

Protein Characterizations by Chiral Vibrational Sum Frequency Generation Spectroscopy

International Symposium on Molecular Spectroscopy

74th Meeting

June 17, 2019

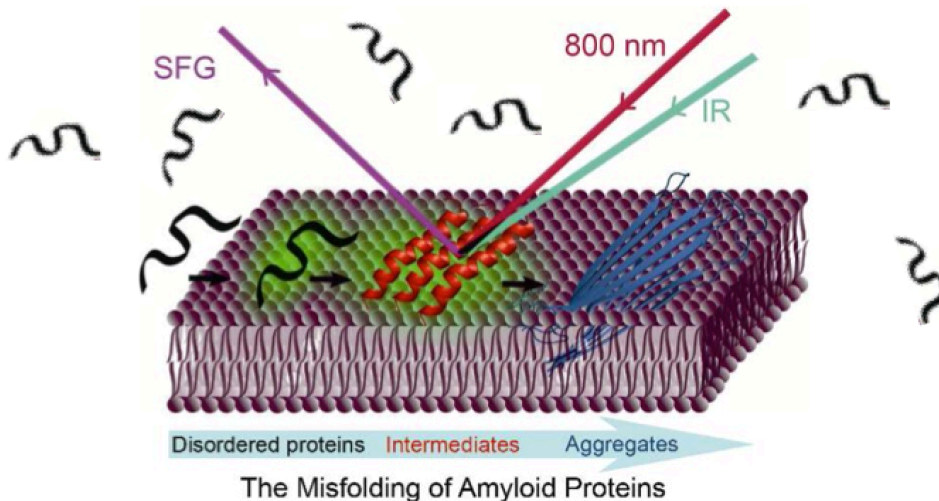
Champaign-Urbana, IL



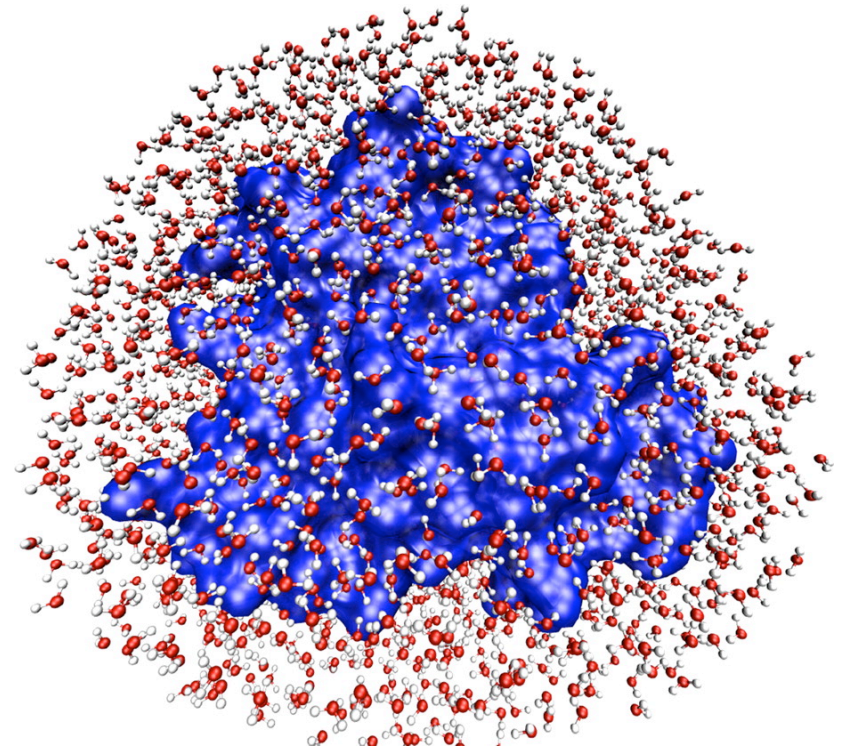
Elsa Chui-Ying Yan
Department of Chemistry
Yale University

Protein Characterizations by Chiral Vibrational Sum Frequency Generation Spectroscopy

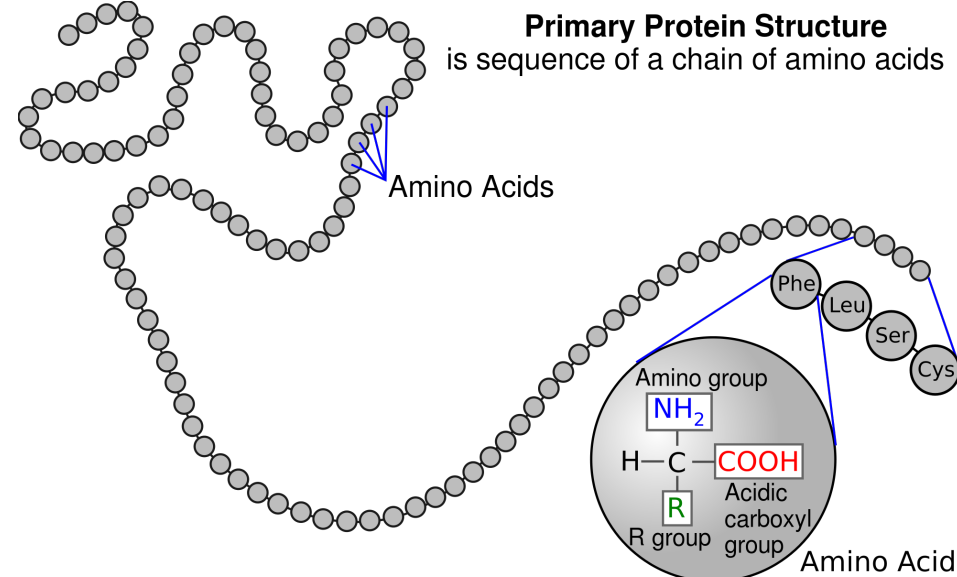
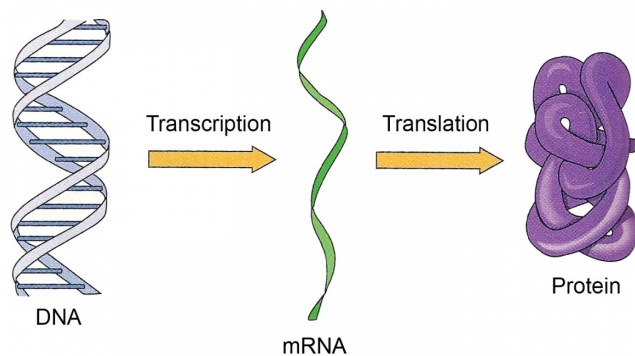
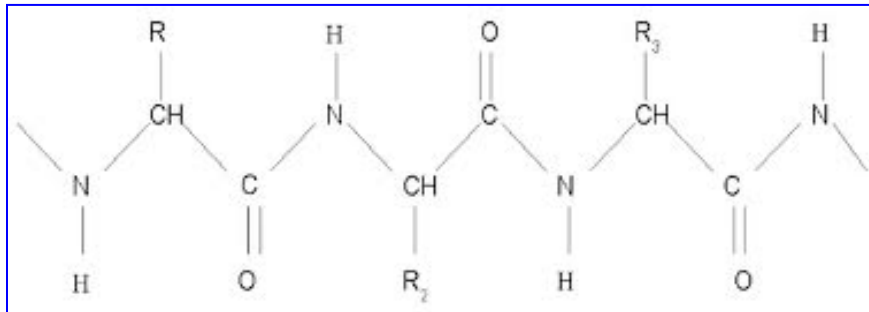
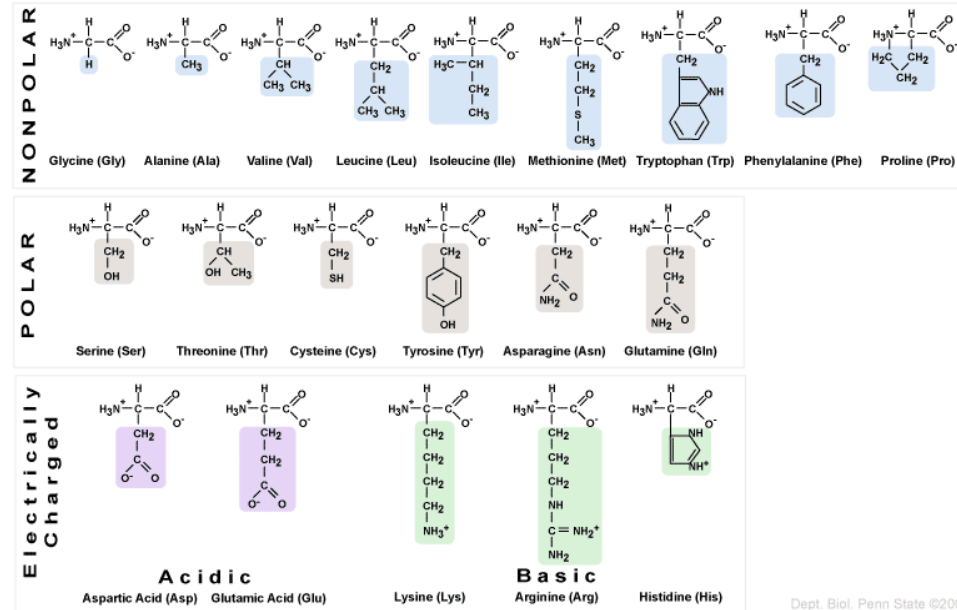
I. Characterizing Protein Secondary Structures at Interfaces



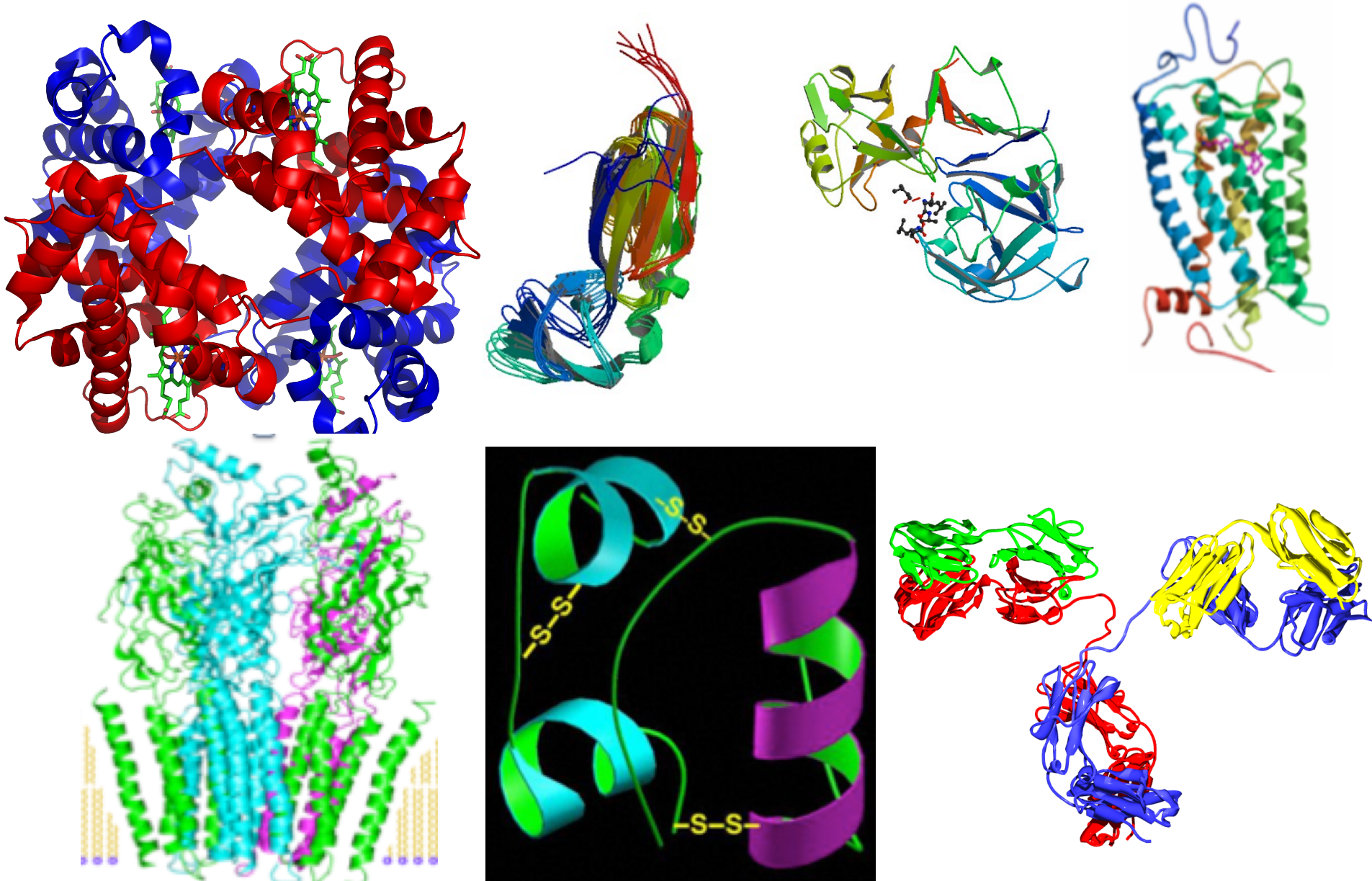
II. Probing Water Structures in Hydration Shell of Proteins



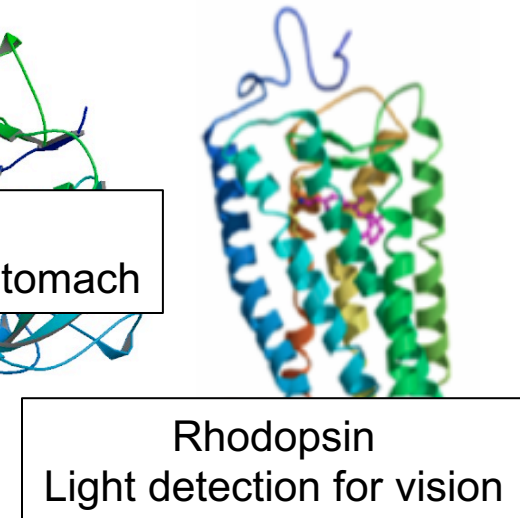
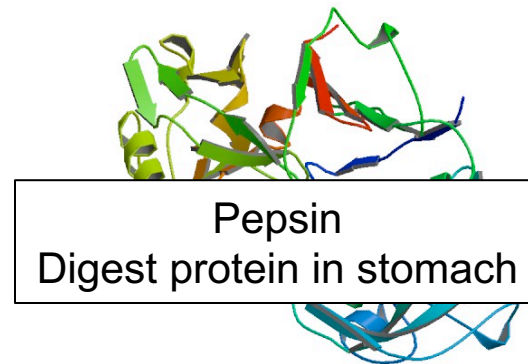
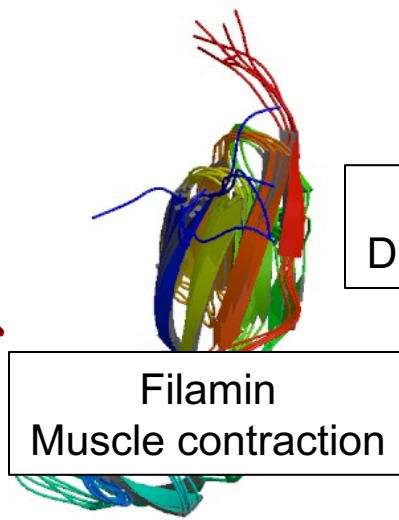
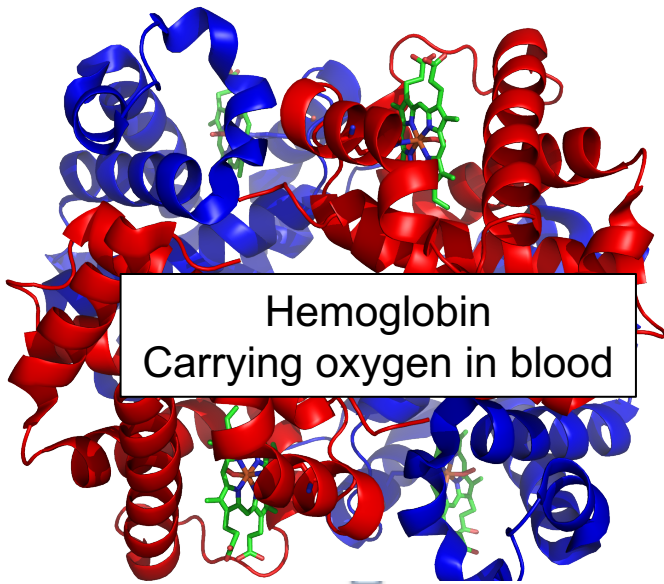
Motivation: Proteins



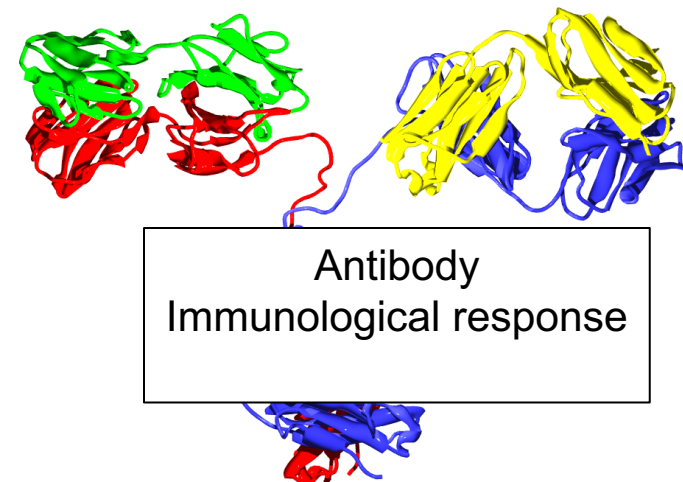
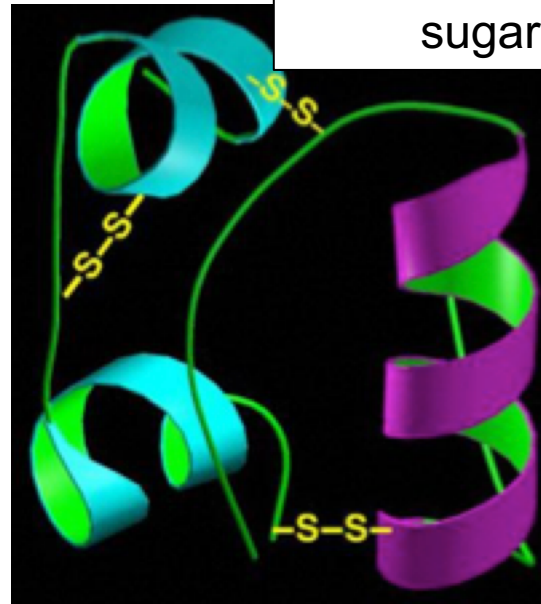
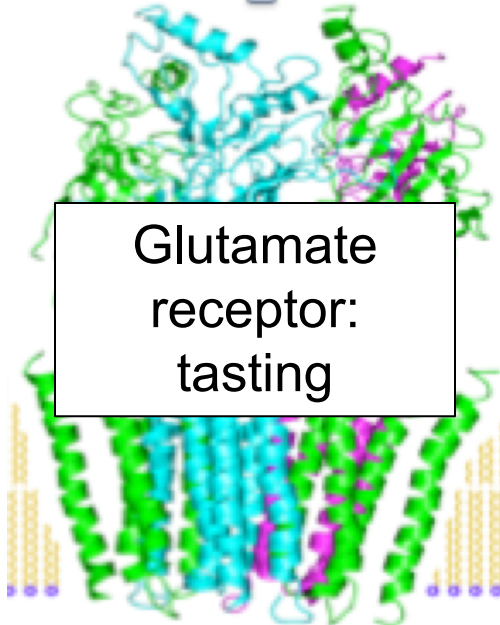
Motivation: Protein Structures and Functions



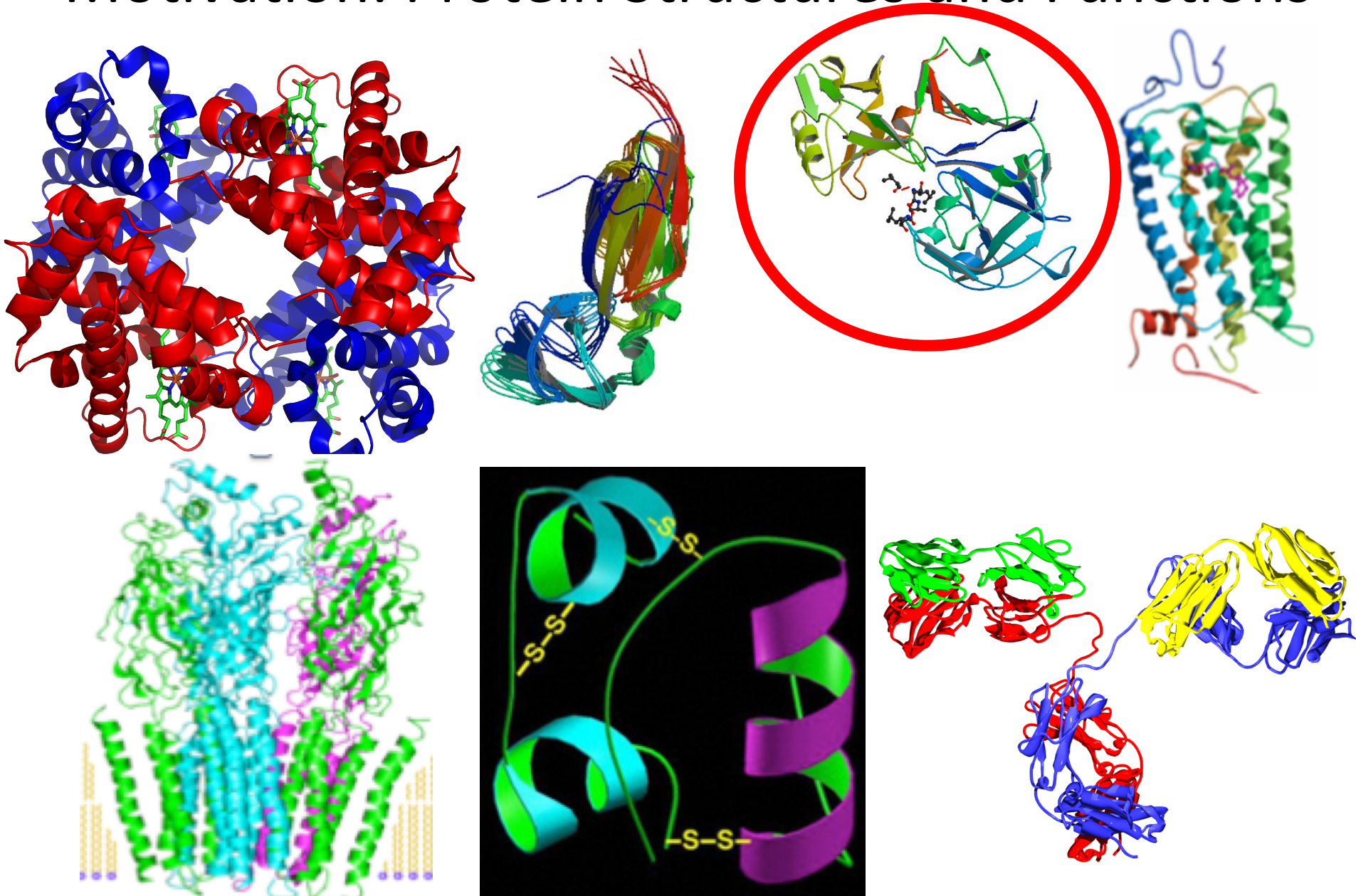
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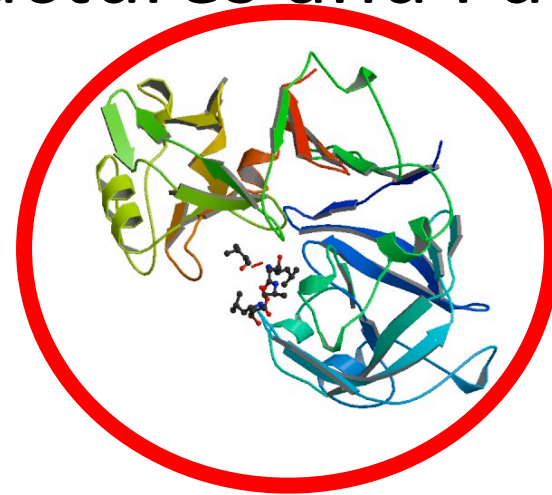
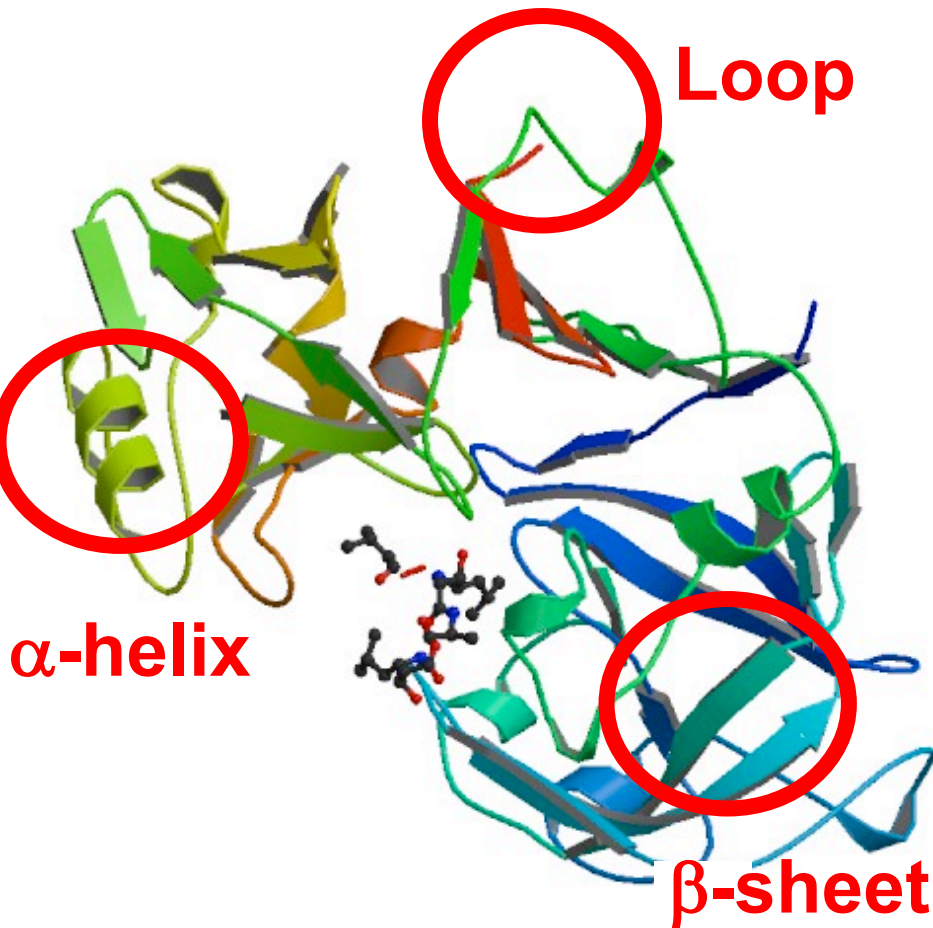
Insulin: hormone for controlling
sugar level in blood



Motivation: Protein Structures and Functions



Motivation: Protein Structures and Functions



Protein Secondary structures:

1. Stability
 2. Dynamics
 3. Organization
- ⇒ Mechanism of protein functions
- To explore disease mechanism
 - To develop therapeutics

Conventional Methods:

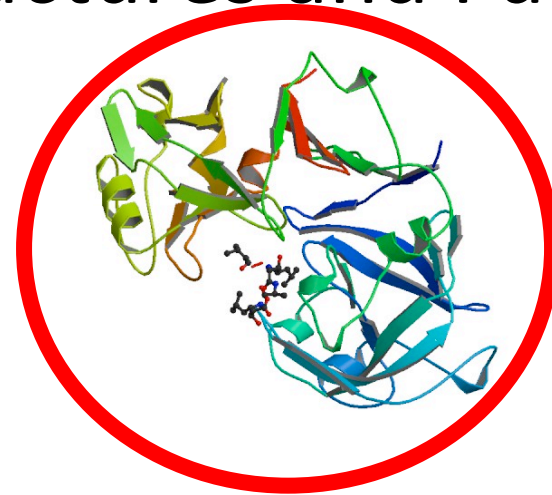
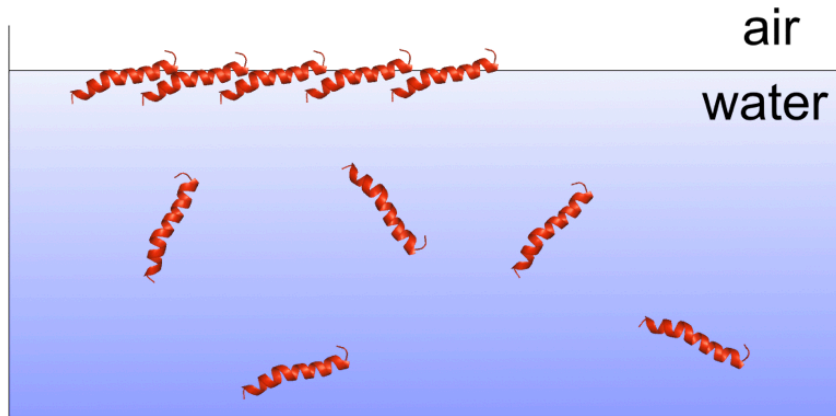
1. Circular Dichroism Spectroscopy
2. NMR Spectroscopy
3. X-ray Crystallography

