2270	PL	1.5	1.5	2.0	1.5	1.0	1.5
35-2328	PL	1.5	2.0	1.0	1.0	1.5	1.0
35-2344	PL	1.0	1.5	1.0	1.5	1.0	1.0
35-2366	PL	1.5	1.5	1.5	1.5	1.0	1.0
35-2666	PL	2.0	1.5	1.5	1.5	1.5	1.5

Table D7. Continued.

Line	Group	Atlantic, IA	Hedrick, IA	Washington, IA	Emery, IL	Metamora, IL	Pesotum, IL
35-2845	PL/RA	2.5	1.5	1.5	1.5	1.5	1.5
35-2874	PL	1.0	1.5	1.0	1.0	1.0	1.0
35-2951	PL	2.5	1.5	1.5	1.5	1.5	1.5
35-3726	PL	1.0	1.0	1.5	1.0	1.0	1.0
35-3757	PL	2.0	1.5	1.5	1.5	1.0	1.5
35-2474	RA	2.0	2.0	1.5	1.5	1.0	1.0
35-2589	RA	1.5	1.5	2.0	1.5	1.0	1.5
35-2728	RA	2.0	2.0	2.0	1.5	1.5	1.5
35-2808	RA	2.0	2.0	1.5	1.5	1.5	1.0
35-3497	RA	2.0	1.0	1.0	1.5	1.0	1.0
35-3536	RA	1.5	1.0	1.0	1.5	1.0	1.0
35-3707	RA	2.0	1.5	1.0	1.0	1.0	1.0
35-3980	RA	2.5	1.5	1.5	1.5	1.5	1.0
CV		7.1	6.3	6.2	8.5	5.9	8.2
s		1.4	0.7	0.7	0.5	0.5	0.5
SE		0.2	0.1	0.1	0.1	0.1	0.1

†GH=genotypic high, GL=genotypic low, PH=phenotypic high, PL=phenotypic low,  
RA=random

‡1 = all plants erect, 5 = all plants prostrate

Table D8. Height by line at individual locations of Population 35 in 2009.

Line	Group	Atlantic, IA	Hedrick, IA	Washington, IA	Emery, IL	Metamora, IL	Pesotum, IL	Napoleon, OH
35-2039	GH†	104‡	86	84	81	94	91	99
35-2152	GH	112	97	89	89	102	91	102
35-2343	GH/RA	99	86	89	84	86	89	104
35-2455	GH	114	91	86	91	91	97	109
35-2592	GH/PH	102	86	81	79	81	84	99
35-2646	GH	104	81	84	81	97	81	99
35-2671	GH	97	81	76	86	97	86	102
35-2820	GH	112	81	86	84	91	84	97
35-2972	GH	109	86	86	86	99	97	102
35-3049	GH	112	89	89	86	94	94	102
35-3071	GH/RA	107	81	79	81	79	79	97
35-3190	GH	109	76	81	84	86	86	99
35-3787	GH	99	76	79	81	86	84	102
35-1983	GL	107	84	81	84	91	89	102
35-2062	GL	99	81	76	79	91	81	97
35-2184	GL	99	86	76	76	79	84	97
35-2244	GL	97	86	74	71	89	89	89
35-2450	GL	109	94	91	89	104	86	107
35-2485	GL	109	86	86	89	94	84	107
35-3253	GL	99	86	84	89	91	86	104

Table D8. Continued.

Line	Group	Atlantic, IA	Hedrick, IA	Washington, IA	Emery, IL	Metamora, IL	Pesotum, IL	Napoleon, OH
35-3388	GL	109	86	89	84	94	89	104
35-3478	GL	102	84	81	81	84	81	99
35-3731	GL	109	81	86	84	94	91	107
35-3801	GL	91	81	76	79	86	76	94
35-2235	PH	109	89	81	81	89	84	99
35-2323	PH	109	91	84	86	89	89	104
35-2684	PH	102	86	79	81	94	84	99
35-2690	PH	99	81	79	81	94	86	102
35-2701	PH	109	86	91	89	89	86	109
35-3018	PH	109	81	89	86	86	91	102
35-3544	PH	117	91	86	91	99	84	99
35-3545	PH	109	86	89	89	84	84	102
35-3927	PH	107	91	84	86	91	89	99
35-3993	PH	117	91	84	86	94	81	107
35-2213	PL	107	97	81	89	86	86	102
35-2270	PL	94	86	79	71	84	84	97
35-2328	PL	104	76	84	89	104	89	109
35-2344	PL	102	81	79	81	86	81	97
35-2366	PL	107	91	91	84	89	81	104
35-2666	PL	91	76	86	74	84	79	89

Table D8. Continued.