} ⇒ Not surface sensitive

Motivation: Protein Structures and Functions

Signals overwhelmed by

- Solvent
- proteins in the bulk



Protein Secondary structures:

1. Stability
2. Dynamics
3. Organization

⇒ Mechanism of protein functions

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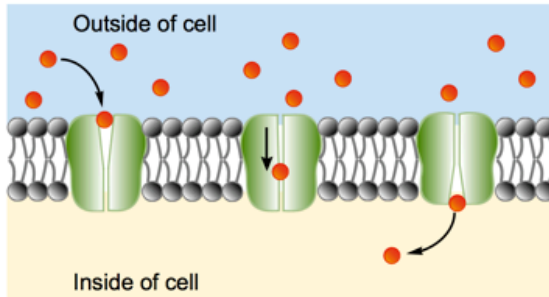
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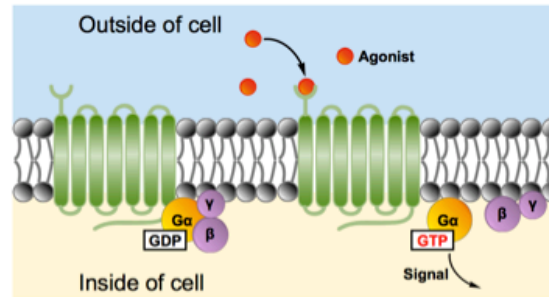
} => Not surface sensitive

Motivation: Proteins at Interfaces

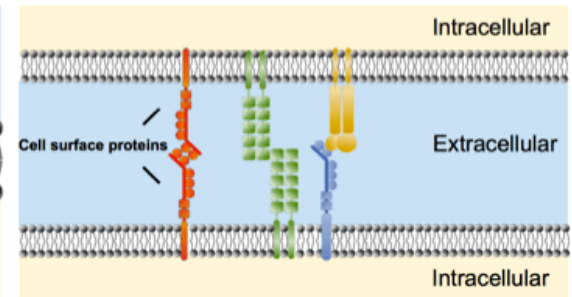
Molecular Transport



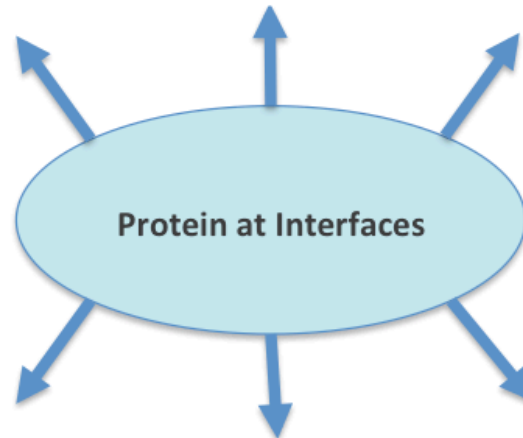
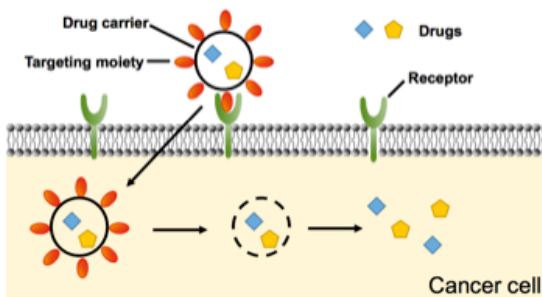
Signal Transduction



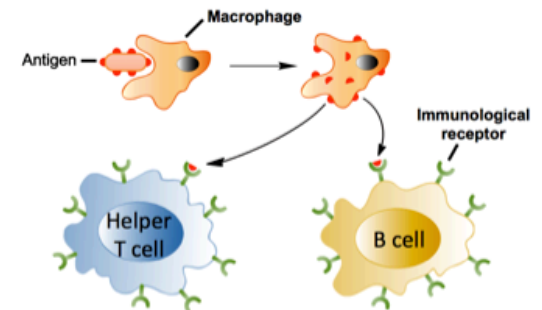
Cell Recognition and Adhesion



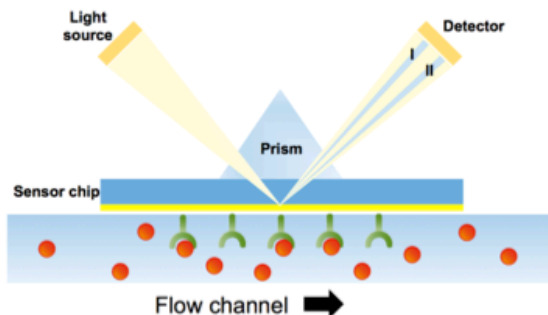
Drug Delivery



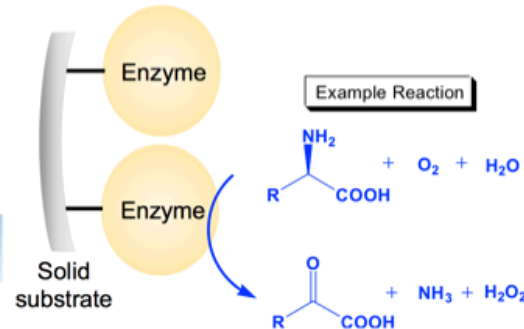
Immunological Response



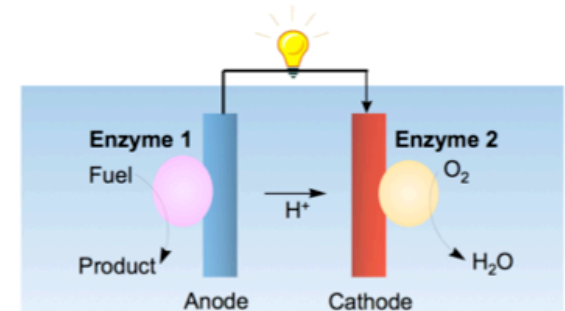
Biosensor



Heterogeneous Biocatalyst



Biofuel Cell



Motivation: Protein Structures at Interfaces

Interfaces and Bulk Media are **DIFFERENT!**

Air

Density 1.2 g/L

Refractive index: 1

Dielectric constant: 1

Water

Density 1000 g/L

Refractive index: 1.3

Dielectric constant: 80

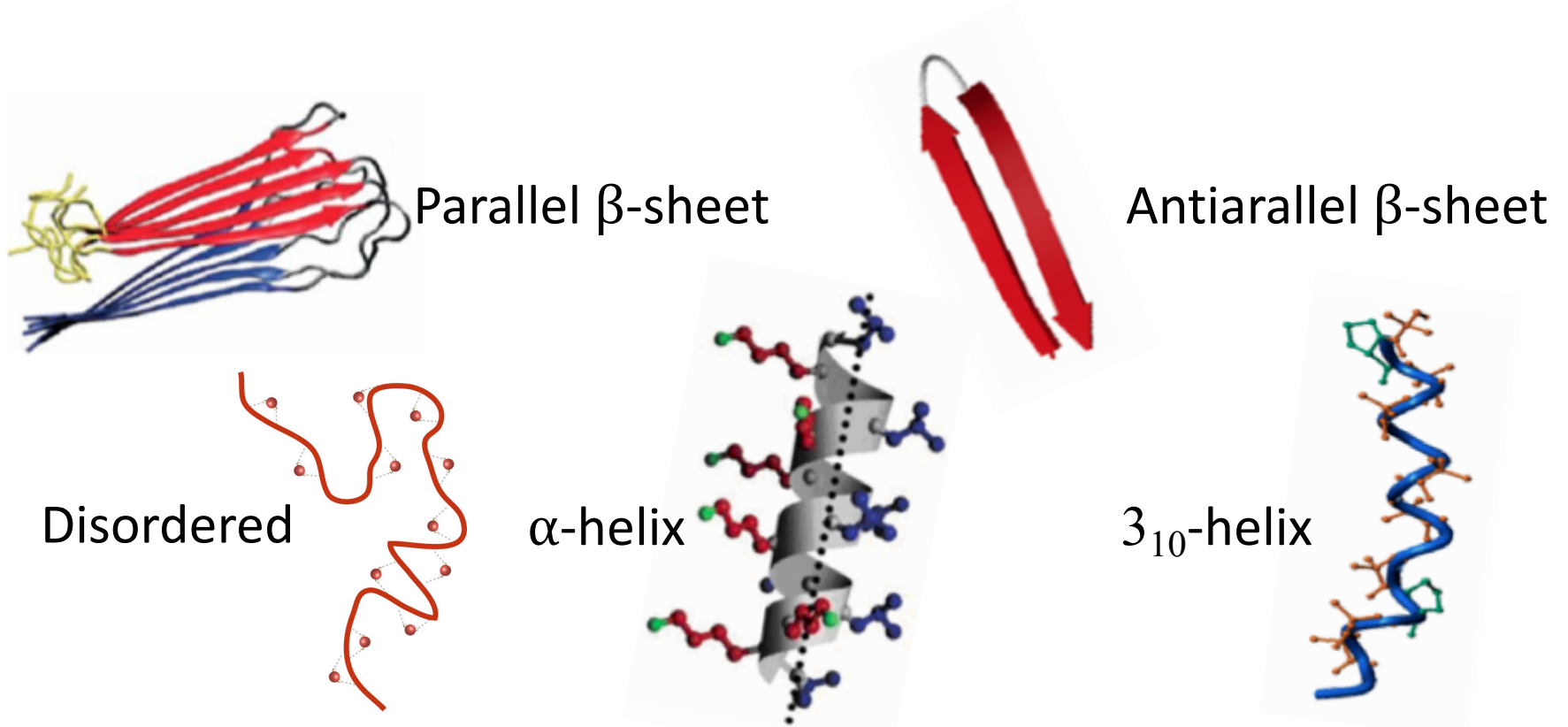
? ? ?
? Interface ?
? ? ?



Need:

Characterization of Protein in situ at Interfaces

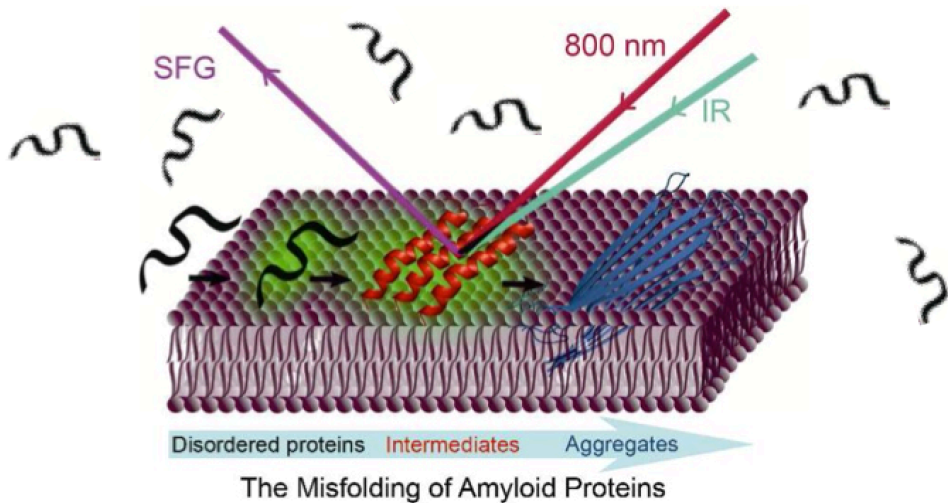
Motivation: Protein Secondary Structures— Basic Functional Units of Proteins



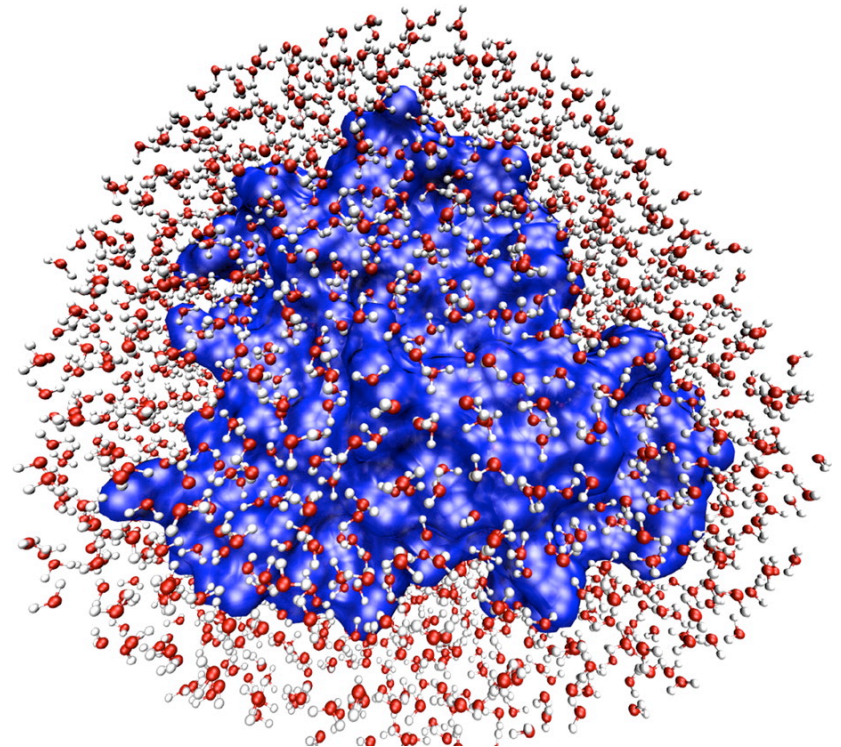
In situ characterization of protein secondary structures at interfaces is important to fundamental biological sciences and engineering.

Protein Characterizations by Chiral Vibrational Sum Frequency Generation Spectroscopy

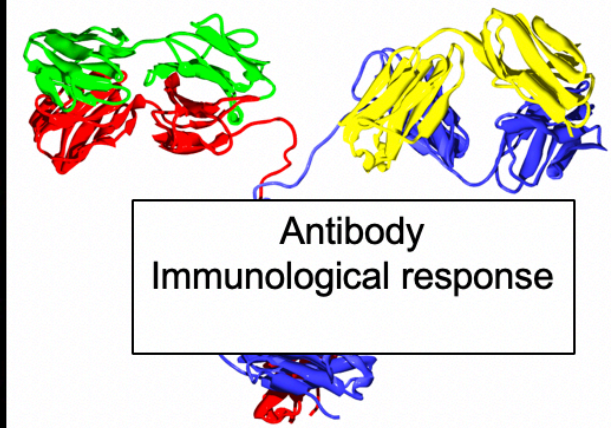
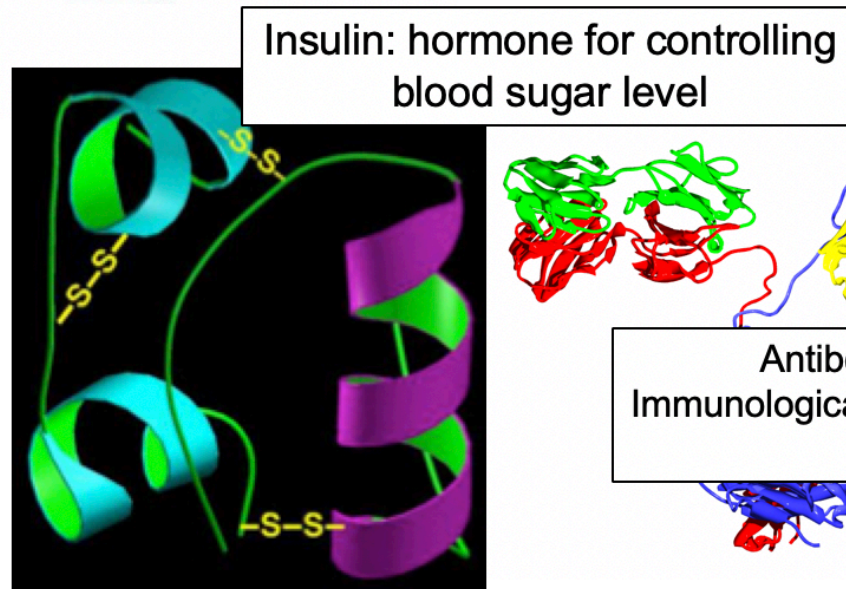
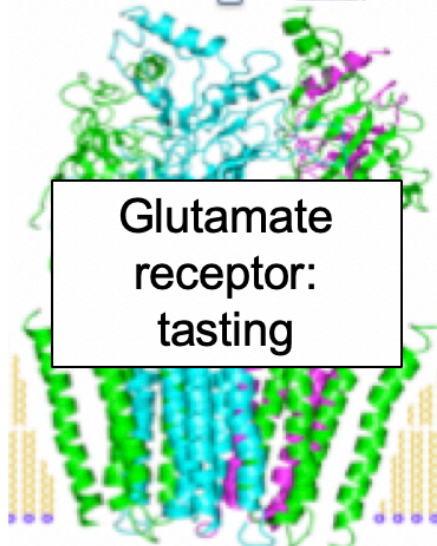
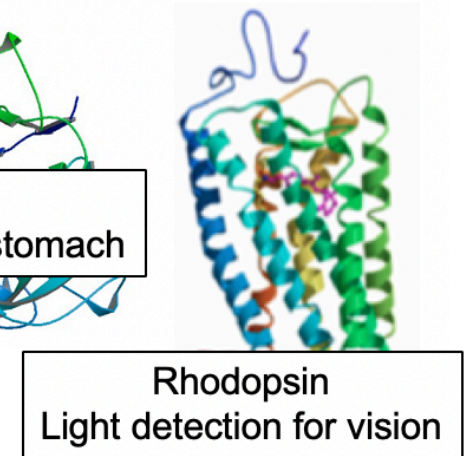
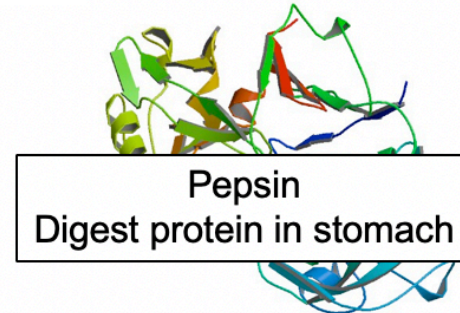
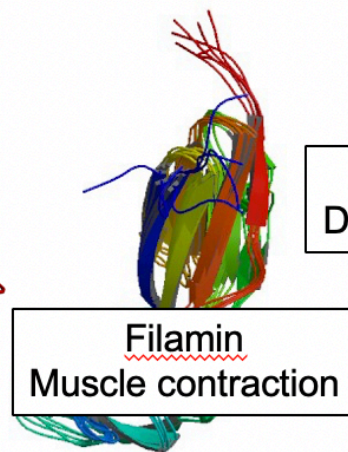
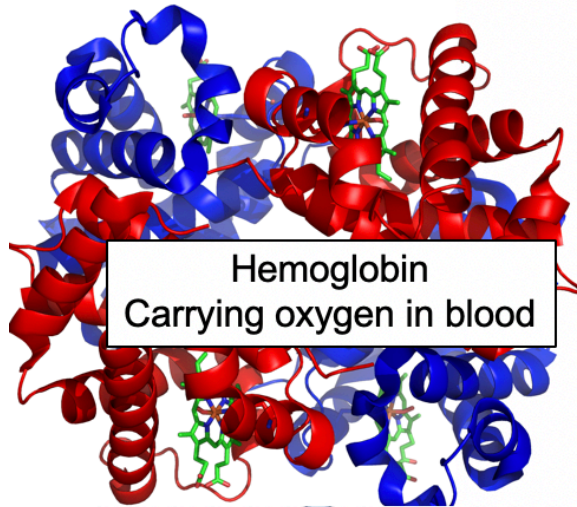
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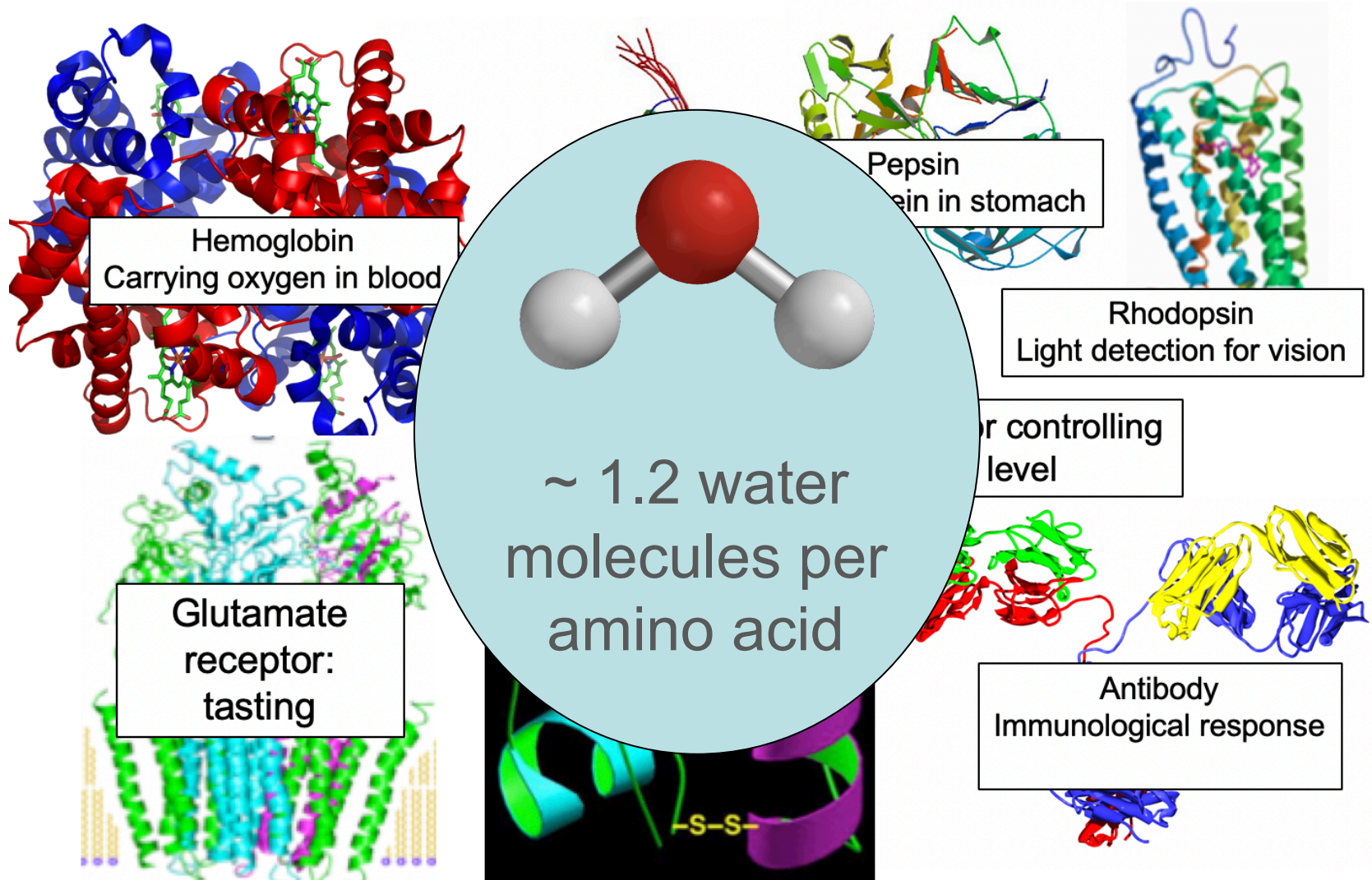
II. Probing Water Structures in Hydration Shell of Proteins



Motivation: Water Structures in Protein Hydration



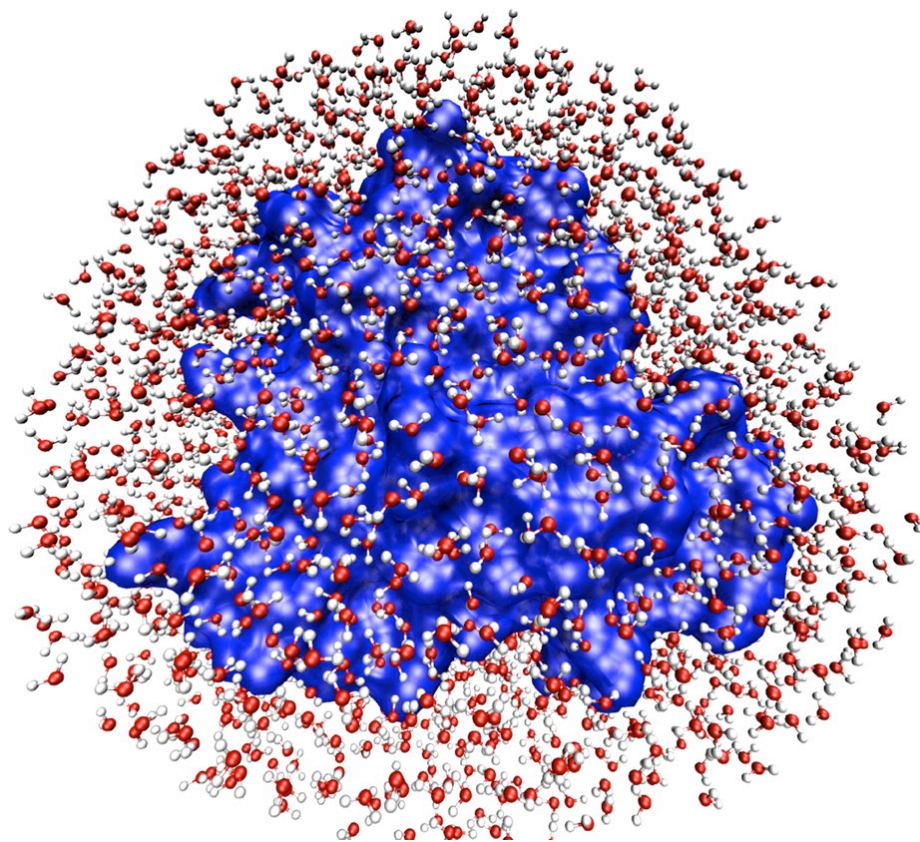
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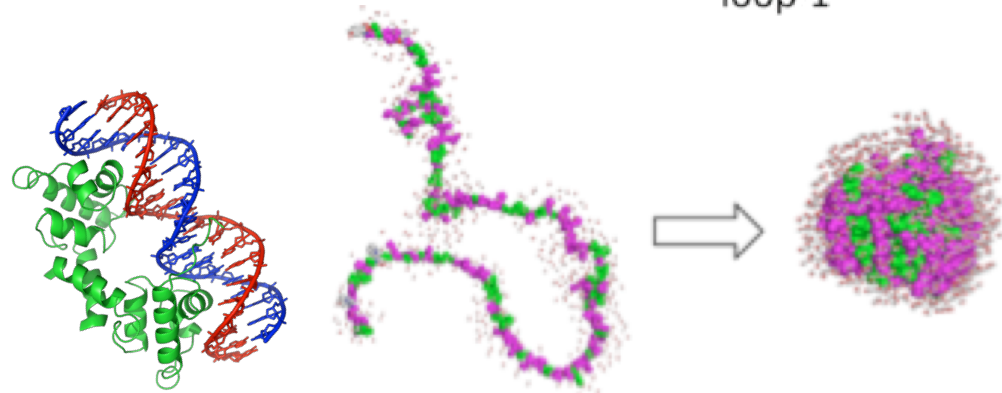
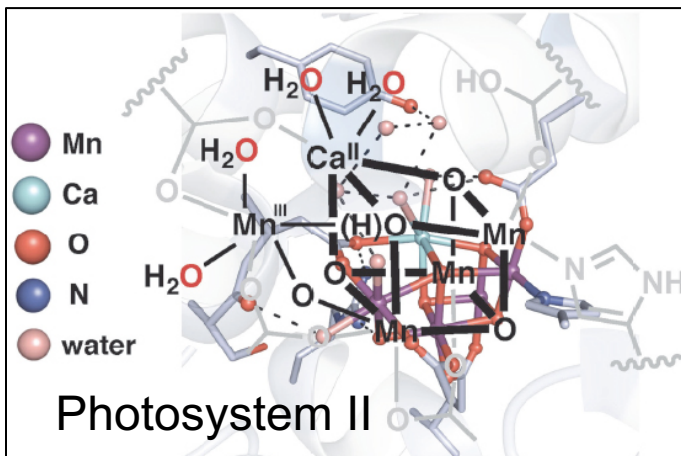
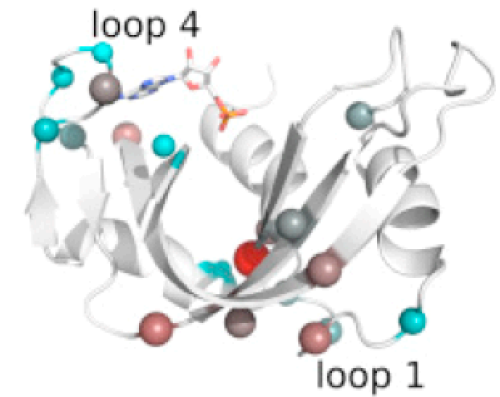
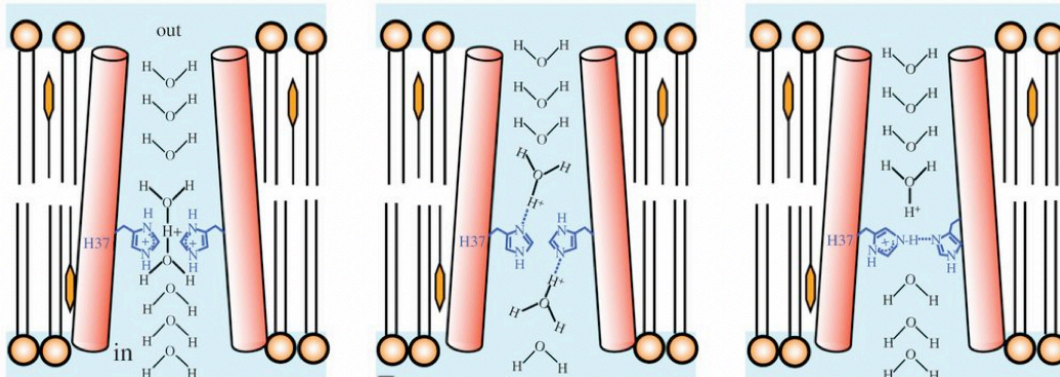
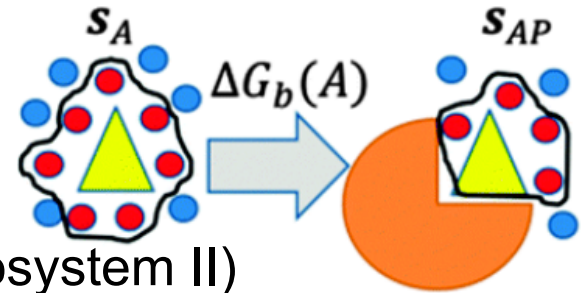
1. Protein dynamics
2. Protein folding/aggregation
3. Protein Interactions
4. Participation in biochemical reactions
5. Translocation ions across membrane

Water molecules are integral parts of proteins.



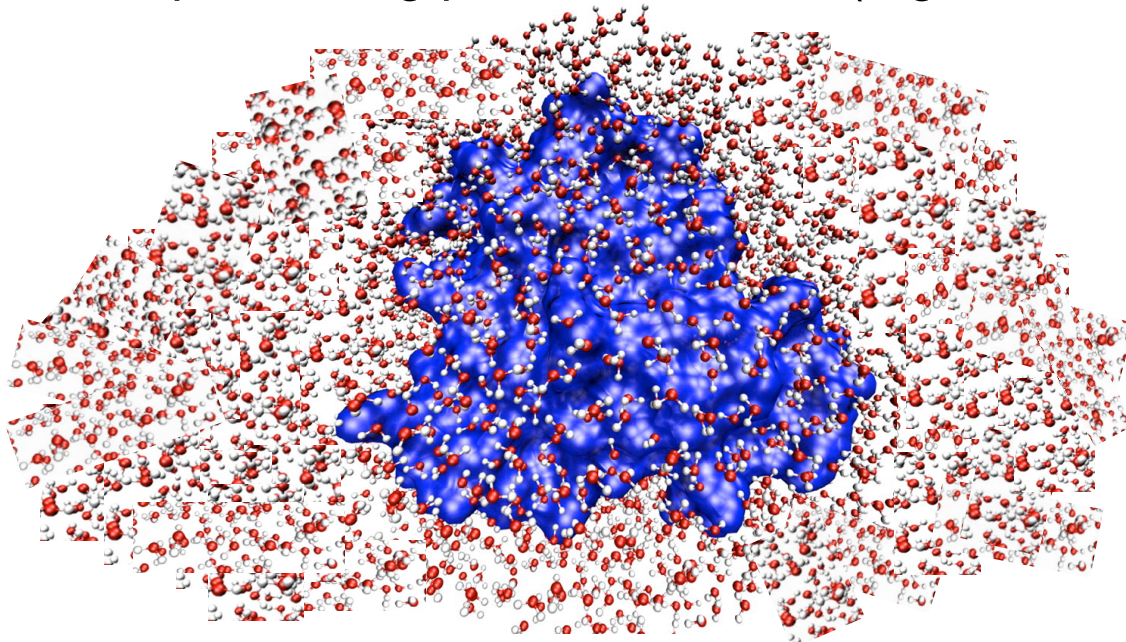
Motivation: Water Structures in Protein Hydration

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3. Protein Interactions
4. Participation in biochemical reactions (e.g., Photosystem II)
5. Involve in performing protein functions (e.g., ions transport)



Motivation: Water Structures in Protein Hydration

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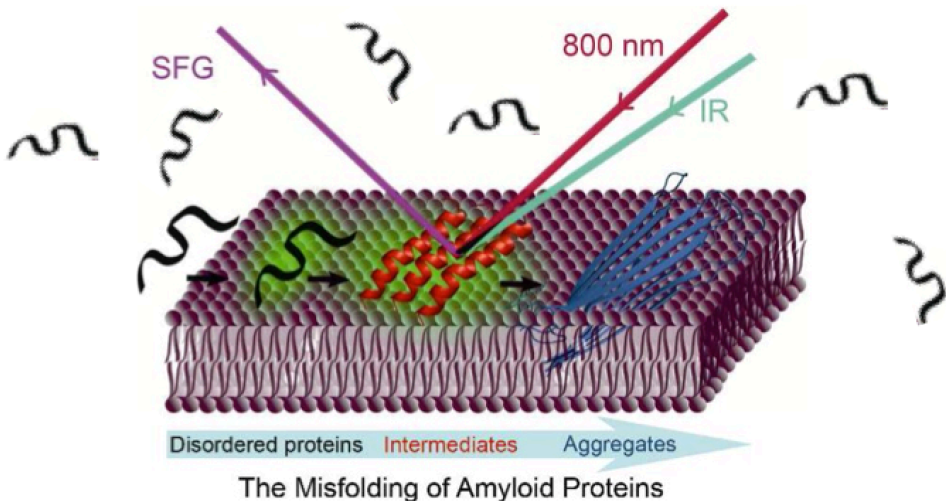


=> Structure-function
correlation of proteins

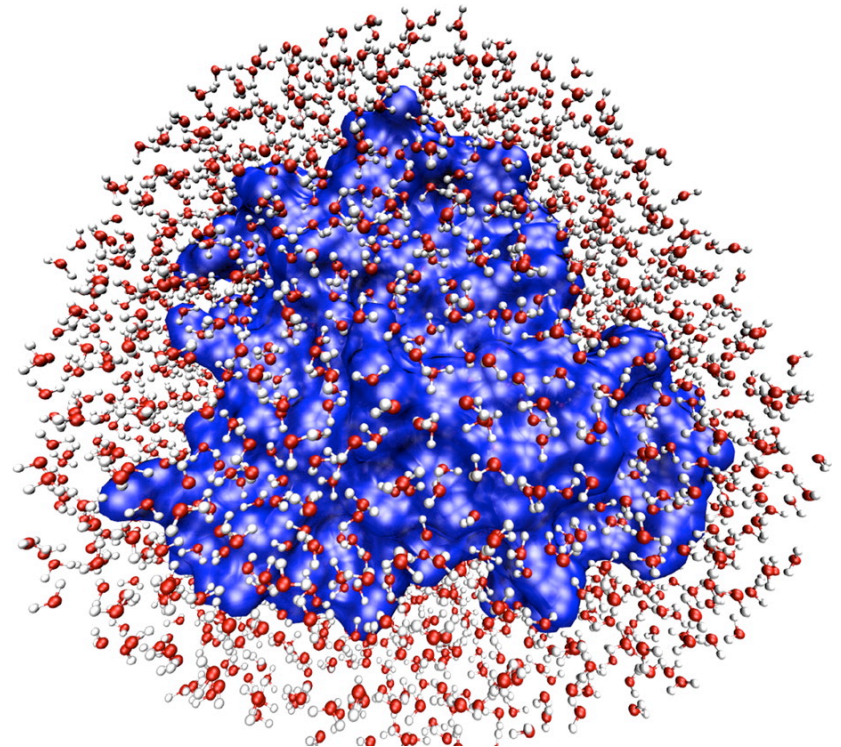
Challenges:
**Background from a large population of water molecules in
bulk solution**

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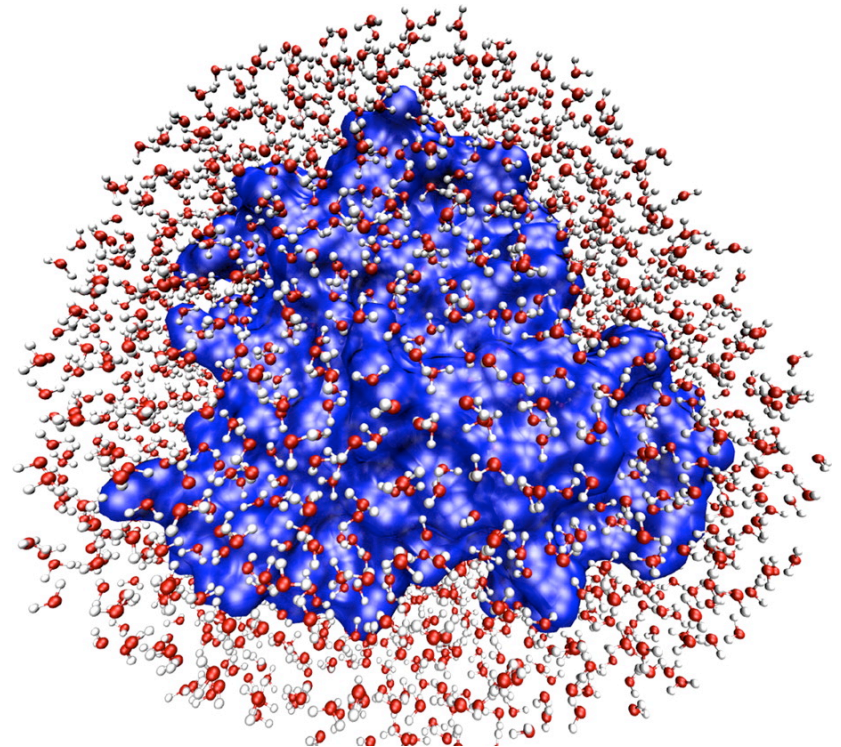
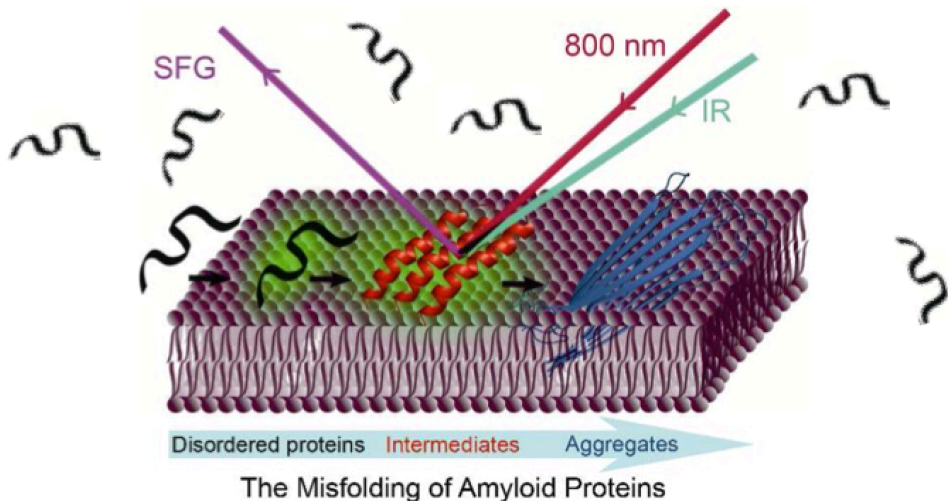


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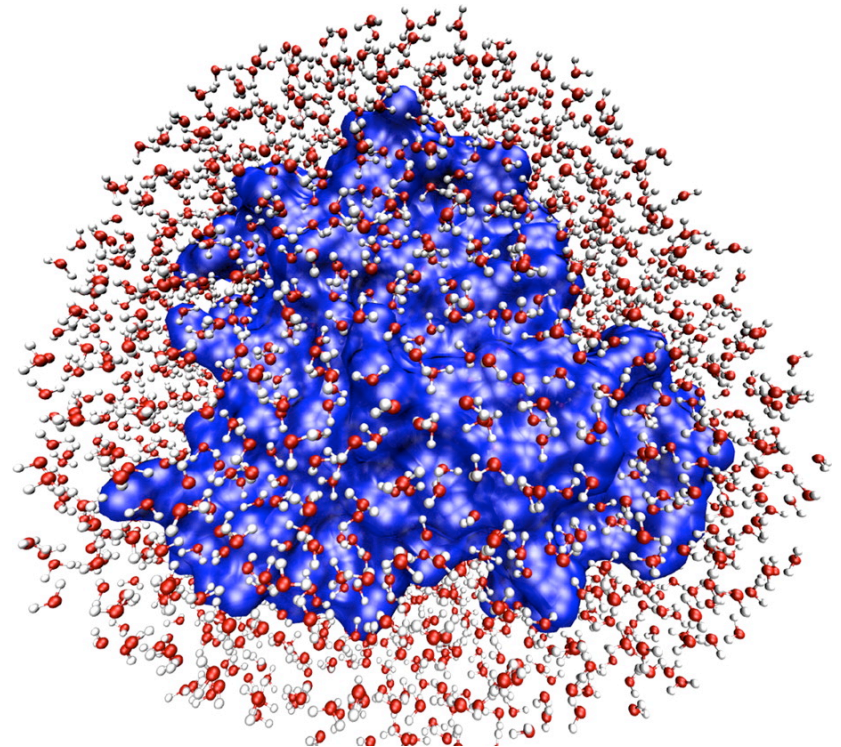
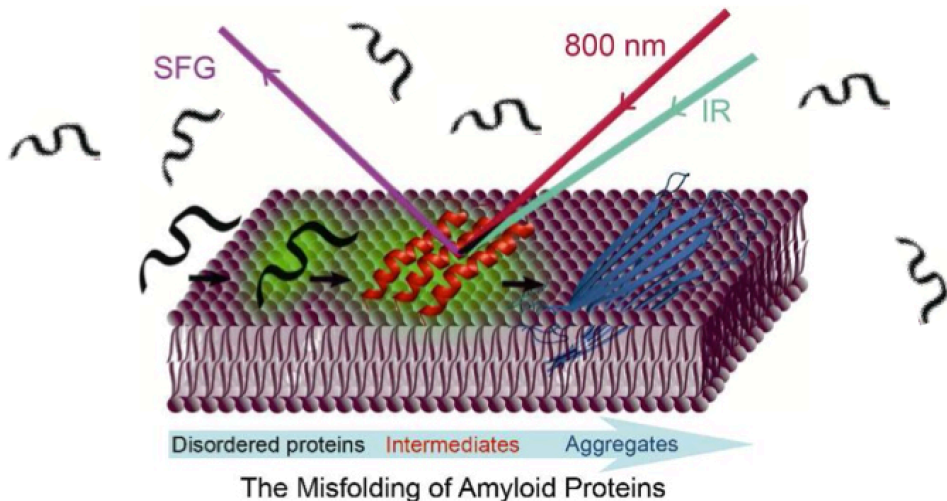
Outline

1. Introduction to Chiral SFG Spectroscopy
2. Characterizing Protein Secondary Structures at Interfaces
3. Probing Water Structures in Hydration Shell of Proteins



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Outline

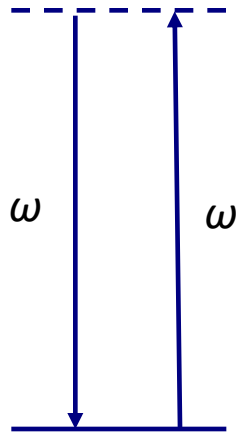
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Development of Chiral SFG

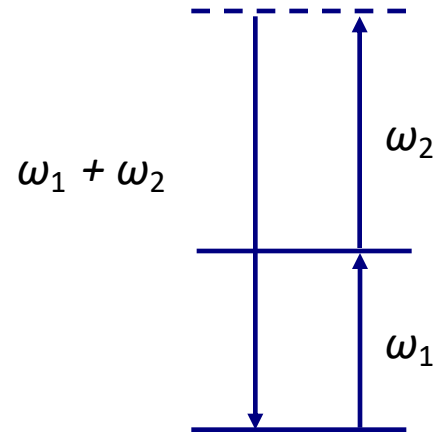
1. Shen (Berkeley): Observation of chiral SFG from pure chiral bulk liquid (Belkin et al, *Phys. Rev. Lett.*, 2000)
2. Simpson (Purdue): Theory of surface-sensitive chiral SFG for probing macroscopic chiral structures (Simpson, *ChemPhysChem*, 2004)
3. Chen (Michigan): First application to study amide I of a β -sheet on polymer surface (Wang et al., *PNAS*, 2005)
4. Geiger (Northwestern): First application to study double helix DNA on glass surface (Stoke et al., *JACS*, 2007,)

Introduction to Chiral SFG Spectroscopy

SFG Spectroscopy



Linear optical process

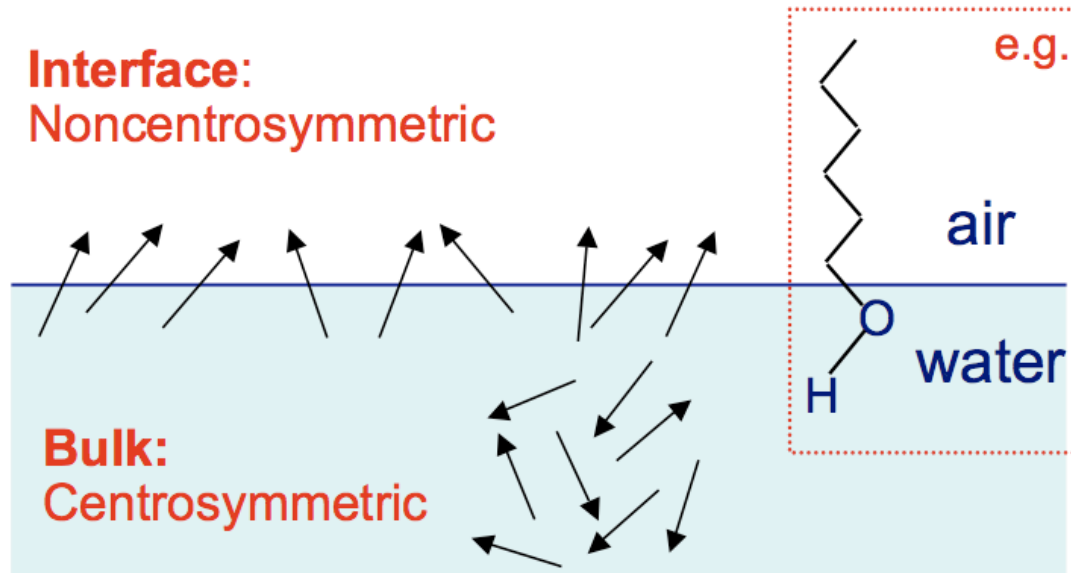


Second-order optical process

A second-order optical technique

Introduction to Chiral SFG Spectroscopy

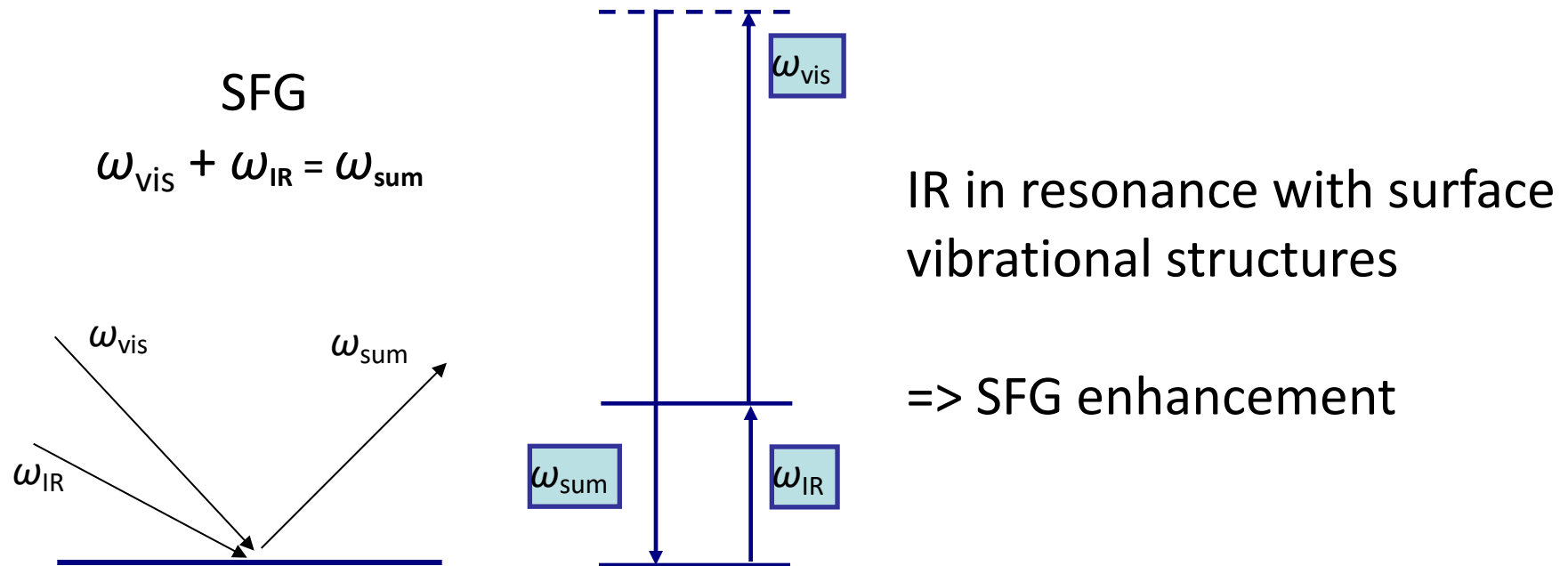
SFG Spectroscopy



Noncentrosymmetric Interface => Surface-selectivity

Introduction to Chiral SFG Spectroscopy

SFG Spectroscopy



SFG: a surface-specific vibrational spectroscopy

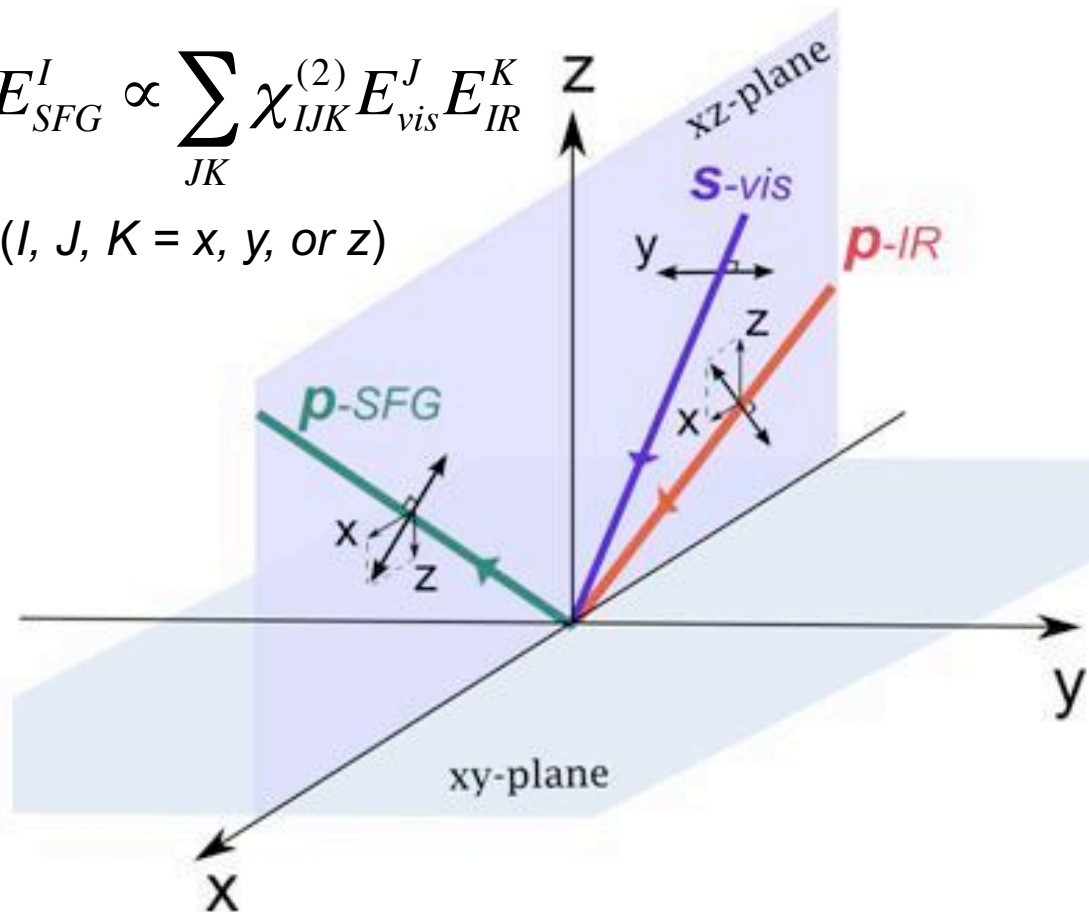
Introduction to Chiral SFG Spectroscopy

$$I_{SFG} = \left| \vec{E}_{SFG} \right|^2$$

$$\vec{E}_{SFG} = E_{SFG}^x \hat{x} + E_{SFG}^y \hat{y} + E_{SFG}^z \hat{z}$$

$$E_{SFG}^I \propto \sum_{JK} \chi_{IJK}^{(2)} E_{vis}^J E_{IR}^K$$

($I, J, K = x, y, \text{ or } z$)



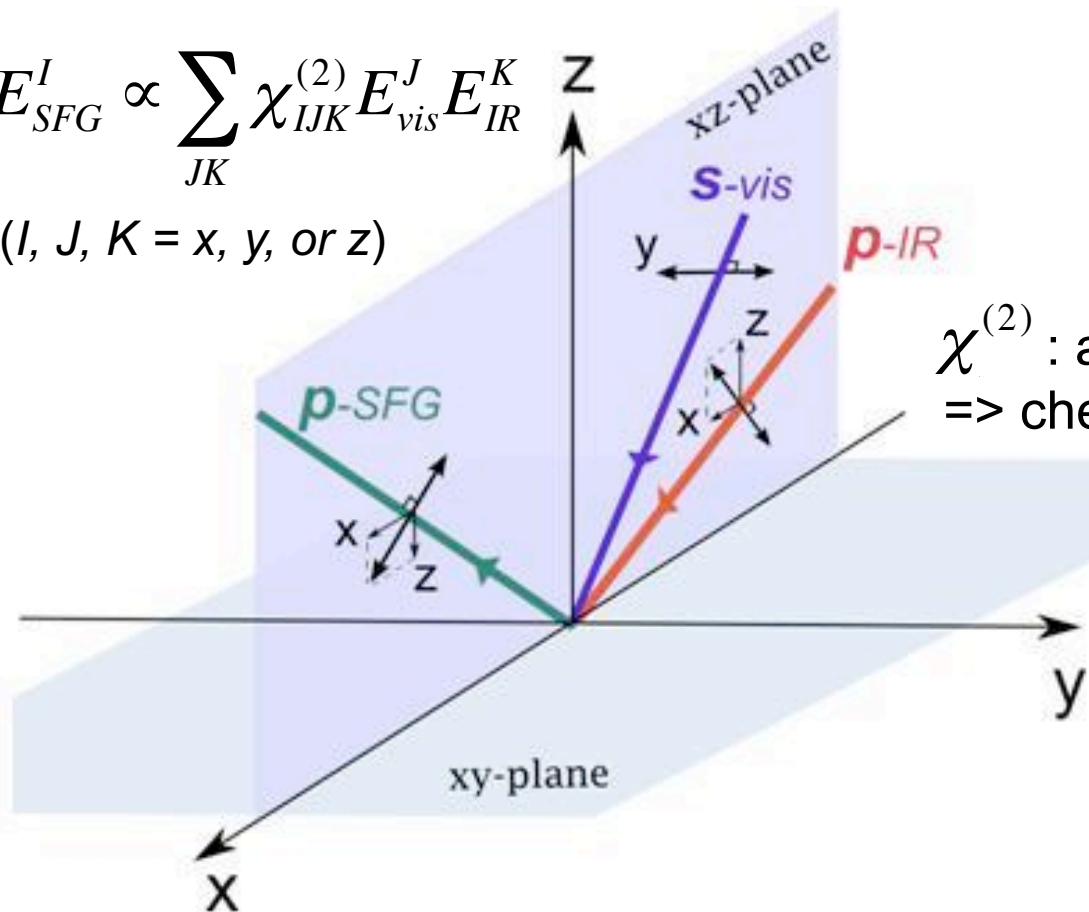
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$\chi^{(2)}$: a tensor with 27 elements
 \Rightarrow chemical & structural information

Individual tensor elements
 can be measured by
 different combinations of
 polarization settings.