Line	Group	Atlantic, IA	Hedrick, IA	Washington, IA	Emery, IL	Metamora, IL	Pesotum, IL	Napoleon, OH
35-2845	PL/RA	99	81	76	81	84	81	94
35-2874	PL	109	86	79	84	81	84	99
35-2951	PL	99	81	79	79	89	86	99
35-3726	PL	94	81	79	76	86	76	94
35-3757	PL	102	76	74	81	89	76	91
35-2474	RA	104	86	99	84	94	89	99
35-2589	RA	99	94	81	79	91	94	99
35-2728	RA	104	84	79	86	86	84	102
35-2808	RA	117	81	89	89	89	86	99
35-3497	RA	109	102	89	91	99	99	109
35-3536	RA	104	91	86	81	102	84	99
35-3707	RA	109	91	89	86	94	89	104
35-3980	RA	109	86	89	94	91	86	102
CV		2	3	2	10	11	3	1
s		6	6	5	5	6	5	5
SE		1	1	1	1	1	1	1

†GH=genotypic high, GL=genotypic low, PH=phenotypic high, PL=phenotypic low, RA=random

‡Height in cm from ground to terminal node

Table D9. Seed size by line at individual locations of Population 35 in 2009.

Line	Group	Emery, IL	Metamora, IL	Pesotum, IL	Napoleon, OH
35-2039	GH†	19.6‡	17.2	-§	16.4
35-2152	GH	19.4	17.2	19.8	18.6
35-2343	GH/RA	18.3	15.4	18.6	17.6
35-2455	GH	19.7	18.1	20.0	18.1
35-2592	GH/PH	17.6	15.4	18.9	16.7
35-2646	GH	16.9	16.3	17.5	16.3
35-2671	GH	19.1	17.2	18.7	16.2
35-2820	GH	19.5	17.2	19.7	18.2
35-2972	GH	18.1	16.3	18.2	17.3
35-3049	GH	19.8	16.3	20.1	19.2
35-3071	GH/RA	19.7	15.4	17.6	16.5
35-3190	GH	21.1	17.2	21.6	16.5
35-3787	GH	20.4	17.2	20.4	18.9
35-1983	GL	-	17.2	18.5	17.8
35-2062	GL	18.1	16.3	18.1	16.2
35-2184	GL	18.9	17.2	18.4	17.0
35-2244	GL	20.5	18.1	20.3	17.5
35-2450	GL	21.0	17.2	19.5	17.9
35-2485	GL	19.3	17.2	19.5	17.2
35-3253	GL	19.6	17.2	18.6	16.7



Table D9. Continued.

Line	Group	Emery, IL	Metamora, IL	Pesotum, IL	Napoleon, OH
35-3388	GL	18.6	17.2	19.5	17.1
35-3478	GL	18.4	17.2	19.6	17.1
35-3731	GL	18.2	16.3	18.9	16.7
35-3801	GL	20.4	17.2	20.5	17.3
35-2235	PH	20.2	18.1	19.9	17.3
35-2323	PH	19.0	16.3	19.5	18.9
35-2684	PH	20.0	16.3	20.0	16.0
35-2690	PH	19.7	17.2	20.2	17.5
35-2701	PH	18.5	16.3	19.1	17.8
35-3018	PH	19.1	18.1	19.9	18.5
35-3544	PH	19.5	16.3	19.2	17.4
35-3545	PH	18.9	17.2	19.3	17.4
35-3927	PH	20.6	18.1	20.7	18.6
35-3993	PH	18.7	16.3	18.6	17.6
35-2213	PL	18.1	16.3	17.9	17.5
35-2270	PL	19.0	16.3	19.8	17.2
35-2328	PL	18.7	17.2	18.2	15.9
35-2344	PL	18.1	17.2	18.8	17.1
35-2366	PL	19.5	17.2	20.0	17.0
35-2666	PL	17.0	15.4	17.5	15.3

Table D9. Continued.