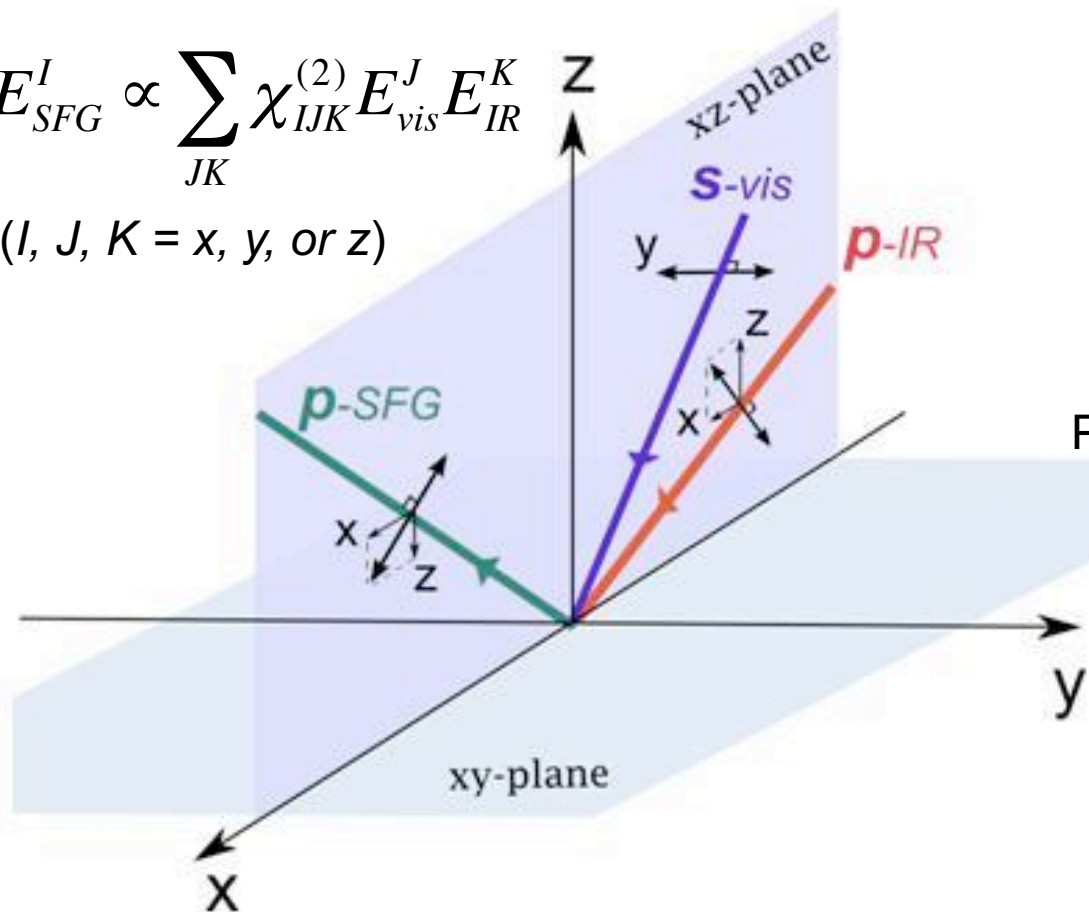
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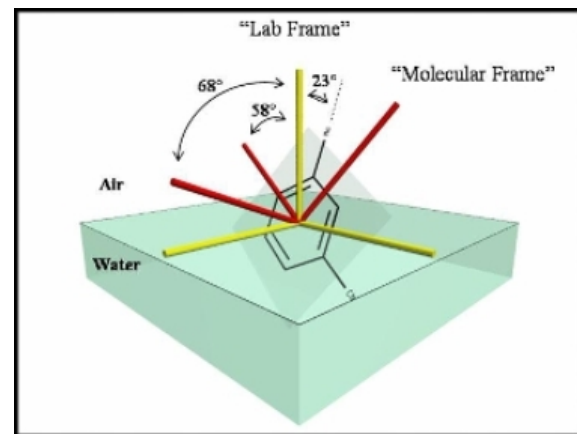
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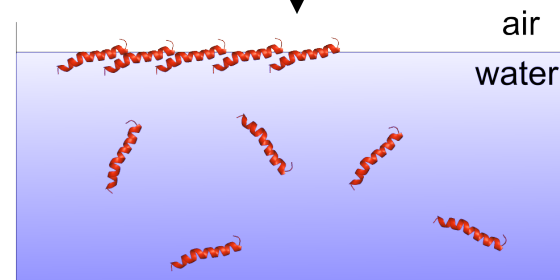


Molecular Orientation



Polarization of beams

$\chi_{IJK}^{(2)}$



Surface chirality

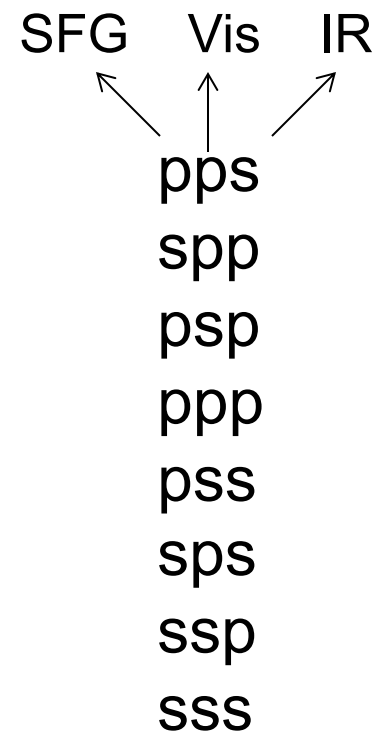
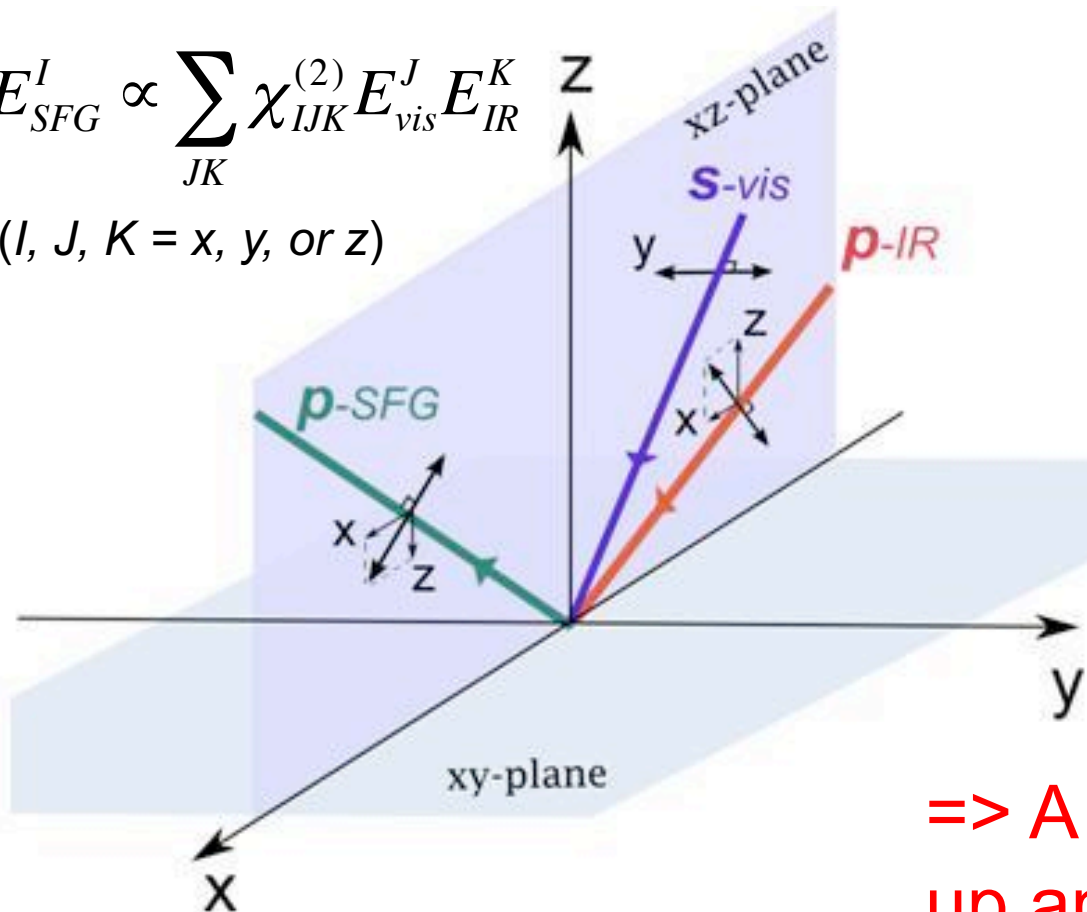
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=> A total of 8 ways to set up an SFG experiment

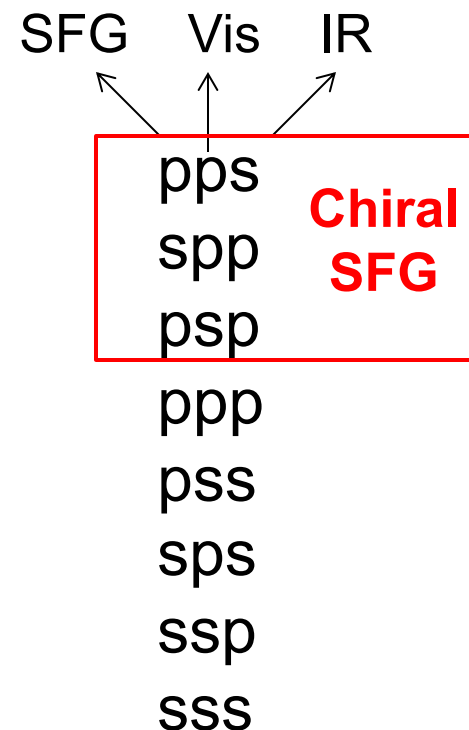
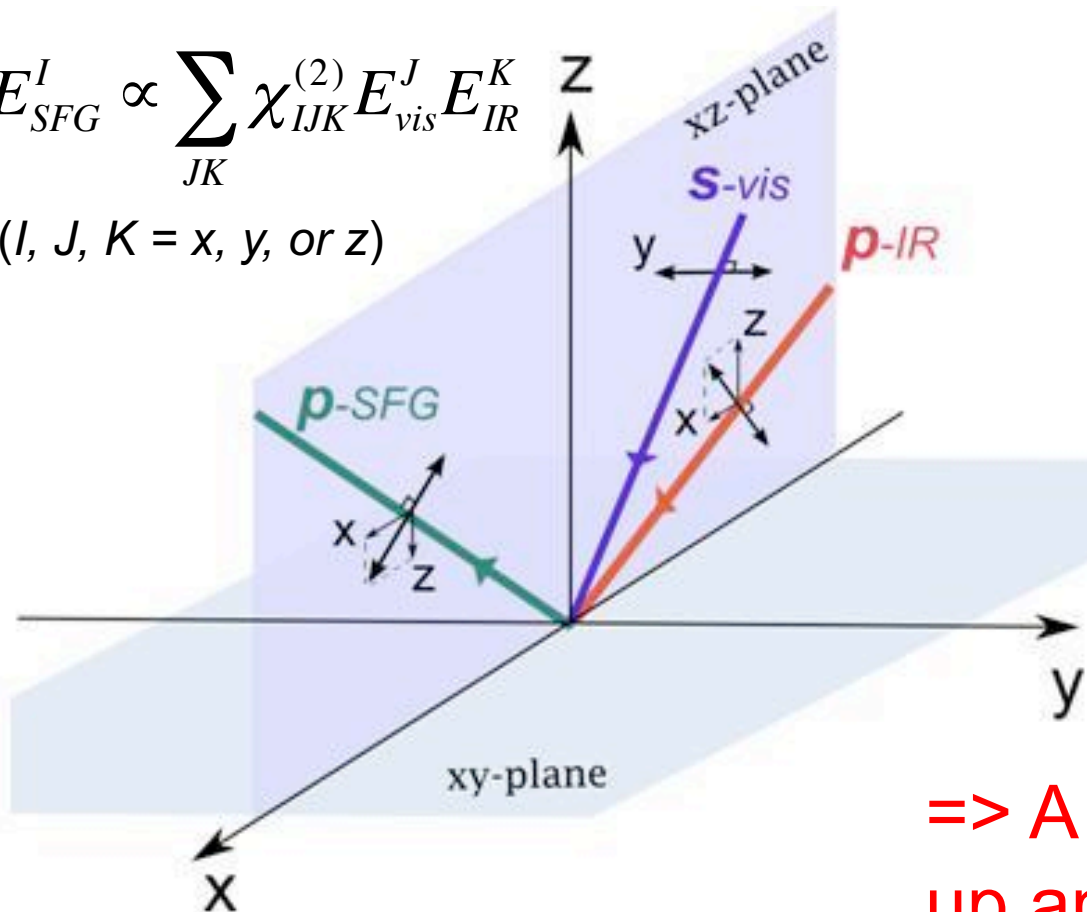
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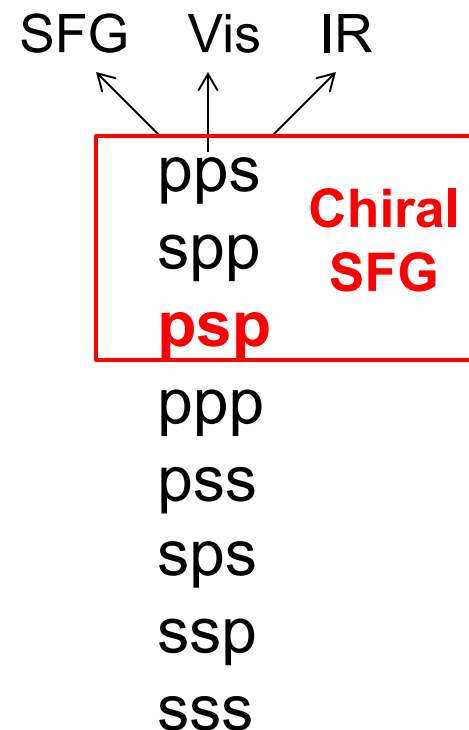
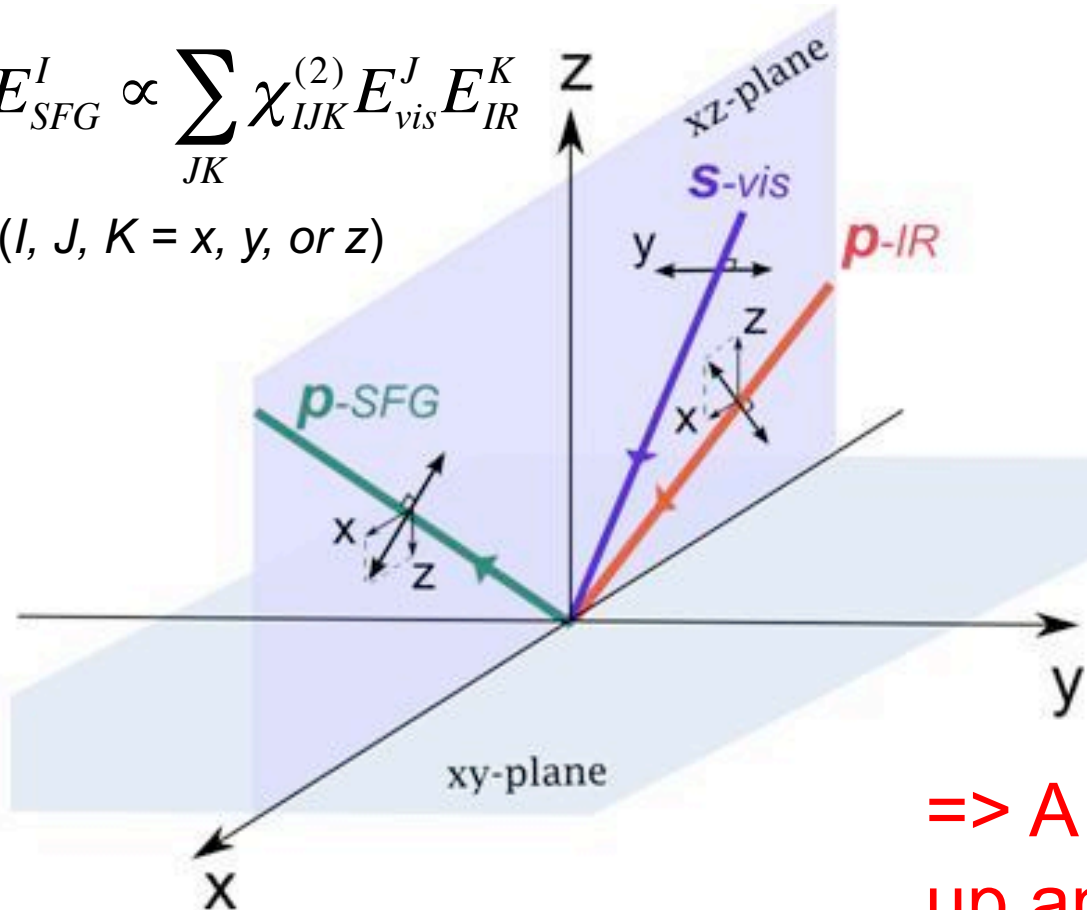
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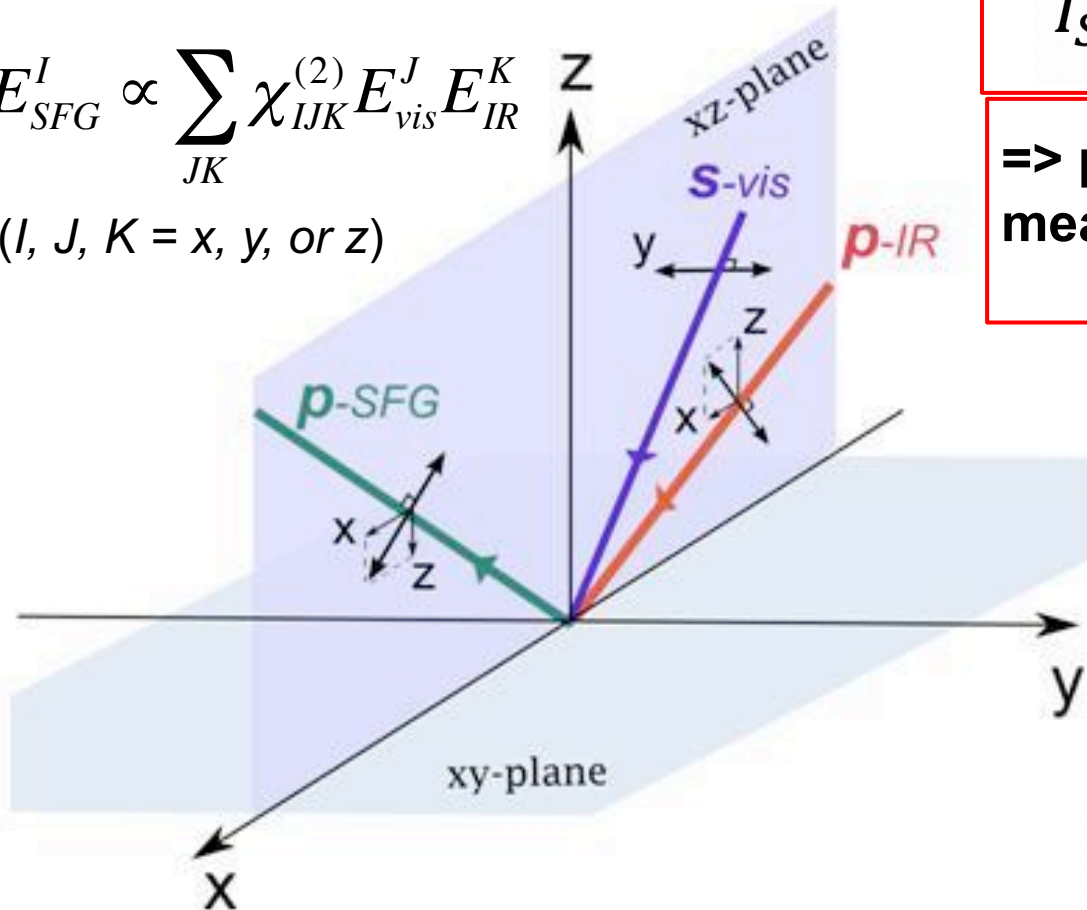
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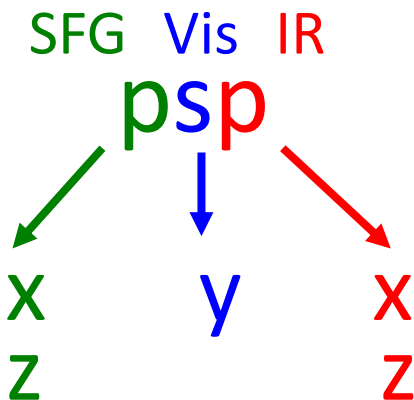
(I, J, K = x, y, or z)



Experimental observable:

$$I_{SFG} \propto |E_{SFG}|^2 \propto |\chi_{zyx}^{(2)}|^2$$

=> psp polarization selectively measuring $\chi_{zyx}^{(2)}$



$$\chi_{ijk}^{(2)}: \overset{0}{\cancel{\chi_{xyx}^{(2)}}}, \overset{0}{\cancel{\chi_{xyz}^{(2)}}}, \chi_{zyx}^{(2)}, \overset{0}{\cancel{\chi_{zyz}^{(2)}}}$$

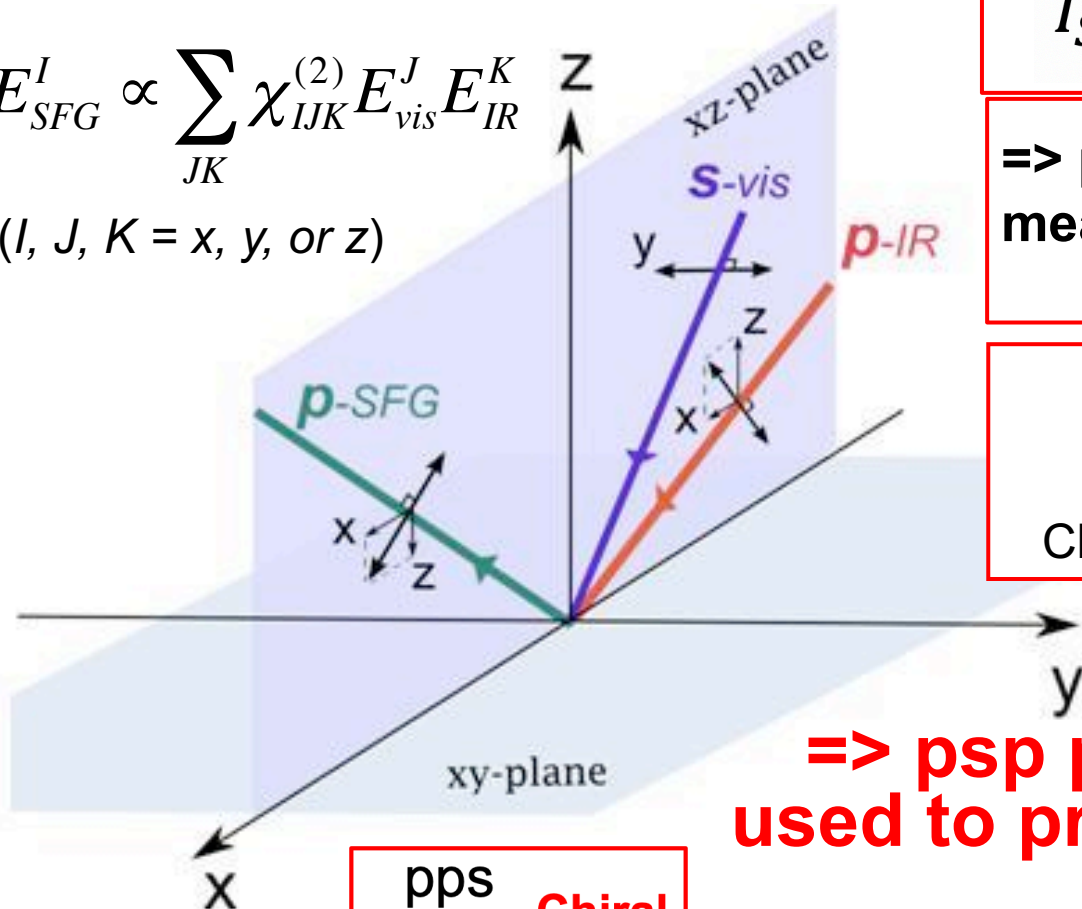
Introduction to Chiral SFG Spectroscopy

$$I_{SFG} = |\vec{E}_{SFG}|^2$$

$$\vec{E}_{SFG} = E_{SFG}^x \hat{x} + E_{SFG}^y \hat{y} + E_{SFG}^z \hat{z}$$

$$E_{SFG}^I \propto \sum_{JK} \chi_{IJK}^{(2)} E_{vis}^J E_{IR}^K$$

(I, J, K = x, y, or z)



Experimental observable:

$$I_{SFG} \propto |E_{SFG}|^2 \propto |\chi_{zyx}^{(2)}|^2$$

=> psp polarization selectively measuring $\chi_{zyx}^{(2)}$

$\chi_{IJK}^{(2)} (I \neq J \neq K):$

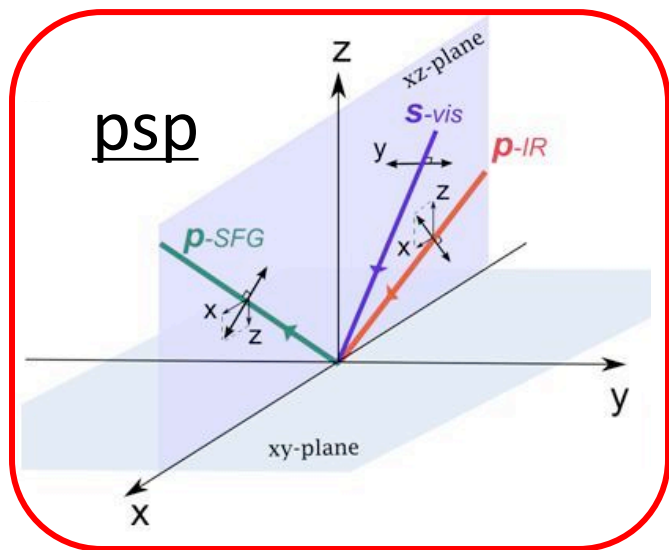
Orthogonal $\chi^{(2)}$ Element

Characteristics of Chiral Surfaces

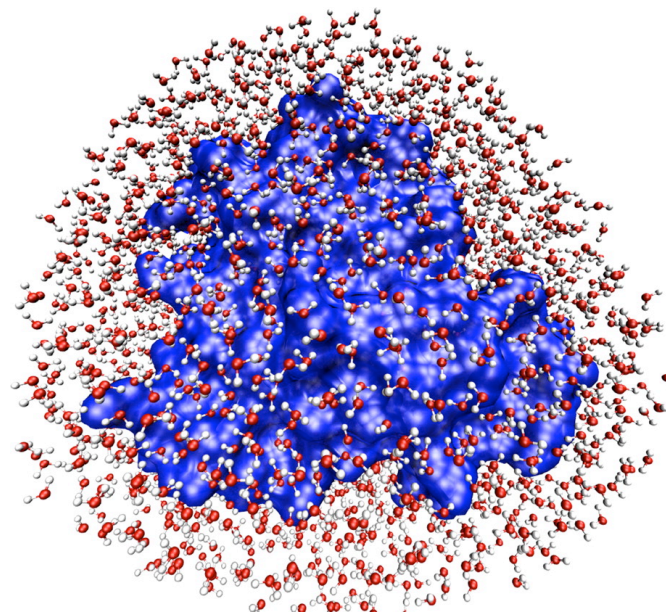
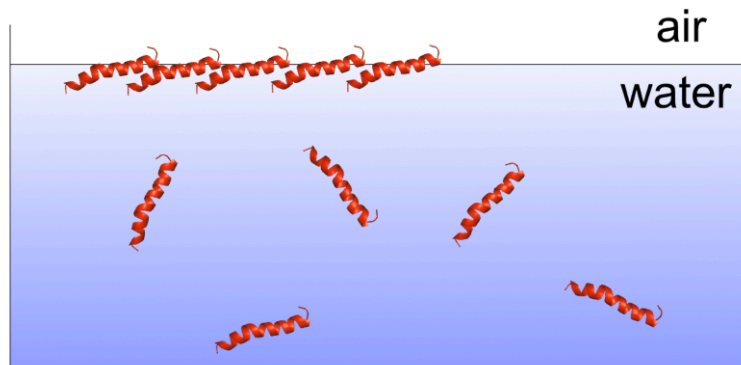
=> psp polarization can be used to probe surface chirality

pps
spp
psp **Chiral SFG**

Introduction to Chiral SFG Spectroscopy



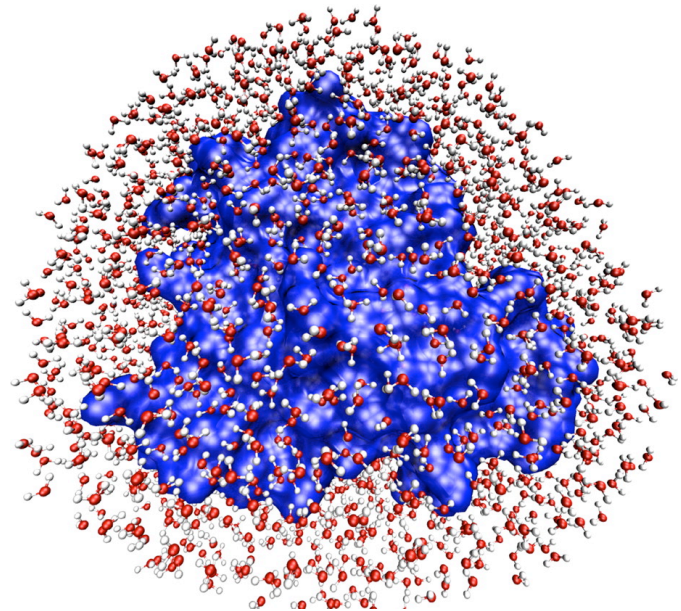
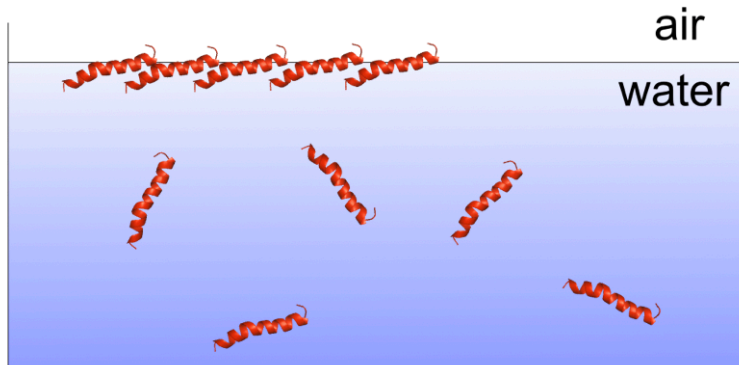
- I. Protein Secondary Structures at Interfaces
- II. Water Structures in Protein Hydration



Protein Characterizations by Chiral Vibrational Sum Frequency Generation Spectroscopy

Outline

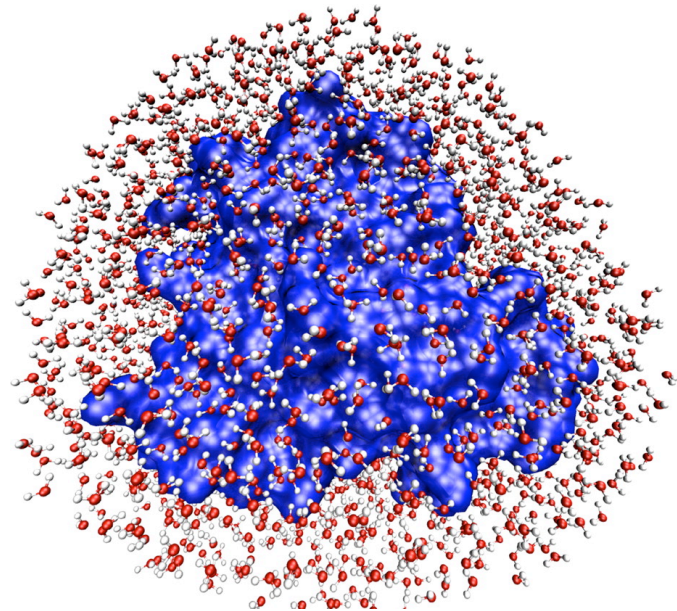
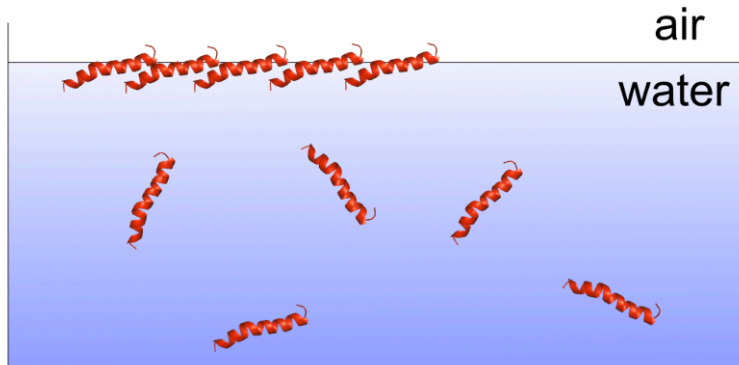
1. Introduction to Chiral SFG Spectroscopy
2. Charactering Protein Secondary Structures at Interfaces
3. Probing Water Structures in Hydration Shell of Proteins



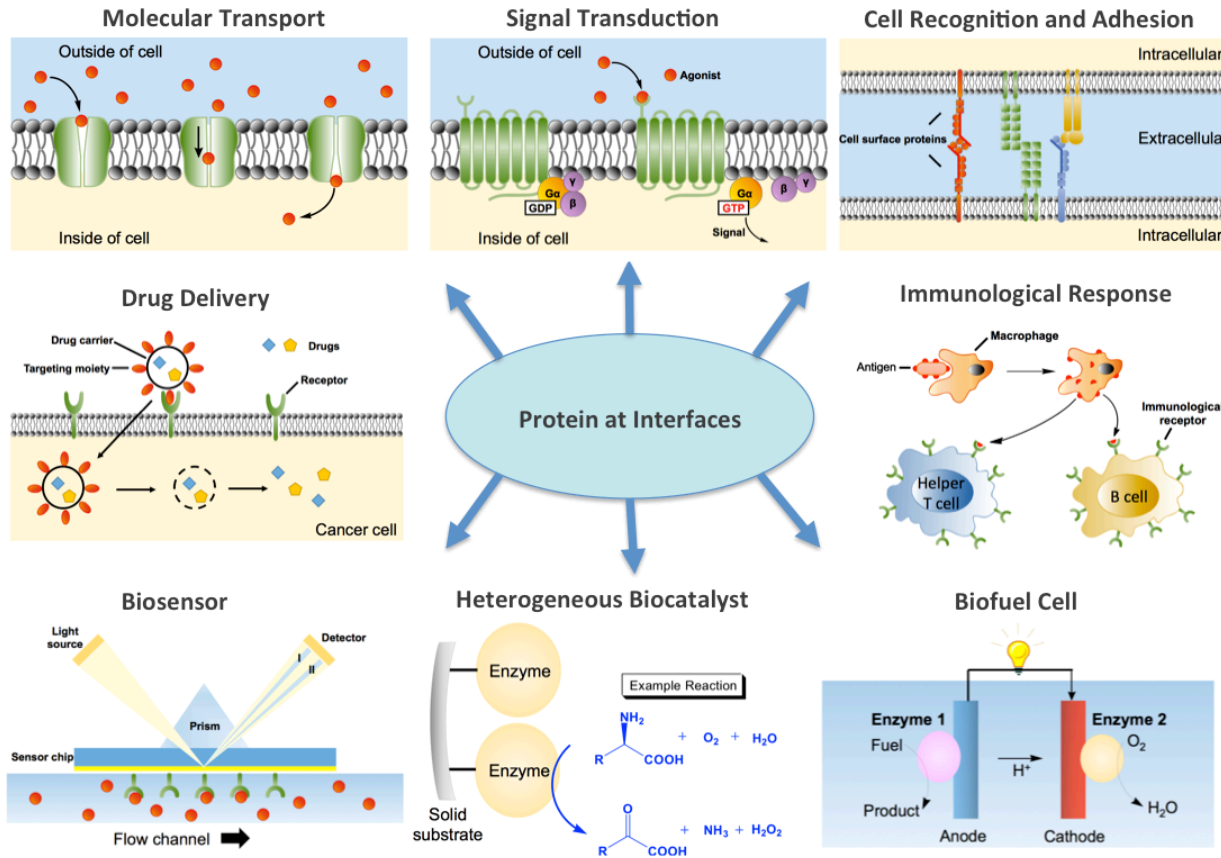
Protein Characterizations by Chiral Vibrational Sum Frequency Generation Spectroscopy

Outline

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2. Charactering Protein Secondary Structures at Interfaces
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Protein Secondary Structures at Interfaces

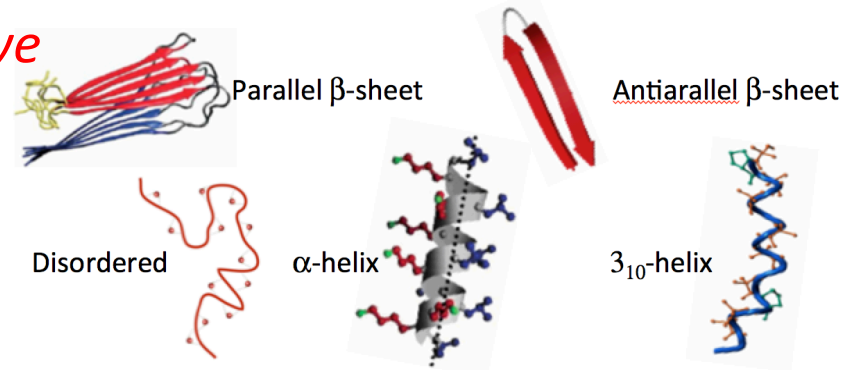


Challenge: Selectivity to
 (1) Interfaces
 (2) Secondary Structures



Conventional Methods—*Not surface selective*

1. Circular Dichroism Spectroscopy
2. NMR Spectroscopy
3. X-ray crystallography



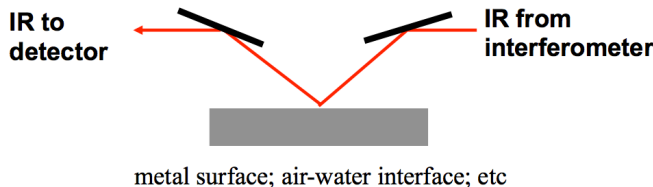
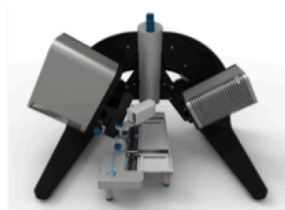
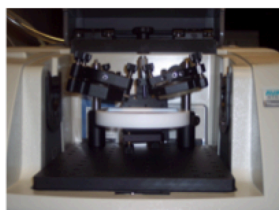
Protein Secondary Structures at Interfaces

Challenge: Selectivity to

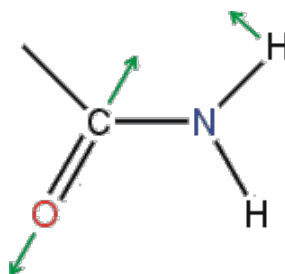
(1) Interfaces

(2) Secondary Structures

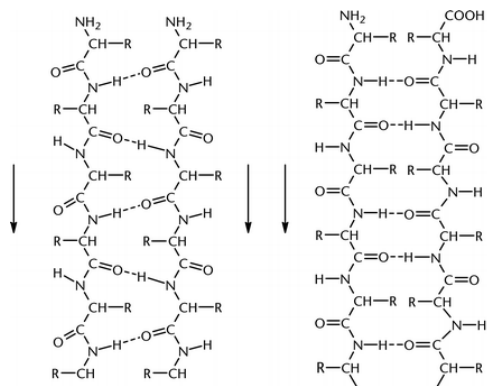
Infrared Reflection Absorption Spectroscopy (IRRAS)



Amide I

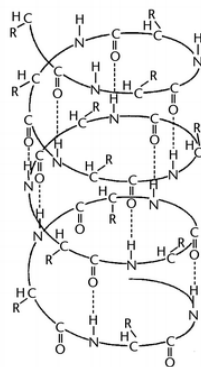


Secondary structure	Band position in H ₂ O/cm ⁻¹	
	Average	Extremes
α-helix	1654	1648-1657
β-sheet	1633	1623-1641
β-sheet	1684	1674-1695
Turns	1672	1662-1686
Disordered	1654	1642-1657



Parallel β pleated sheet

Antiparallel β pleated sheet



Right-handed α helix

Spectral overlapping

- Deconvolution
- Overlap with water bending
- D₂O as solvent

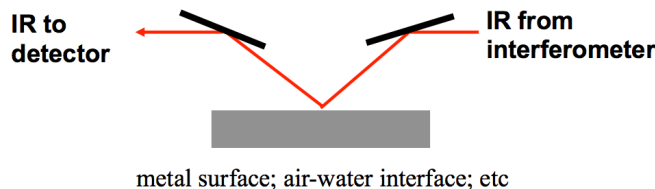
Protein Secondary Structures at Interfaces

Challenge: Selectivity to

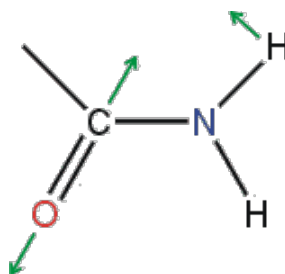
(1) Interfaces

(2) Secondary Structures

Infrared Reflection Absorption Spectroscopy (IRRAS)

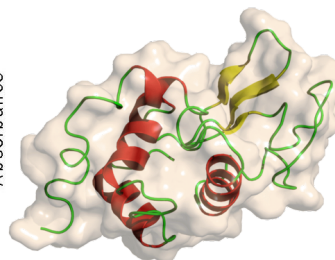
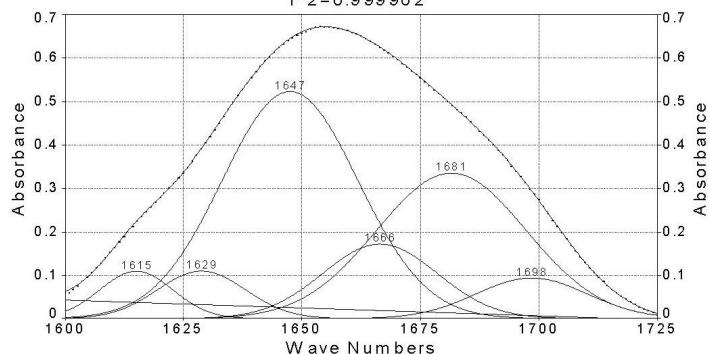


Amide I



Secondary structure	Band position in H ₂ O/cm ⁻¹	
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β -sheet	1684	1674-1695
Turns	1672	1662-1686
Disordered	1654	1642-1657

Lysozyme
Gauss 6 Peaks
 $r^2=0.999962$



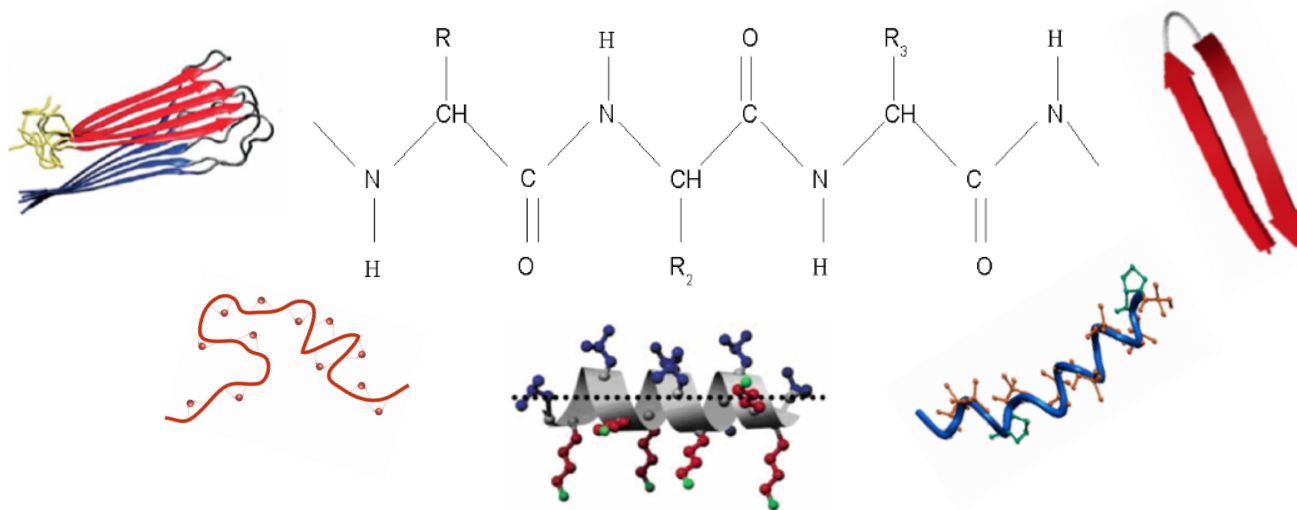
Spectral overlapping

- Deconvolution
- Overlap with water bending
- D₂O as solvent

Protein Secondary Structures at Interfaces

Challenge: Selectivity to
(1) Interfaces
(2) Secondary Structures

Chiral Sum Frequency Generation Spectroscopy

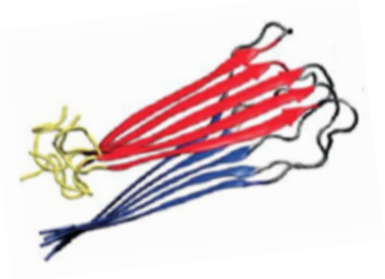


- Ron Shen (UC Berkeley)
- Garth Simpson (Purdue)
- Zhan Chen (U Michigan)
- Franz Geiger (Northwestern U)

Protein Backbone (Amide I and N-H Stretch) Chiral Vibrational Structures
Characterization of Protein Secondary Structures at Interfaces

Characterization of Secondary Structures at Interfaces

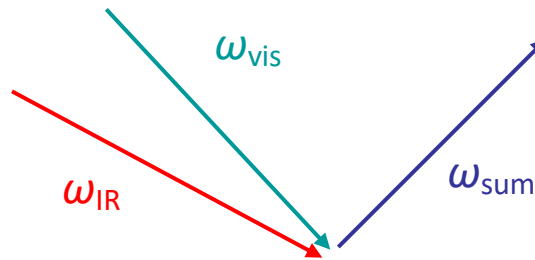
Parallel β -sheet



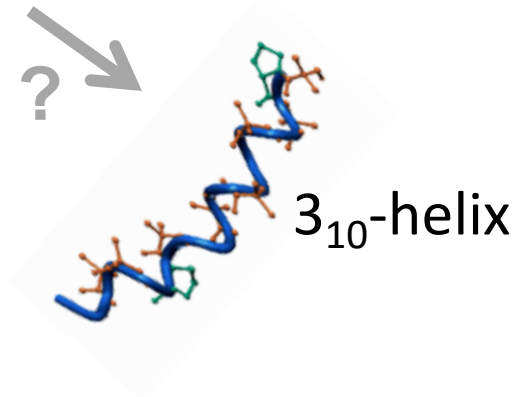
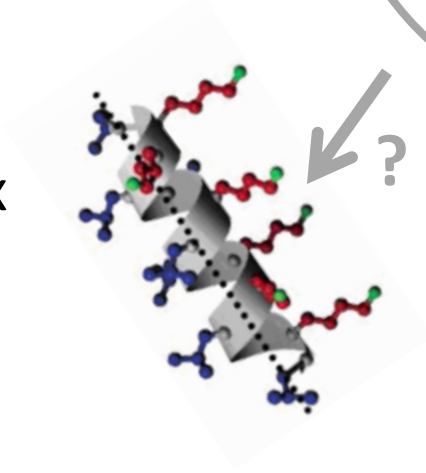
Antiparallel β -sheet



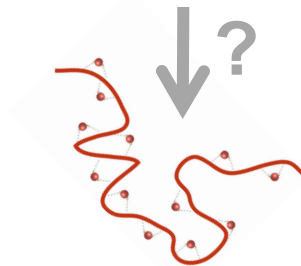
**SFG Chiral
Vibrational Signatures**



α -helix

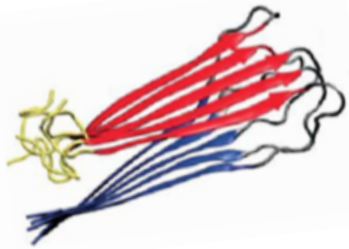


Unstructured



Characterization of Secondary Structures at Interfaces

Parallel β -sheet



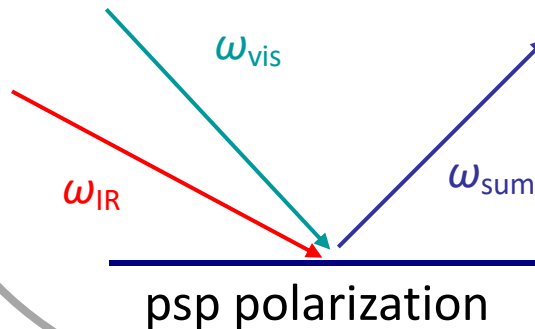
Model:
The hIAPP
aggregate

Antiparallel β -sheet

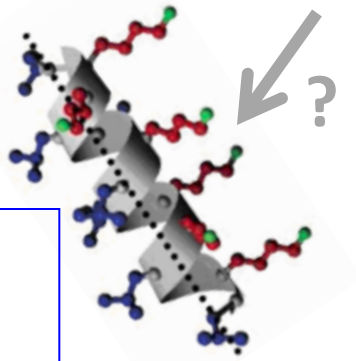


Model:
Tachyplesin I
LK $_7\beta$

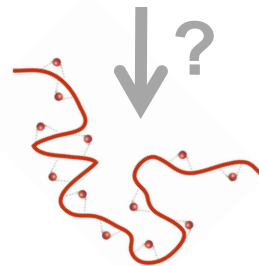
**SFG Chiral
Vibrational Signatures**



α -helix

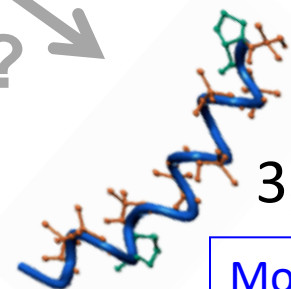


Model:
Rhodopsin
pHLIP
LK α_{14}



Unstructured

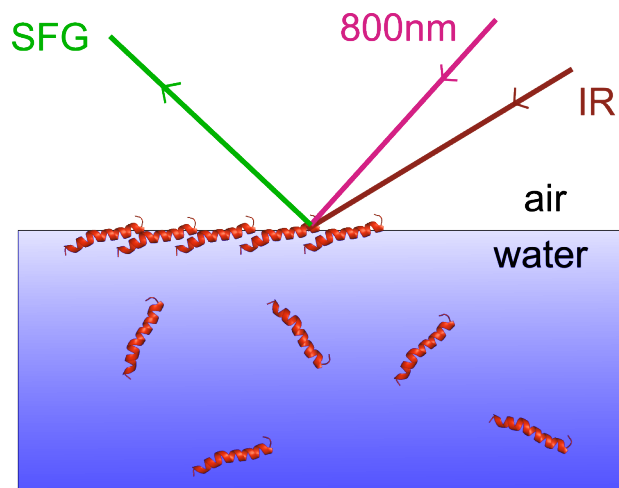
Model:
disordered
rIAPP



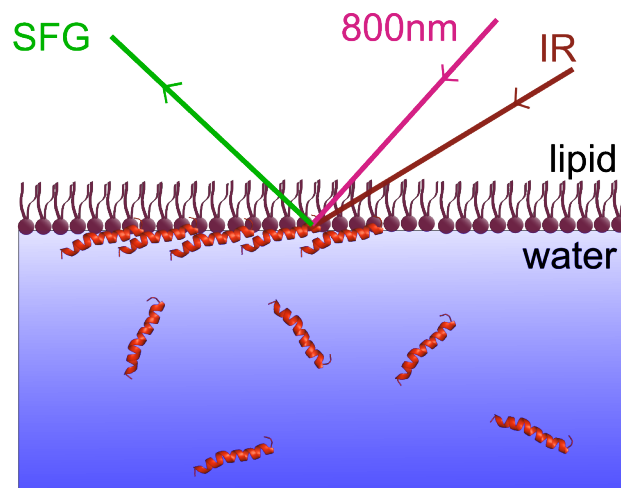
3_{10} -helix

Model:
Alamethicin
n

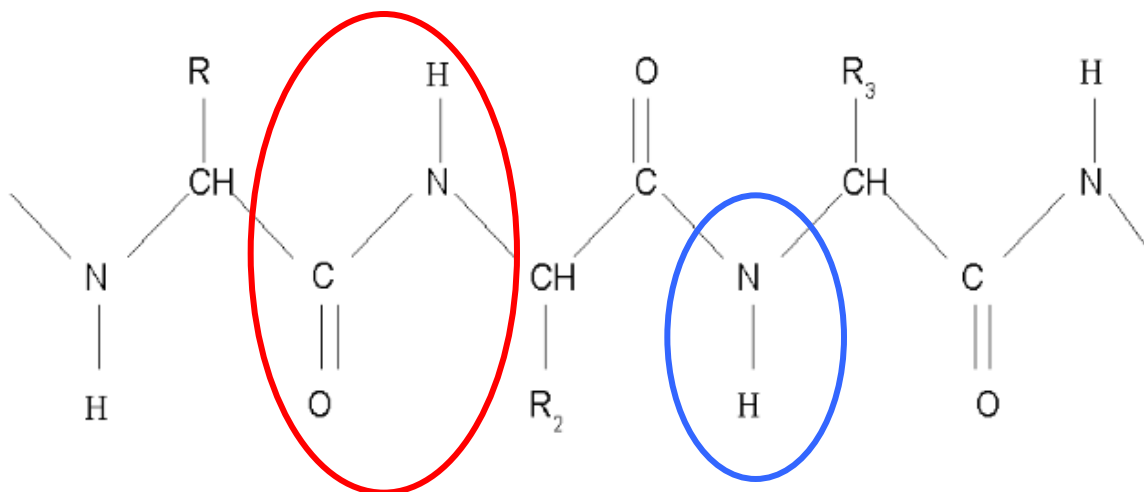
Chiral SFG Experiments—psp Polarization



or



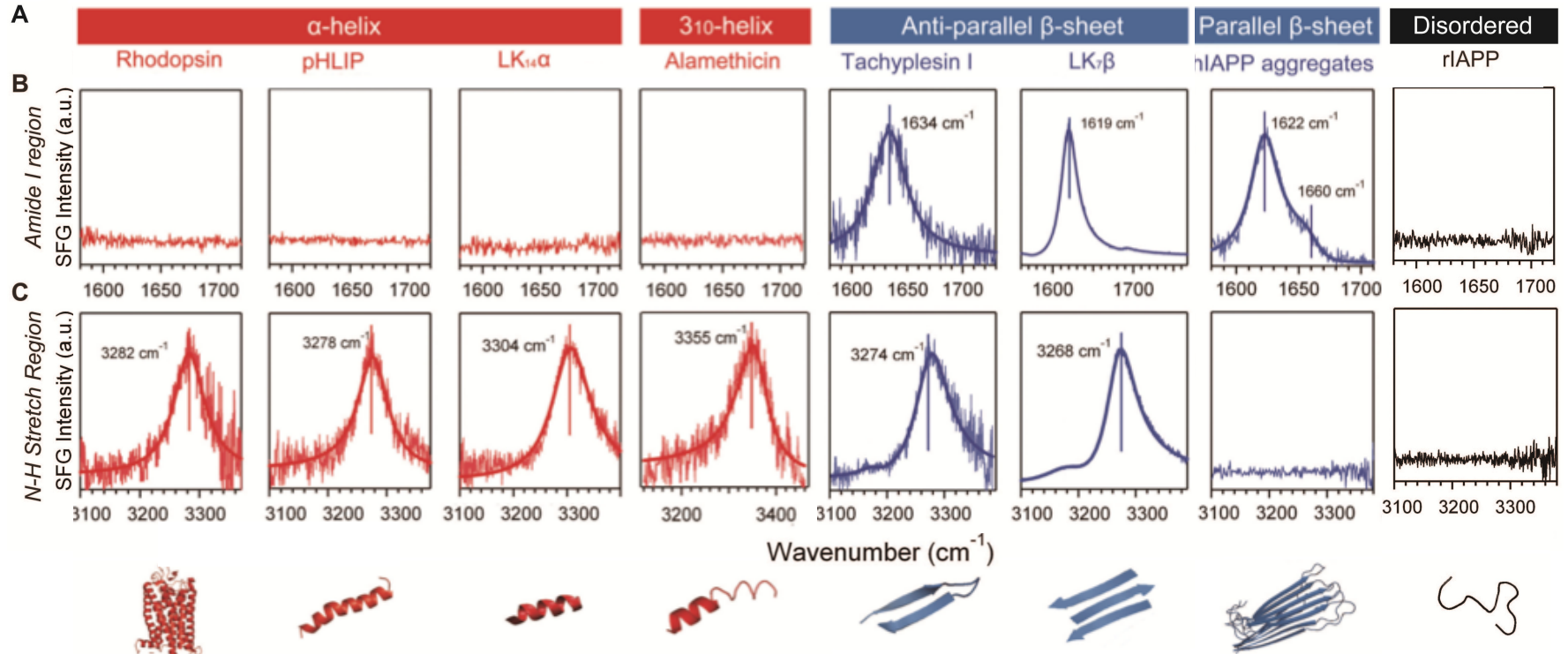
Peptide Backbone



The Amide I Band ($1600-1700\text{ cm}^{-1}$)