Line	Group	Emery, IL	Metamora, IL	Pesotum, IL	Napoleon, OH
35-2845	PL/RA	19.5	17.2	19.6	17.8
35-2874	PL	19.4	16.3	20.5	14.6
35-2951	PL	18.5	18.1	18.3	16.0
35-3726	PL	19.1	18.1	19.2	18.8
35-3757	PL	19.8	17.2	22.1	18.3
35-2474	RA	18.9	17.2	18.7	18.2
35-2589	RA	19.3	18.1	19.5	17.6
35-2728	RA	19.8	17.2	20.4	18.3
35-2808	RA	19.3	17.2	18.8	16.9
35-3497	RA	17.2	16.3	17.8	16.2
35-3536	RA	19.2	17.2	21.2	18.6
35-3707	RA	19.9	16.3	19.7	16.8
35-3980	RA	19.0	17.2	17.9	16.4
CV		4.0	4.0	2.7	3.8
s		0.9	0.8	1.0	1.0
SE		0.1	0.1	0.1	0.1

†GH=genotypic high, GL=genotypic low, PH=phenotypic high,  
 PL=phenotypic low, RA=random

‡Seed size expressed in g 100 seed<sup>-1</sup>

§A dash sign indicates data not reported

Table D10. Maturity by line at individual locations of Population 35 in 2009.

Line	Group	Atlantic, IA	Hedrick, IA	Washington, IA	Emery, IL	Metamora, IL	Pesotum, IL
35-2039	GH†	132‡	124	132	122	122	118
35-2152	GH	134	129	136	126	125	121
35-2343	GH/RA	134	127	133	124	124	118
35-2455	GH	135	127	134	123	125	118
35-2592	GH/PH	129	123	128	122	121	117
35-2646	GH	132	129	132	125	122	117
35-2671	GH	132	125	132	120	121	118
35-2820	GH	131	130	135	123	124	121
35-2972	GH	131	129	135	123	125	118
35-3049	GH	132	127	132	126	124	118
35-3071	GH/RA	129	123	128	124	121	112
35-3190	GH	130	129	129	121	120	118
35-3787	GH	129	124	128	121	121	117
35-1983	GL	131	126	132	120	124	121
35-2062	GL	130	127	129	127	122	118
35-2184	GL	130	127	132	122	121	116
35-2244	GL	129	123	129	123	122	117
35-2450	GL	128	123	132	120	120	120
35-2485	GL	134	124	133	121	124	117
35-3253	GL	130	127	132	120	124	117

Table D10. Continued.

Line	Group	Atlantic, IA	Hedrick, IA	Washington, IA	Emery, IL	Metamora, IL	Pesotum, IL
35-3388	GL	136	128	135	120	125	121
35-3478	GL	130	124	129	120	122	120
35-3731	GL	134	124	131	121	122	114
35-3801	GL	131	124	129	123	120	121
35-2235	PH	134	125	135	126	126	118
35-2323	PH	132	126	132	121	125	120
35-2684	PH	136	130	139	126	126	121
35-2690	PH	134	128	135	125	125	121
35-2701	PH	136	130	140	128	127	124
35-3018	PH	135	128	136	125	126	119
35-3544	PH	132	127	131	123	124	120
35-3545	PH	136	129	139	121	127	118
35-3927	PH	134	129	132	121	124	122
35-3993	PH	130	127	132	124	122	117
35-2213	PL	136	130	140	126	126	118
35-2270	PL	128	122	127	122	117	116
35-2328	PL	134	130	135	121	127	122
35-2344	PL	131	127	133	122	122	118
35-2366	PL	134	130	136	125	125	121
35-2666	PL	127	123	127	120	116	111

Table D10. Continued.