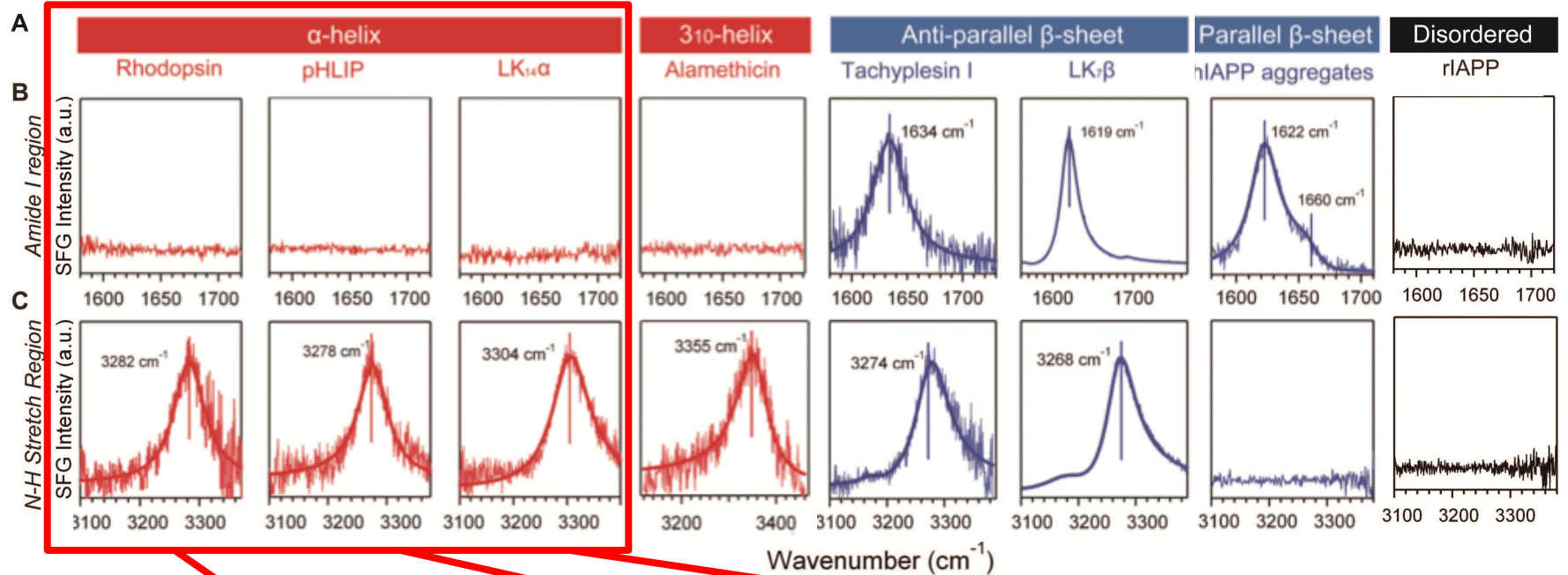
The N-H Stretch ($3000-3400\text{ cm}^{-1}$)

cSFG Vibrational Signatures for Secondary Structures



- Yan *et al* *Chem. Rev.* 2014
- Fu *et al* *Int. J. Mol. Sci.* 2011
- Yan *et al.* *J. Phys. Chem. B* 2015
- Wang *et al.* *Langmuir*, 2015

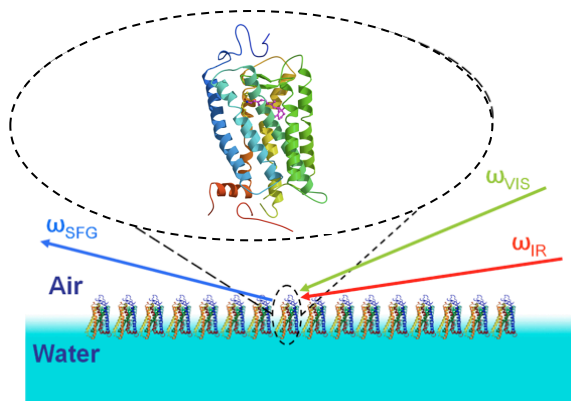
cSFG Vibrational Signatures for Secondary Structures



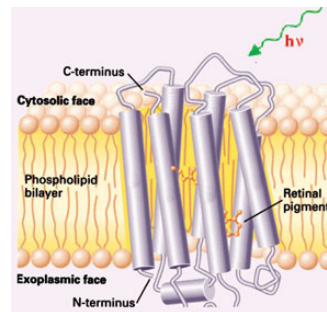
Rhodopsin

pHLIP

LK₁₄ α

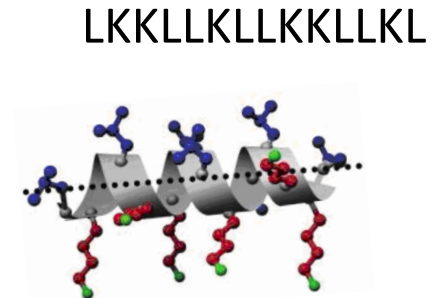


(Lavoie et al. *Biochemistry*, 2002)

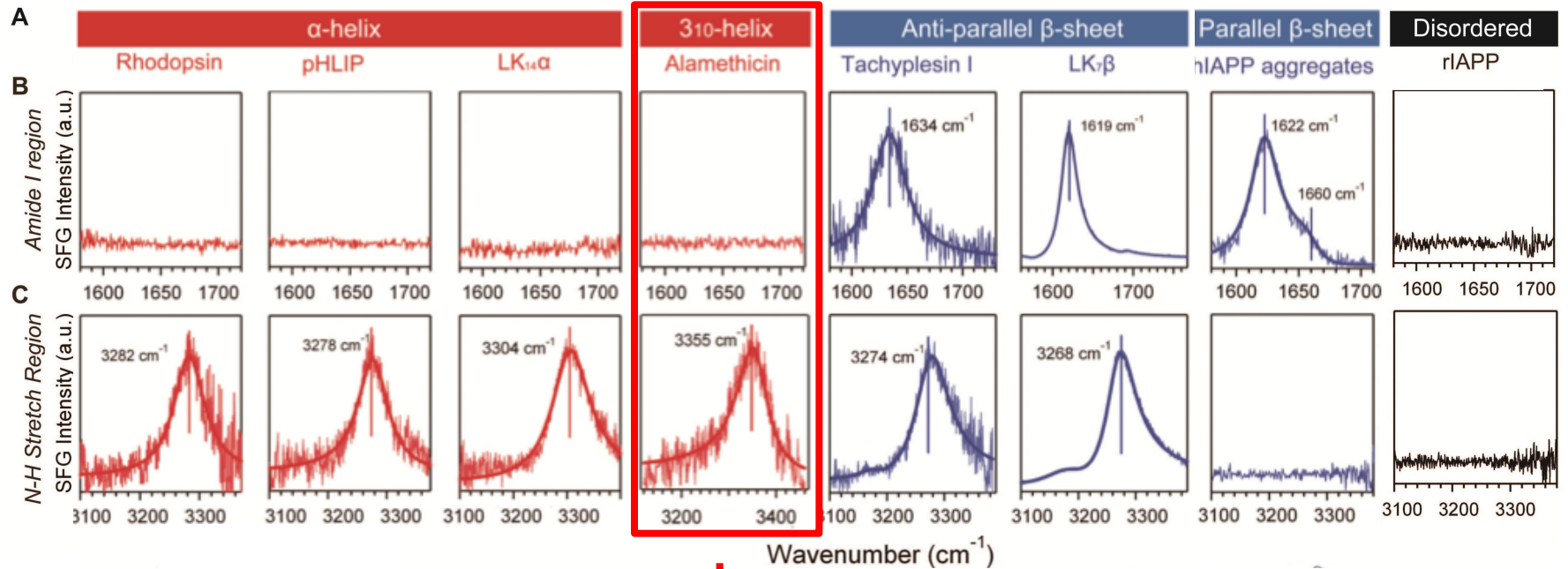


Helix 3 of
Bacteriorhodopsin

(Englman and coworkers, PNAS, 2010) (DeGrado and Coworkers JACS, 1985)

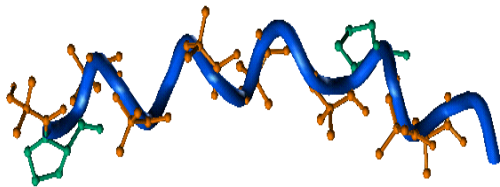


cSFG Vibrational Signatures for Secondary Structures



A mixture of α -helix & 3_{10} helix

Alamethicin

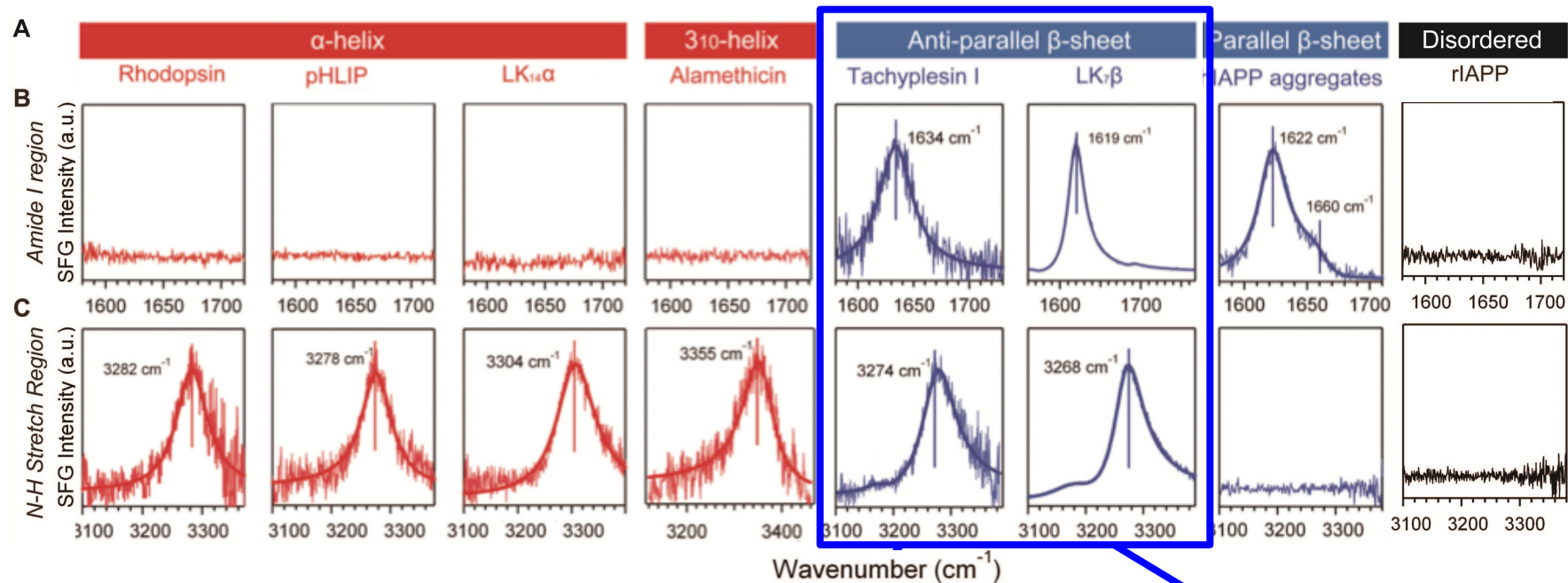


(Ionov et al. *J. Phys. Chem.* 2004)

Ac-Aib-L-Pro-Aib-L-Ala-Aib-L-
Ala-L-Gln-Aib-L-Val-Aib-Gly-L-
Leu-Aib-L-Pro-L-Val-Aib-Aib-L-
Glu-L-Gln-L-Phol

- Yan et al *Chem. Rev.* 2014
- Fu et al *Int. J. Mol. Sci.* 2011
- Yan et al. *J. Phys. Chem. B* 2015
- Wang et al. *Langmuir*, 2015

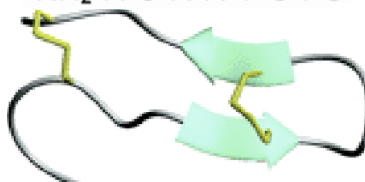
cSFG Vibrational Signatures for Secondary Structures



Tachyplesin I

H-K-W-C-F-R-V-C-Y-R

NH₂-R-C-R-R-Y-C-I-G



(Nakamura *et al.* *J. Biol. Chem.* **1988**)

(Chen and coworker, *PNAS*, **2005**)

LK₇ β

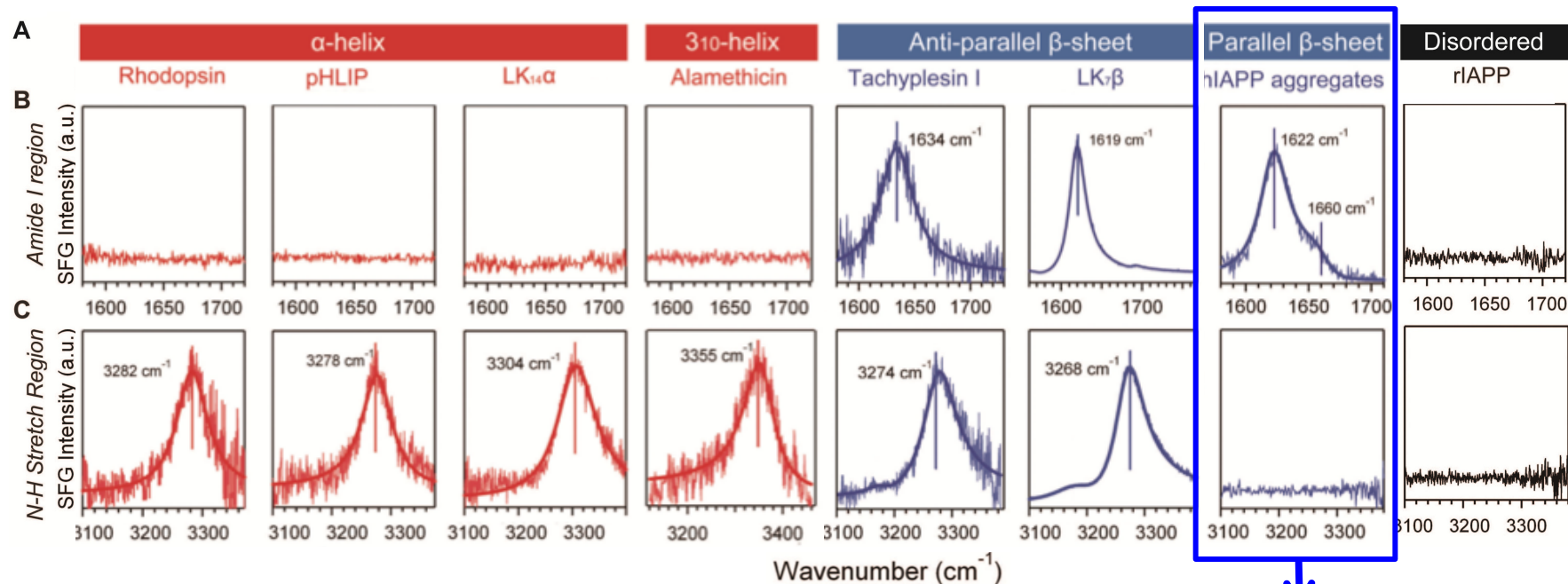
LK₇ β



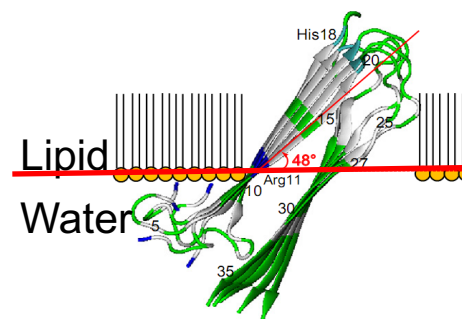
(DeGraau and coworker, *JACS* **1985**)

- Yan *et al* *Chem. Rev.* 2014
- Fu *et al* *Int. J. Mol. Sci.* 2011
- Yan *et al.* *J. Phys. Chem. B* 2015
- Wang *et al.* *Langmuir*, 2015

cSFG Vibrational Signatures for Secondary Structures

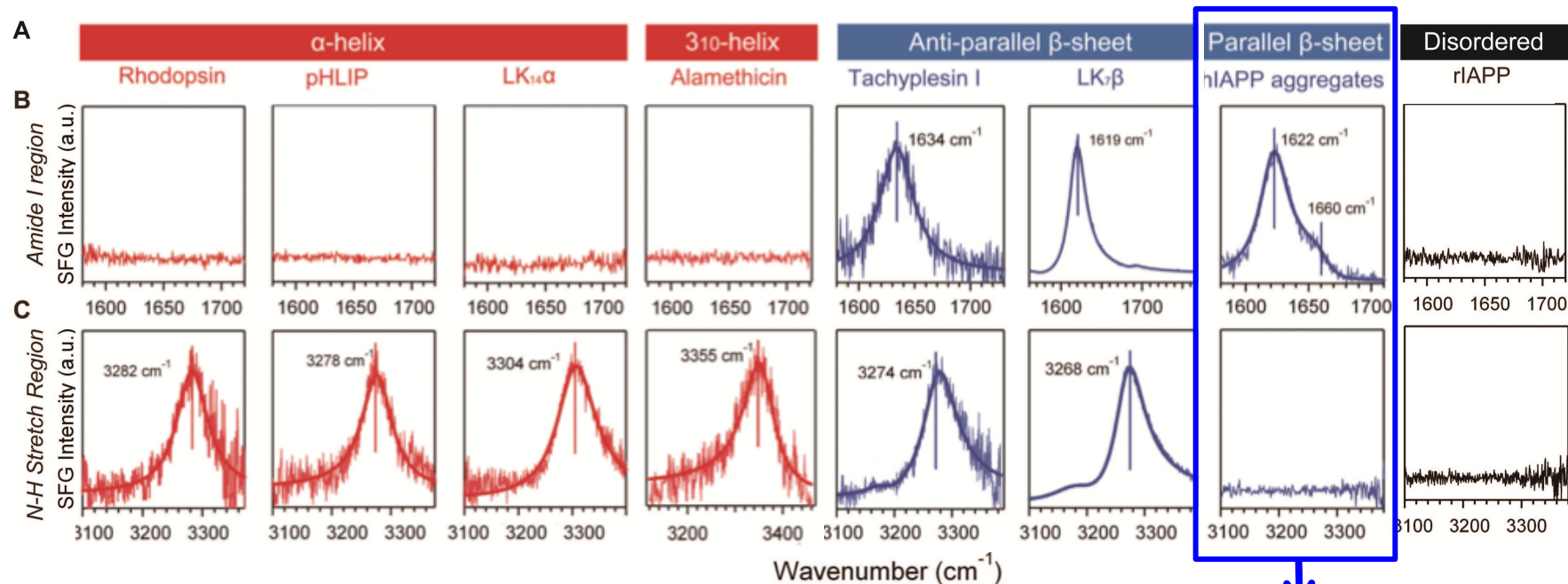


hIAPP aggregate

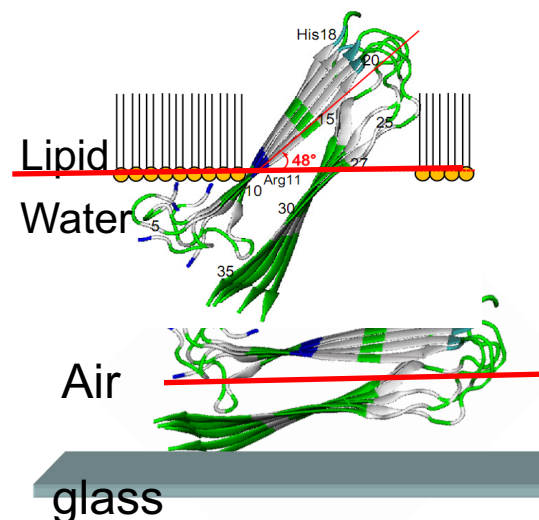


- Yan *et al* *Chem. Rev.* 2014
- Fu *et al* *Int. J. Mol. Sci.* 2011
- Yan *et al.* *J. Phys. Chem. B* 2015
- Wang *et al.* *Langmuir*, 2015

cSFG Vibrational Signatures for Secondary Structures

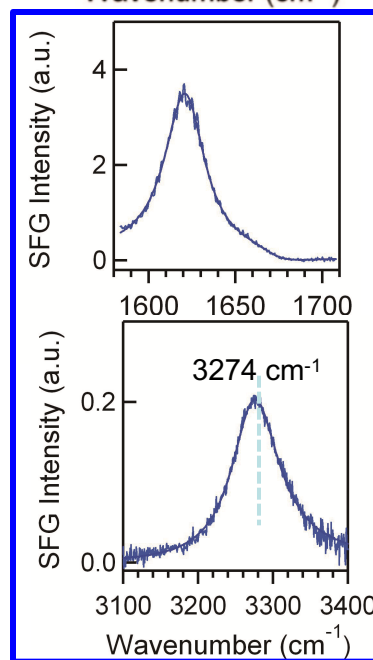
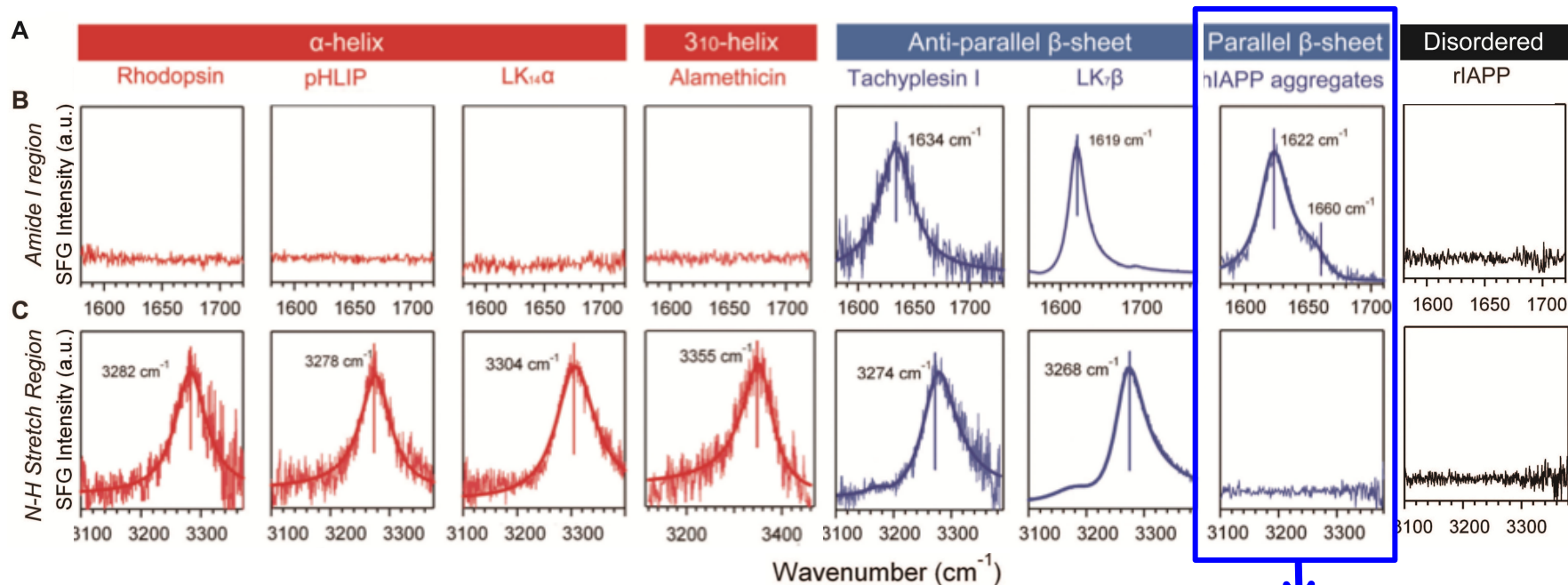


hIAPP aggregate

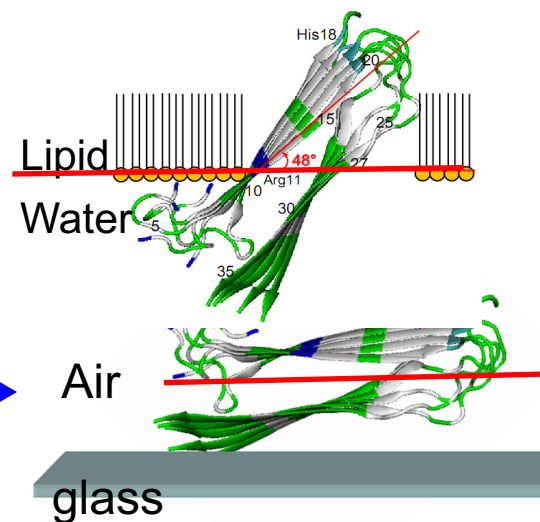


- Yan *et al* *Chem. Rev.* 2014
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- Yan *et al.* *J. Phys. Chem. B* 2015
- Wang *et al.* *Langmuir*, 2015

cSFG Vibrational Signatures for Secondary Structures

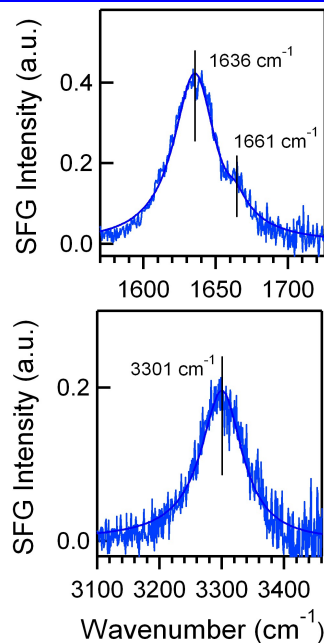
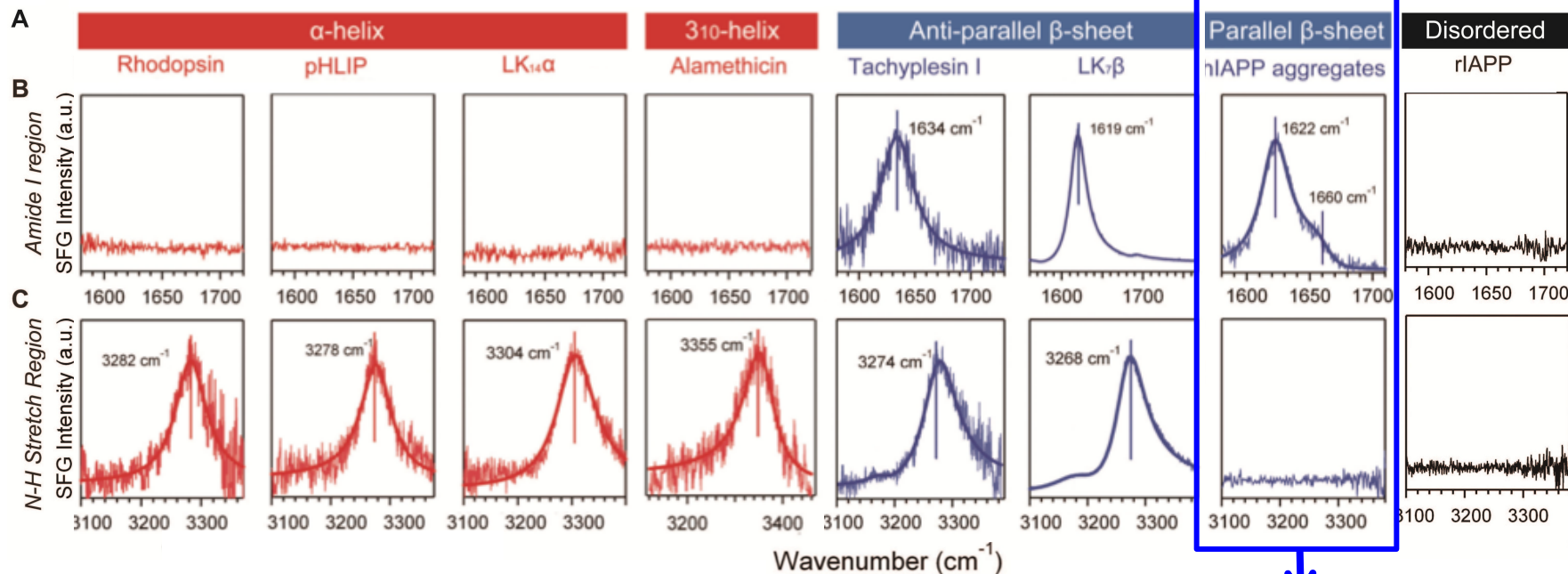


hIAPP aggregate

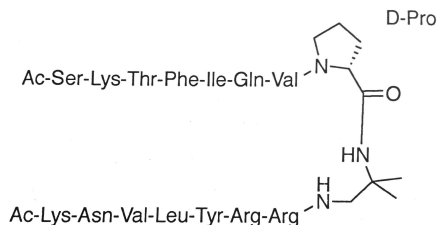


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cSFG Vibrational Signatures for Secondary Structures

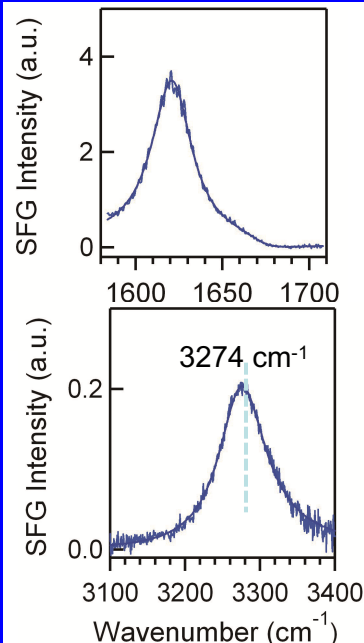


SJM2116a (Peptide 1 in the paper)

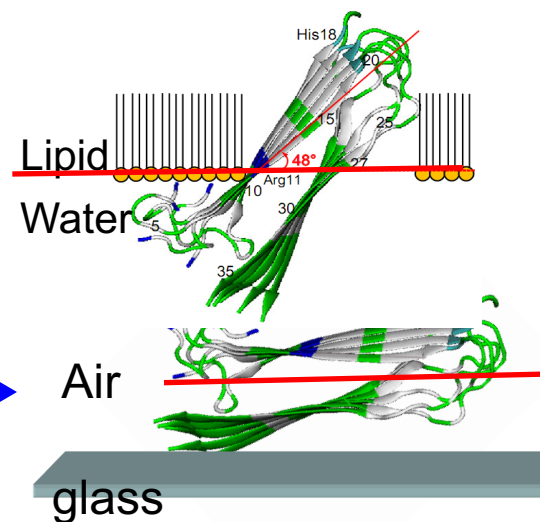


(Gellman, U Wisconsin)

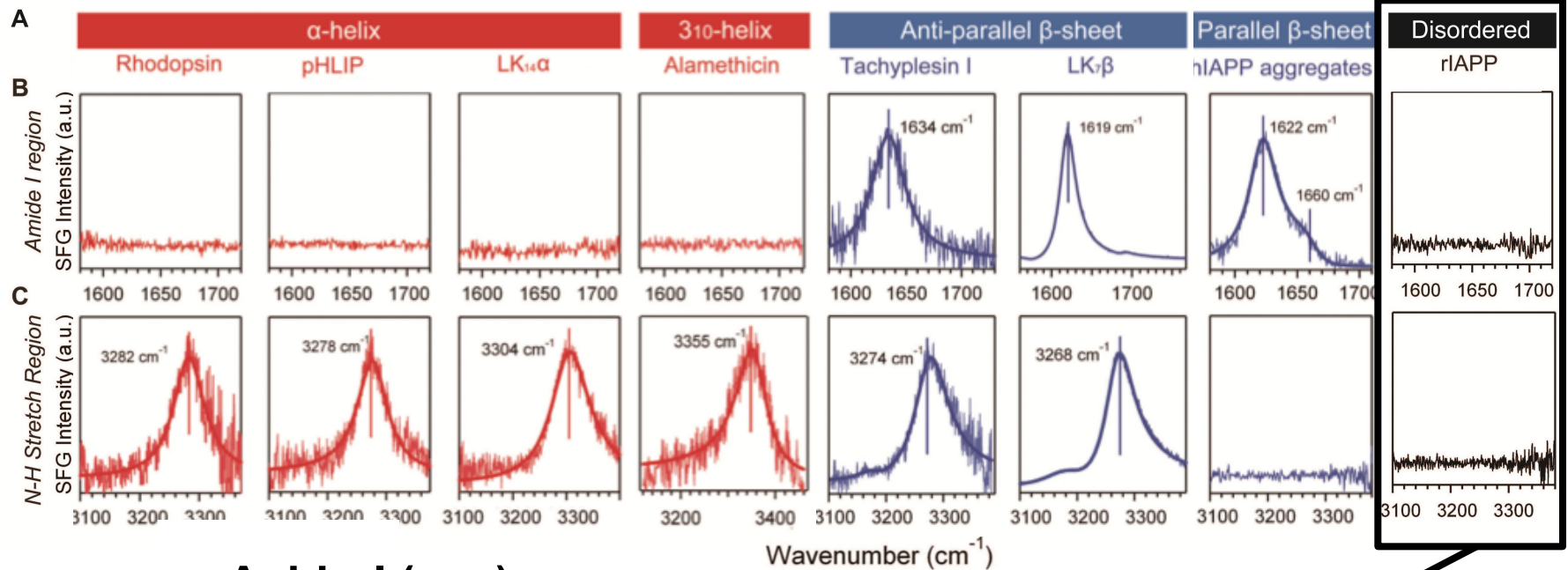
J. Am. Chem. Soc. **2001**,
123, 343-344



hIAPP aggregate



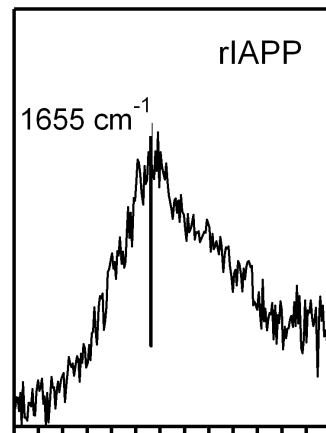
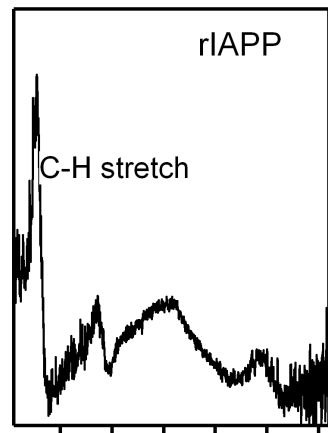
cSFG Vibrational Signatures for Secondary Structures



Achiral (ssp)

N-H Stretch

Amide I



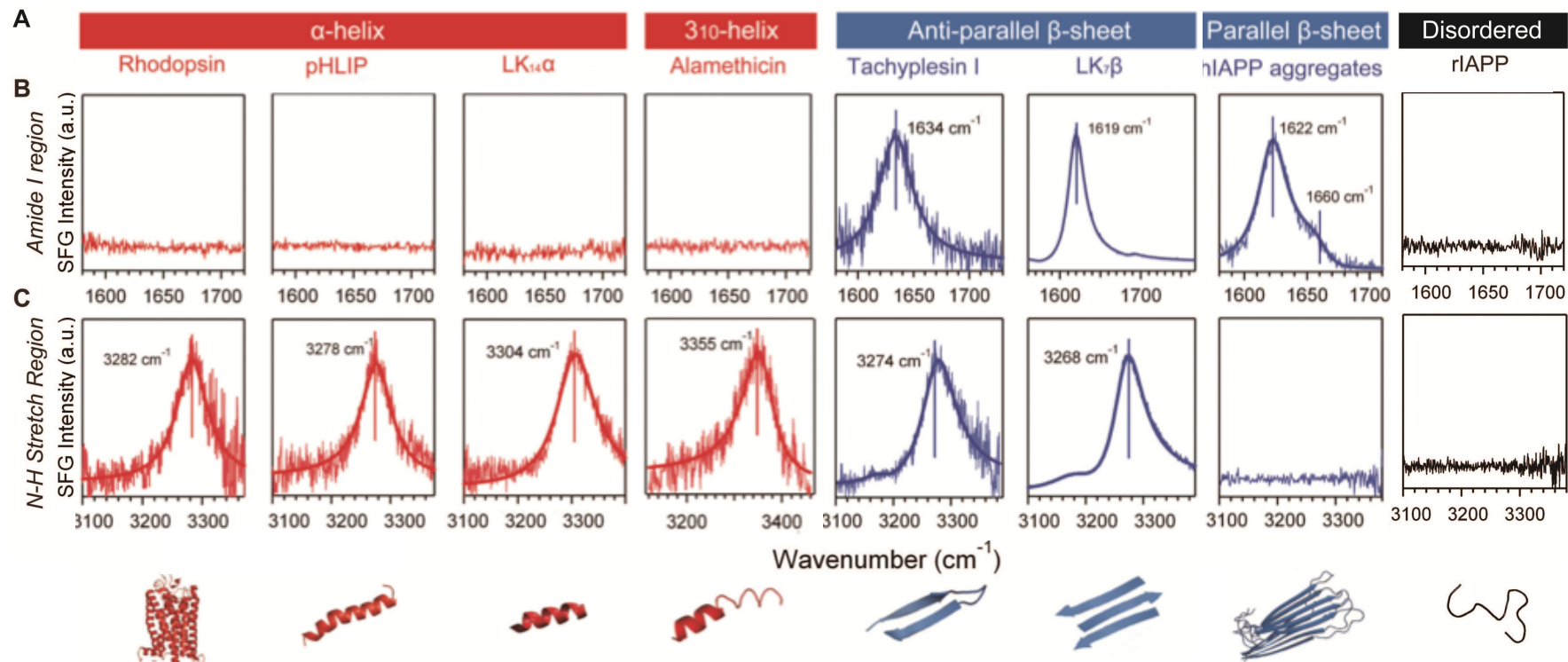
Wavenumber (cm⁻¹) Wavenumber (cm⁻¹)

Rat Islet Amyloid Polypeptide

Disordered



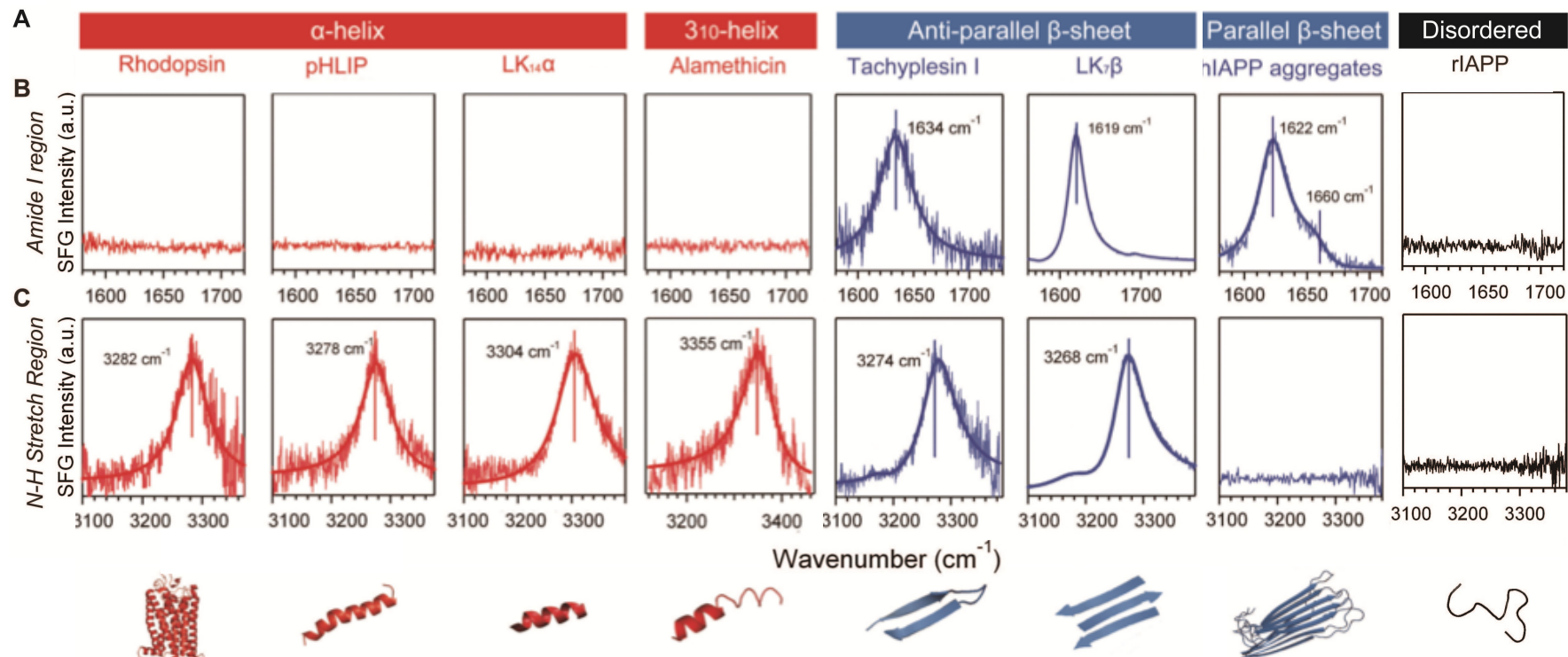
cSFG Vibrational Signatures for Secondary Structures



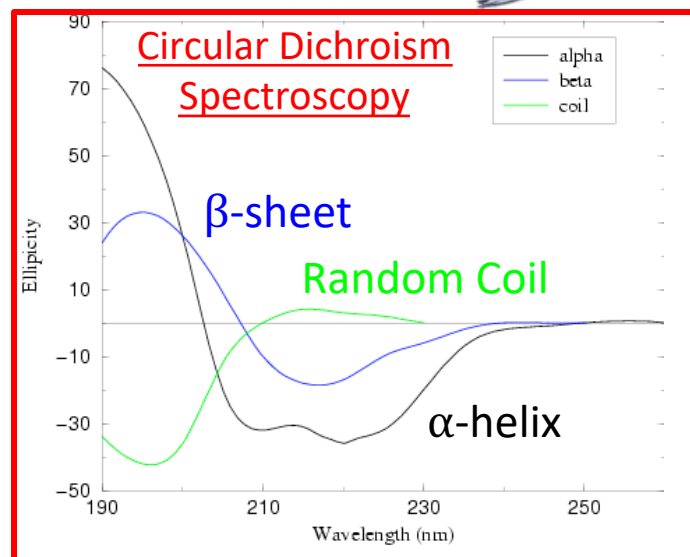
Chiral SFG: characterize
protein secondary
structures at interfaces

- Yan *et al* *Chem. Rev.* 2014
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cSFG Vibrational Signatures for Secondary Structures

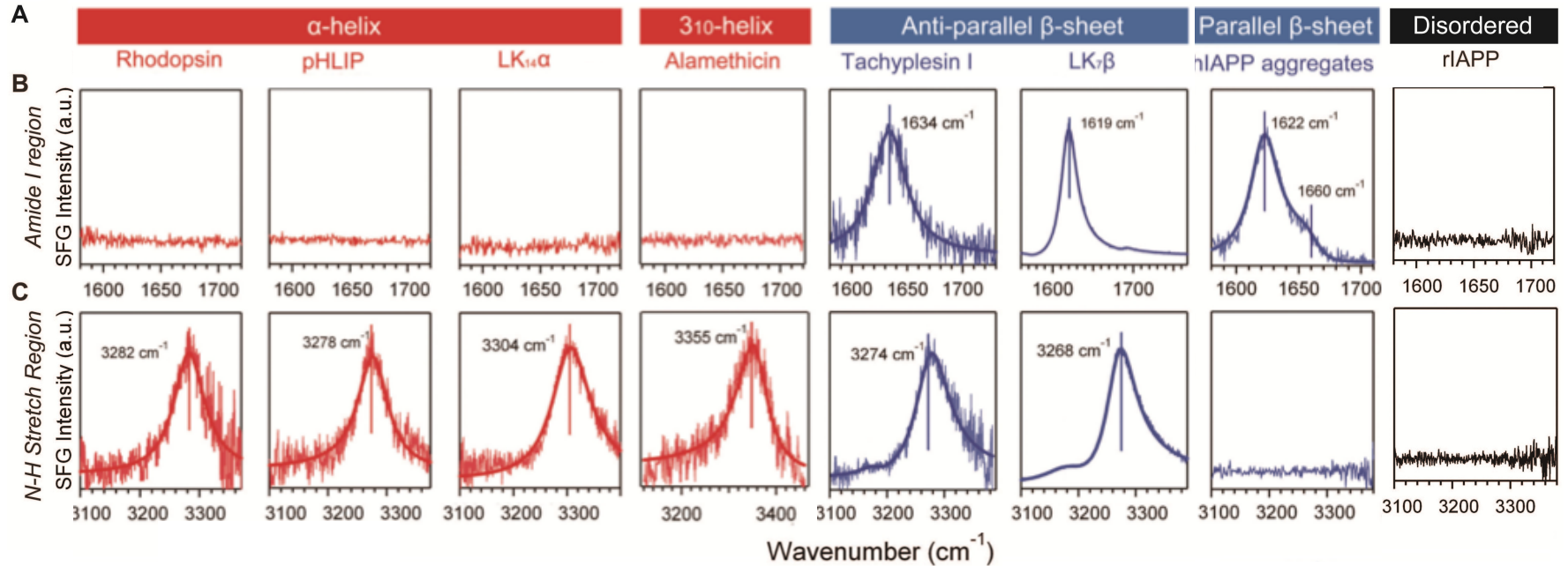


Chiral SFG: characterize protein secondary structures at interfaces



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cSFG Vibrational Signatures for Secondary Structures



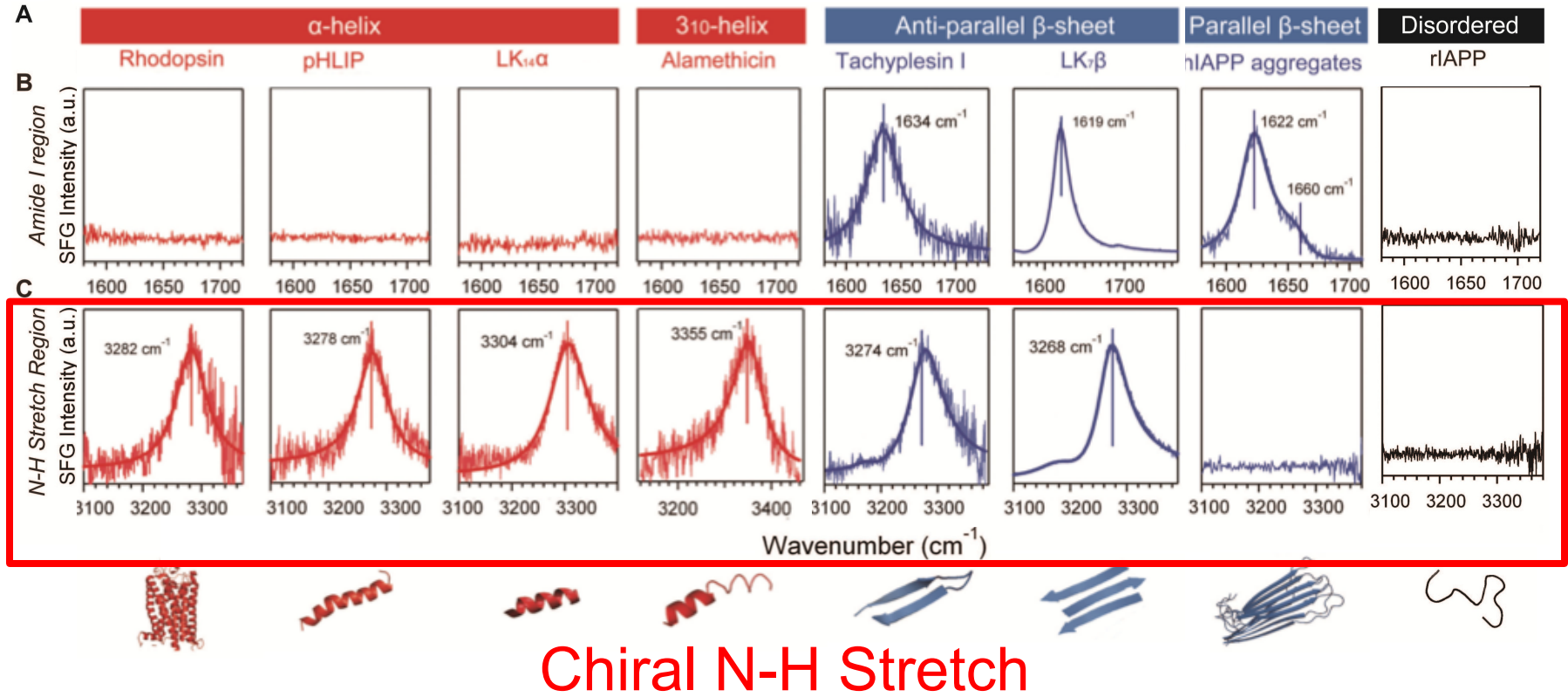
Compared to IR Spectroscopy

Chiral SFG

Secondary structure	Band position in H ₂ O/ cm^{-1}	
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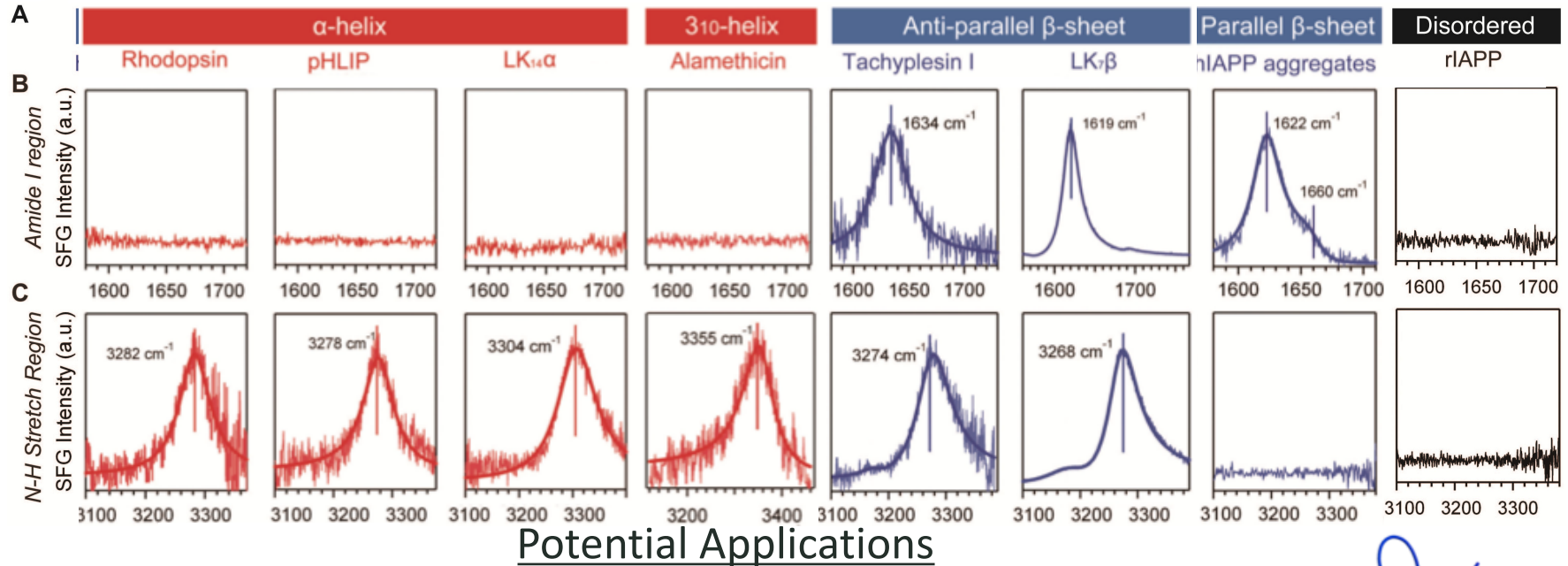
1. Spectrally well separated
2. Background Free from water
3. Optically clean
4. H₂O as solvent

cSFG Vibrational Signatures for Secondary Structures

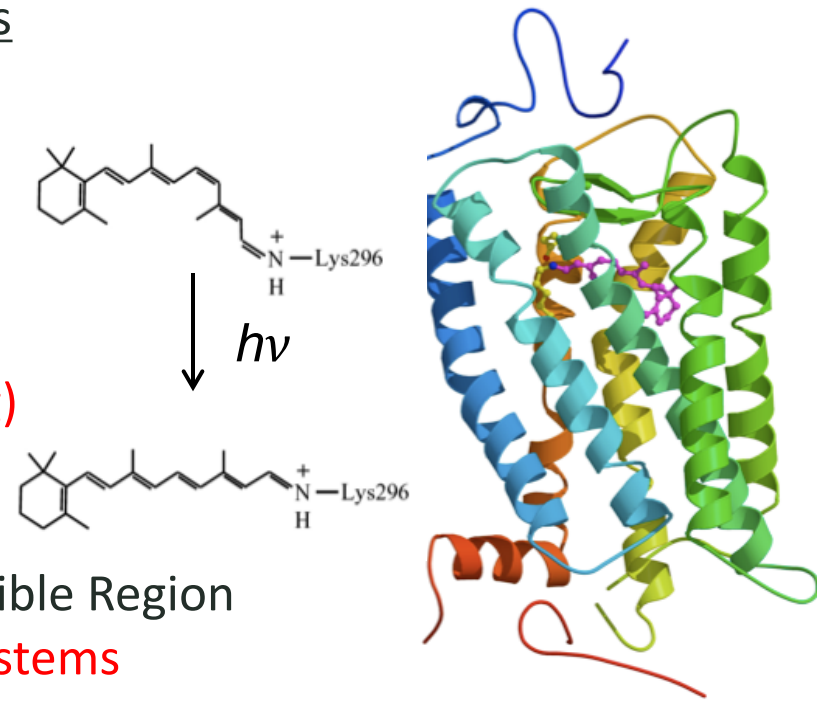


- Chiral SFG—Background-free from water O-H stretch
- Chiral SFG—Background-free from proteins in disordered structures
- N-H frequency—Secondary structures (Stabilization by H-bonds at N-H)
=> Characteristic of secondary structures

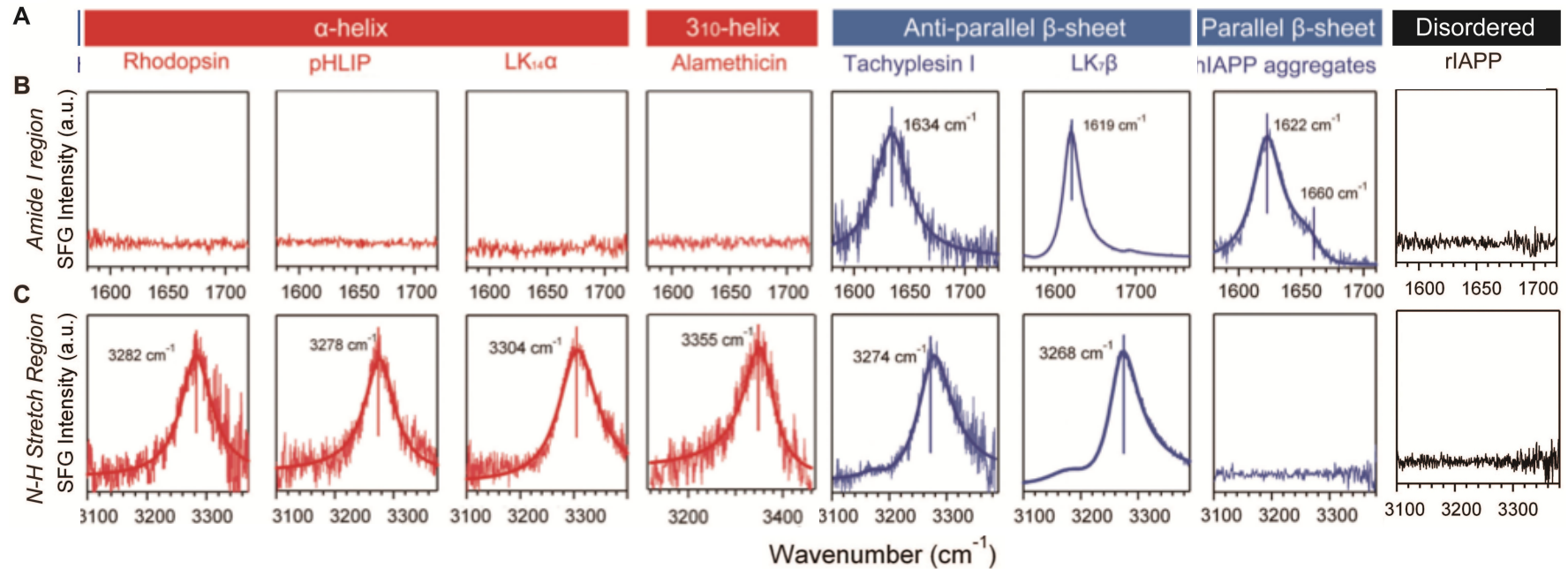
cSFG Vibrational Signatures for Secondary Structures



- Changes in Secondary Structures
=> Protein Folding at Interfaces
- Changes in Conformation and Orientation
=> Protein Functions at Interfaces
- Surface Sensitive—Monolayer of Proteins
=> Membrane Proteins--Small Sample Size ($\sim\mu\text{g}$)
- Background Free
=> Imaging of Biological Samples
- Ultrafast laser & Vibrational Optical Signal in Visible Region
=> Ultrafast Dynamics—IVR, Photobiological Systems



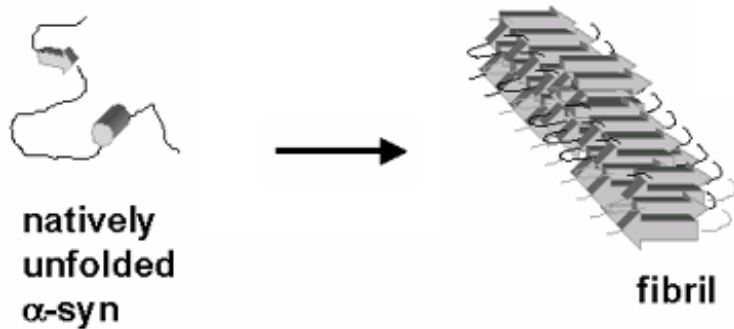
cSFG Vibrational Signatures for Secondary Structures



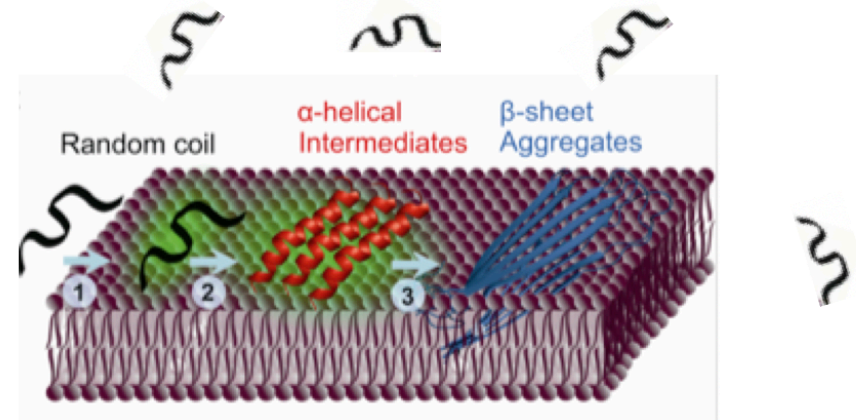
Chiral SFG Studies of Proteins at Interfaces

1. Amyloid Aggregation at Lipid/Water Interface (Li et al, *JACS*, 2010, 2011)
2. Proton Exchange of Proteins at Interfaces (Li et al, *JACS*, 2013)
3. Orientation of Proteins at Interfaces (Li et al, *JPCL*, 2015; Xiao et al *JMB*, 2012)
4. Functions of Biofilms Proteins (Wang et al. *Chem Comm*, 2016; Liu et al, *Langmuir* 2017)
5. Crowding Effect on Protein Folding at Interfaces (Liu et al. *PCCP*, 2018)

Aggregation of Amyloid Proteins



Alzheimer's Disease, Parkinson's Disease,
Huntington's Disease, Prion Disease, Type II Diabetes