Line	Group	Atlantic, IA	Hedrick, IA	Washington, IA	Emery, IL	Metamora, IL	Pesotum, IL
35-2845	PL/RA	128	122	128	121	119	112
35-2874	PL	129	124	128	120	121	117
35-2951	PL	128	123	127	121	119	116
35-3726	PL	129	124	131	121	122	120
35-3757	PL	129	122	129	121	119	117
35-2474	RA	131	127	132	120	122	117
35-2589	RA	134	128	134	125	125	120
35-2728	RA	132	127	132	127	125	120
35-2808	RA	135	129	136	125	125	120
35-3497	RA	130	124	132	124	121	118
35-3536	RA	134	125	133	120	121	117
35-3707	RA	134	129	135	126	124	118
35-3980	RA	134	130	138	123	126	122
CV		1	0	1	1	1	2
s		3	3	3	2	3	3
SE		0	0	0	0	0	0

†GH=genotypic high, GL=genotypic low, PH=phenotypic high, PL=phenotypic low,  
RA=random

‡Days after planting

**APPENDIX E**

**MEAN GERMINATION OF EACH LINE**

Table E1. Mean germination of each selected line of Population 34.

Line	Group	Germination (%)
34-0414	GH†	91
34-0692	GH	89
34-0777	GH	91
34-1046	GH	77
34-1055	GH	91
34-1058	GH	88
34-1553	GH	90
34-1627	GH	91
34-1730	GH	88
34-1757	GH	90
34-0724	GH/PH	84
34-0351	GL	93
34-0450	GL	83
34-0485	GL	82
34-0503	GL	88
34-0760	GL	90
34-0873	GL	88
34-1242	GL	87
34-1288	GL	82
34-1716	GL	81
34-1806	GL	85
34-1897	GL	89
34-0506	PH	87
34-0518	PH	91
34-0626	PH	90
34-0662	PH	88
34-0754	PH	91
34-0947	PH	75
34-1118	PH	86
34-1157	PH	90
34-1340	PH	85
34-1440	PH	90
34-0297	PL	82

Table E1. Continued.

34-0378	PL	84
34-0405	PL	84
34-0538	PL	90
34-0778	PL	90
34-1256	PL	89
34-1426	PL	90
34-1486	PL	88
34-1609	PL	86
34-1654	PL	87
34-1808	PL	92
34-0467	RA	92
34-0679	RA	88
34-0761	RA	82
34-1029	RA	95
34-1032	RA	88
34-1140	RA	89
34-1333	RA	87
34-1625	RA	91
34-1666	RA	88
34-1779	RA	92
34-1847	RA	90
CV		5
S		5
SE		1

†GH=genotypic high, GL=genotypic low,  
 PH=phenotypic high, PL=phenotypic low,  
 RA=random



Table E2. Mean germination of each selected line of Population 35.

Line	Group	Germination (%)
35-2039	GH†	83
35-2152	GH	81
35-2455	GH	85
35-2646	GH	88
35-2671	GH	85
35-2820	GH	83
35-2972	GH	82
35-3049	GH	86
35-3190	GH	77
35-3787	GH	81
35-2592	GH/PH	85
35-2343	GH/RA	83
35-3071	GH/RA	82
35-1983	GL	83
35-2062	GL	82
35-2184	GL	86
35-2244	GL	74
35-2450	GL	89
35-2485	GL	88
35-3253	GL	85
35-3388	GL	84
35-3478	GL	81
35-3731	GL	84
35-3801	GL	74
35-2235	PH	83
35-2323	PH	83
35-2684	PH	83
35-2690	PH	88
35-2701	PH	88
35-3018	PH	84
35-3544	PH	83
35-3545	PH	86
35-3927	PH	83

Table E2. Continued.

35-3993	PH	80
35-2213	PL	75
35-2270	PL	79
35-2328	PL	89
35-2344	PL	78
35-2366	PL	89
35-2666	PL	80
35-2874	PL	82
35-2951	PL	74
35-3726	PL	71
35-3757	PL	77
35-2845	PL/RA	79
35-2474	RA	71
35-2589	RA	86
35-2728	RA	69
35-2808	RA	85
35-3497	RA	82
35-3536	RA	85
35-3707	RA	85
35-3980	RA	85
CV		7
s		7
SE		1

†GH=genotypic high, GL=genotypic low, PH=phenotypic high, PL=phenotypic low, RA=random