1. Small oligomer interrupt cell membrane and cause cytotoxicity:
2. Aggregation catalyzed by interactions with lipid membrane

Challenge: Selectivity to

(1) Interfaces

(2) Secondary Structures

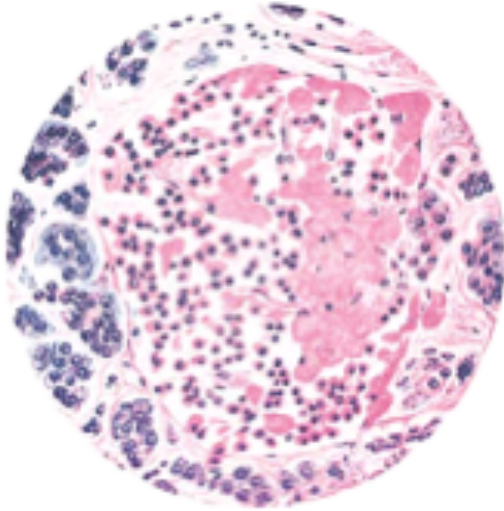
- NMR, EPR, CD—not surface-selective
- Reflective IR—spectral overlapping

Aggregation of Amyloid Proteins

Human Islet Amyloid Polypeptide (hIAPP)

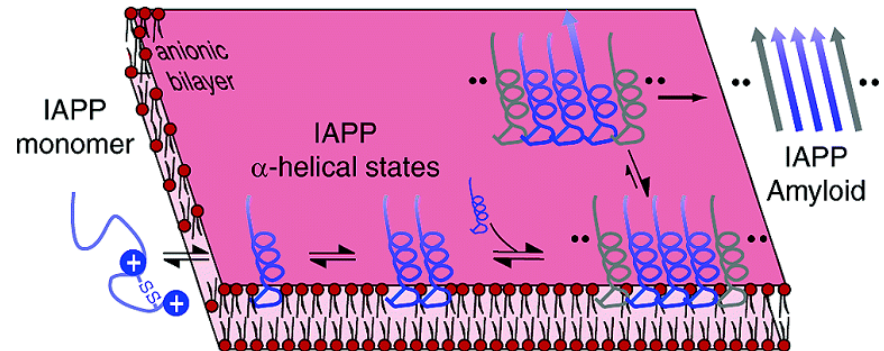


Pathology of type II diabetes



(Jo et al. *The New England Journal of Medicine*)

Model of IAPP aggregation



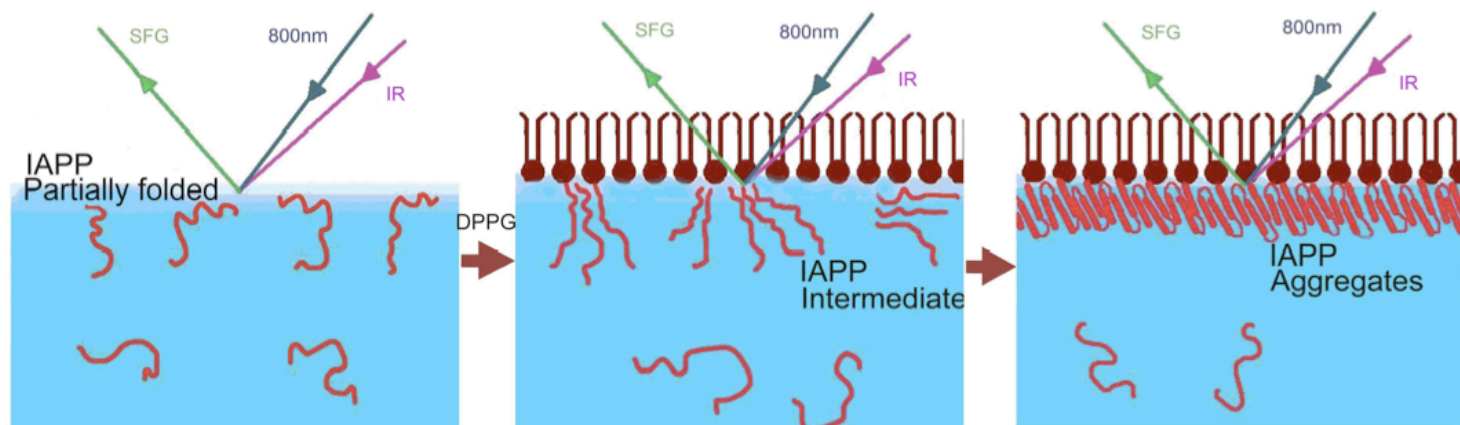
(Knight et al *Biochemistry* 2006)

- IAPP forms fibrils in pancreas
- IAPP aggregation
=> Death of β cells
=> Production of Insulin

- Adsorption as α -helices
- Alignment of α -helices
- Conversion to β -sheets

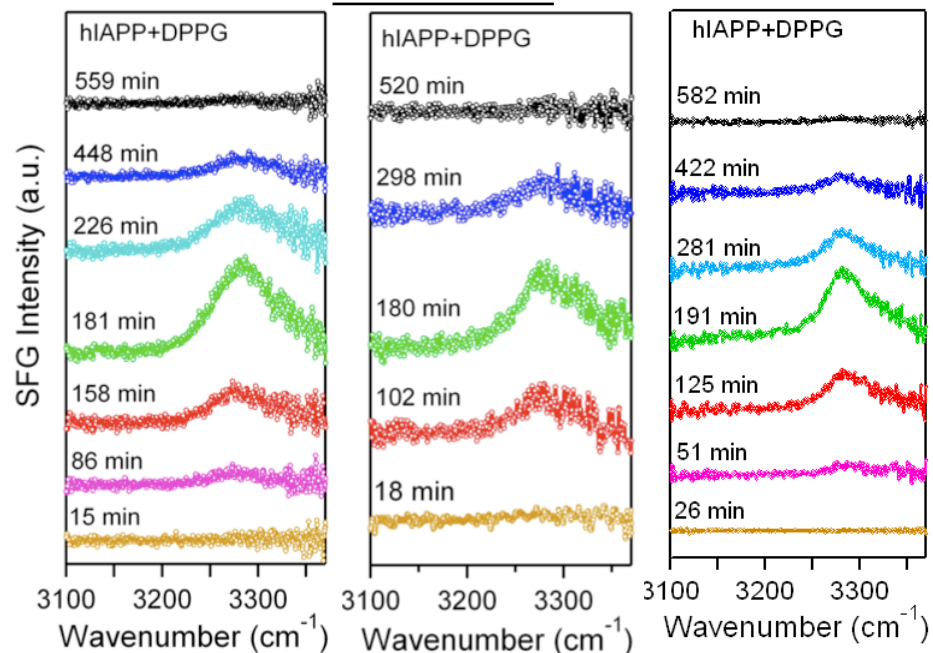
Aggregation of Amyloid Proteins

Kinetics: Chiral N-H vs. Chiral Amide I

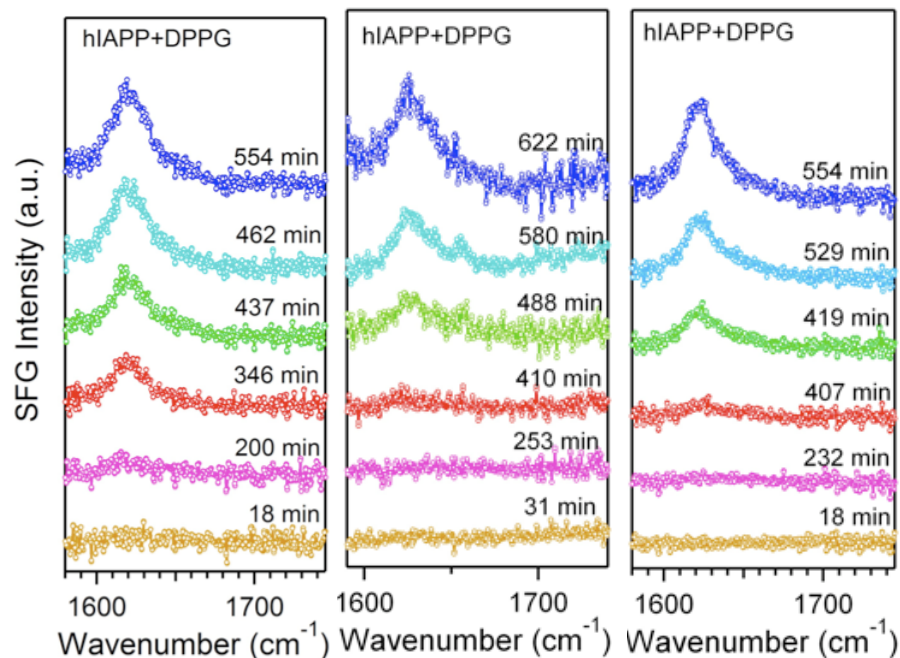


(Fu, L; Liu, J;
Yan, ECY
JACS., 2011)

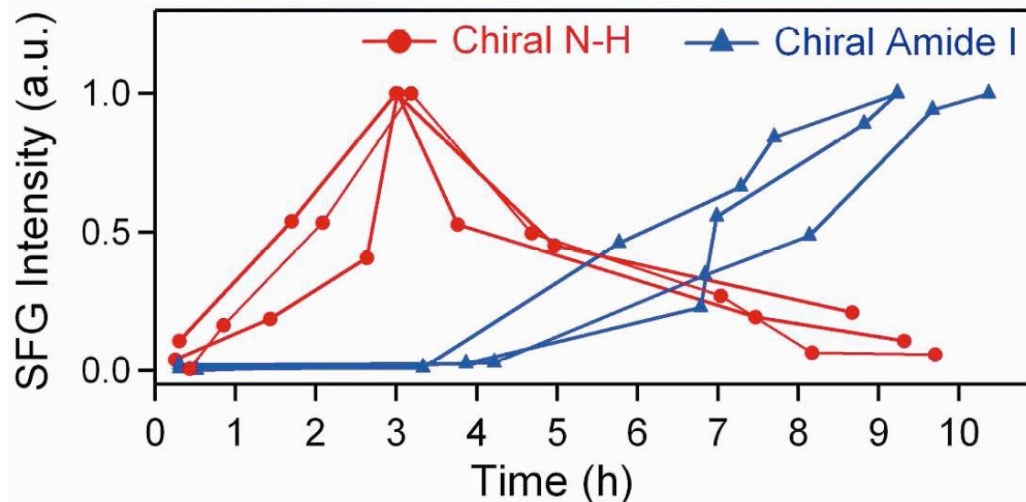
Chiral N-H



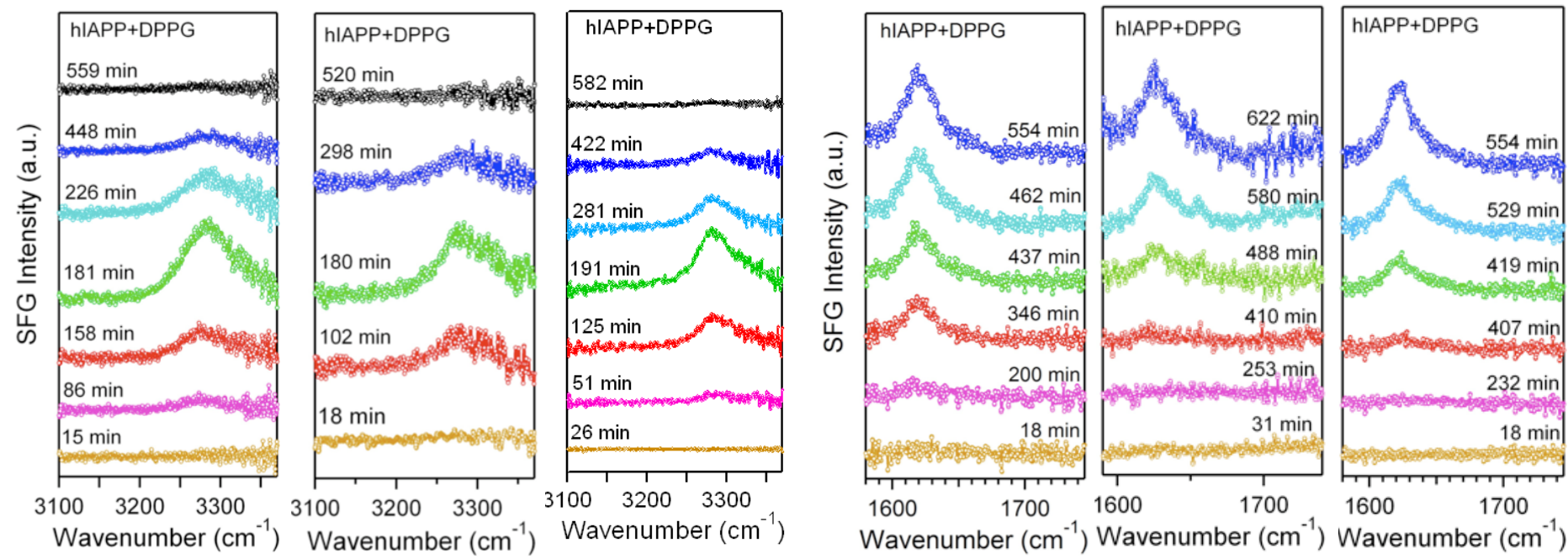
Chiral Amide I



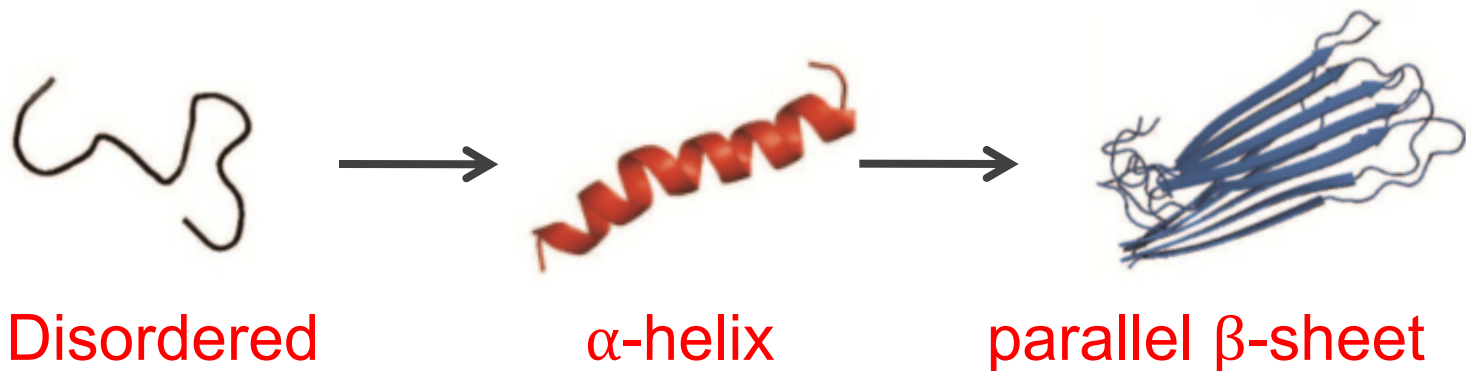
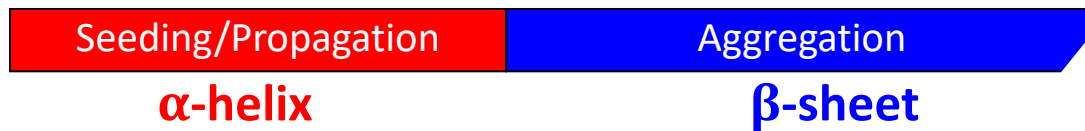
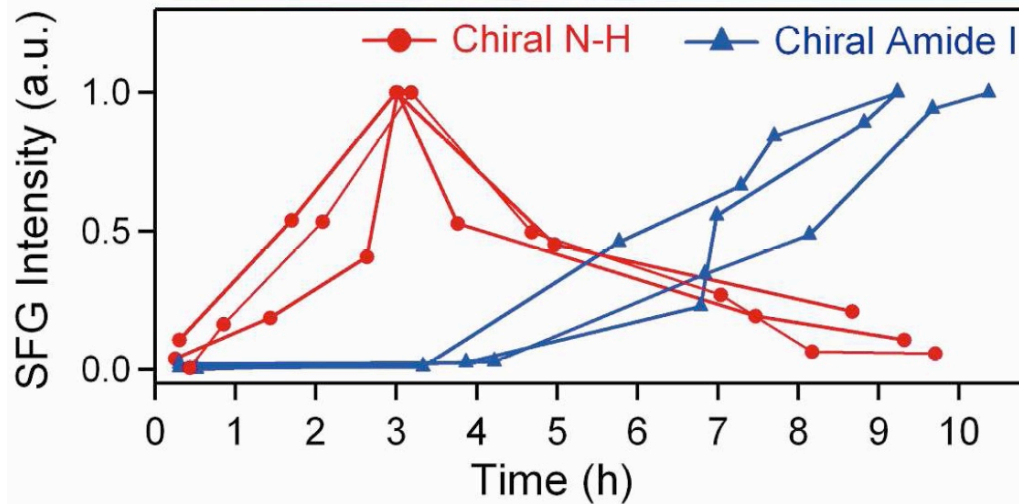
Aggregation of Amyloid Proteins



(Fu, L; Liu, J;
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JACS., 2011)



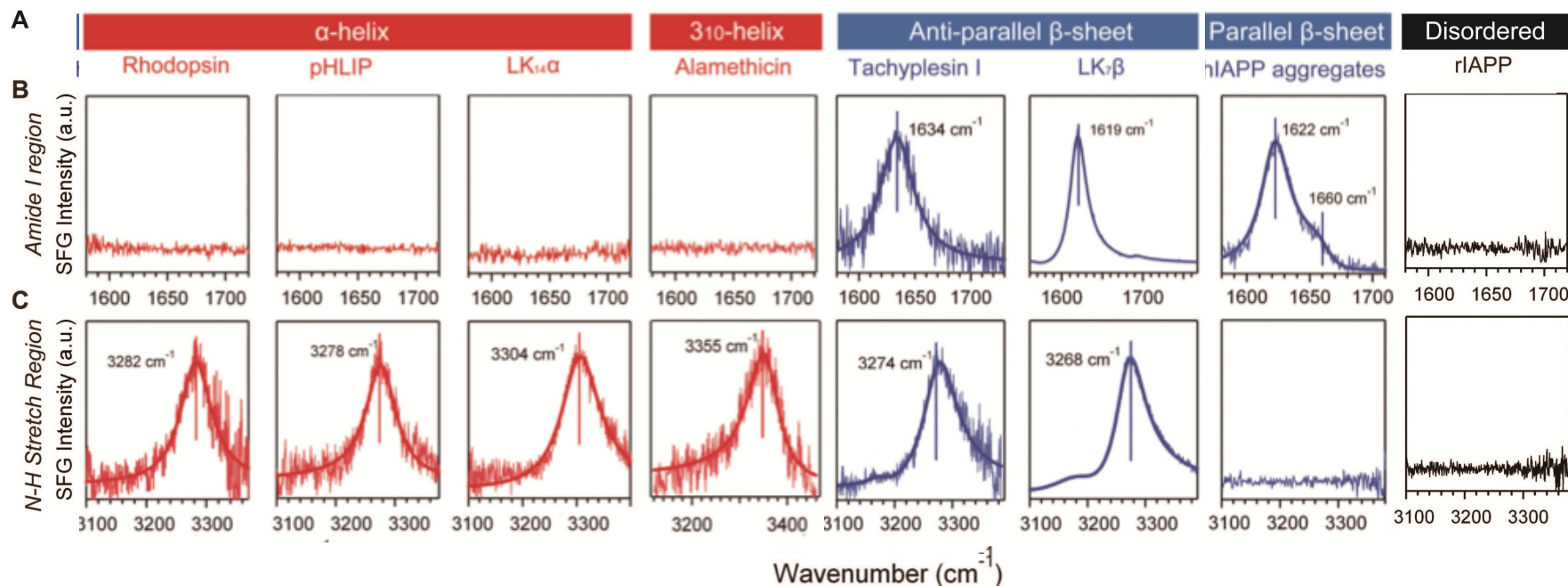
Aggregation of Amyloid Proteins



(Fu, L; Ma, G; Yan, ECY *J. Am. Chem. Soc.*, 132, 5405, 2010)

(Fu, L; Liu, J; Yan, ECY *J. Am. Chem. Soc.*, 133, 8094, 2011)

Summary: Characterization of Protein Secondary Structures at Interfaces

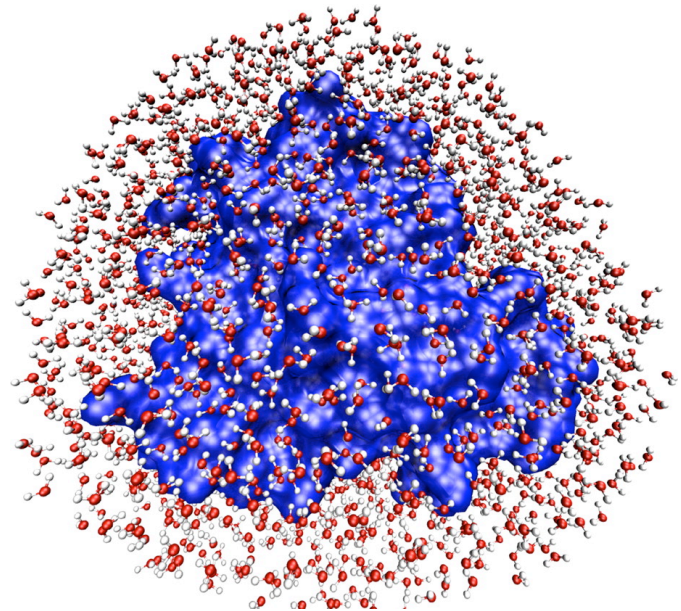
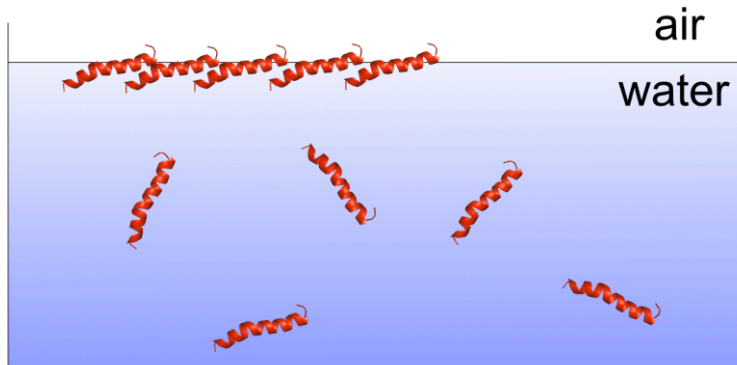


1. Amyloid Aggregation at Lipid/Water Interface (Li et al, *JACS*, 2010, *JACS* 2011)
2. Orientation of Proteins at Interfaces (Li et al, *JPCL*, 2015; Xiao et al *JMB*, 2012)
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5. Crowding Effect on Protein Folding at Interfaces (Liu et al. *PCCP*, 2018)

Protein Characterizations by Chiral Vibrational Sum Frequency Generation Spectroscopy

Outline

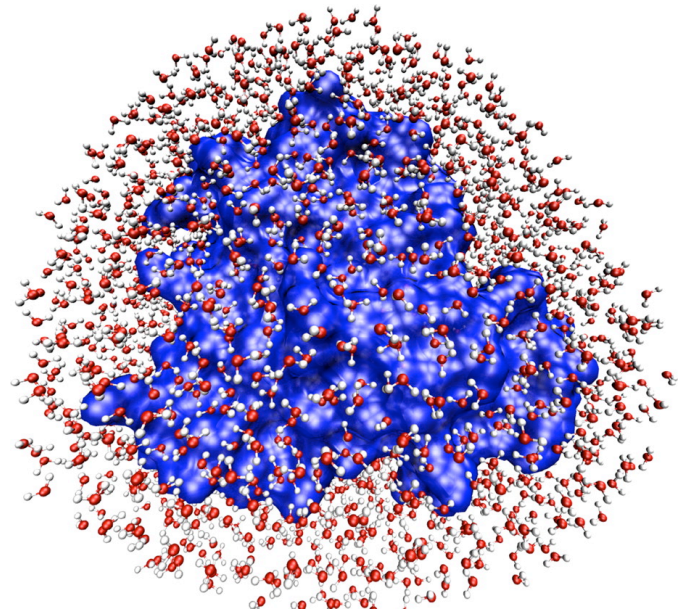
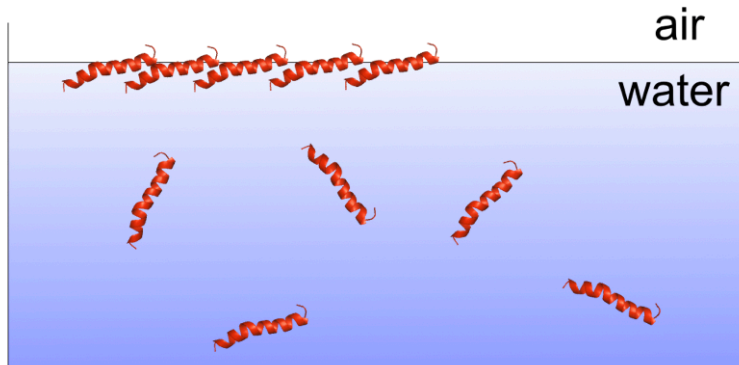
1. Introduction to Chiral SFG Spectroscopy
2. Charactering Protein Secondary Structures at Interfaces
3. Probing Water Structures in Hydration Shell of Proteins



Protein Characterizations by Chiral Vibrational Sum Frequency Generation Spectroscopy

Outline

1. Introduction to Chiral SFG Spectroscopy
2. Charactering Protein Secondary Structures at Interfaces
3. Probing Water Structures in Hydration Shell of Proteins



Chiral Water Superstructures Imprinted by DNA Detected by Chiral SFG

ACS
central
science

Research Article

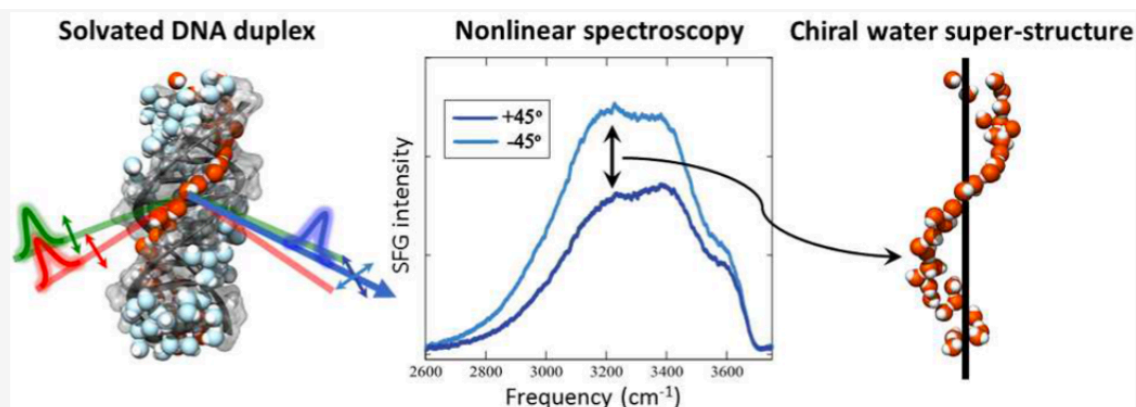
<http://pubs.acs.org/journal/acscii>

DNA's Chiral Spine of Hydration

M. Luke McDermott,[†] Heather Vanselow,[†] Steven A. Corcelli,[‡] and Poul B. Petersen^{*,†}

[†]Department of Chemistry and Chemical Biology, Cornell University, Ithaca, New York, United States

[‡]Department of Chemistry and Biochemistry, University of Notre Dame, Notre Dame, Indiana, United States



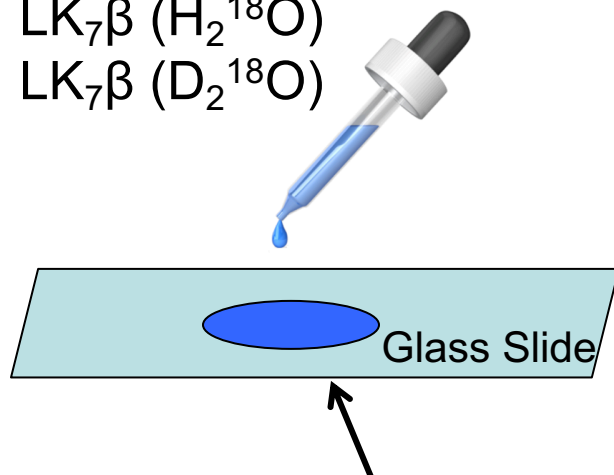
ACS Cent. Sci. 2017, 3, 708–714

**Can proteins orient waters into chiral superstructures?
Can these structures be observed by chiral SFG?**

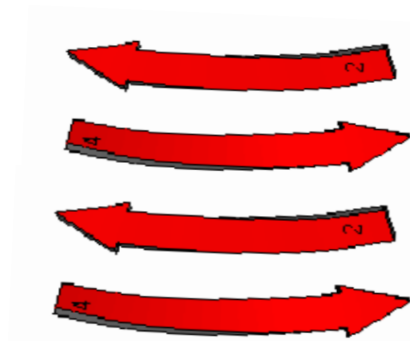
Chiral SFG Experiments: Chiral Water Superstructures Around Proteins

Model peptide, LK₇β (LK₇LKL): antiparallel β-sheet

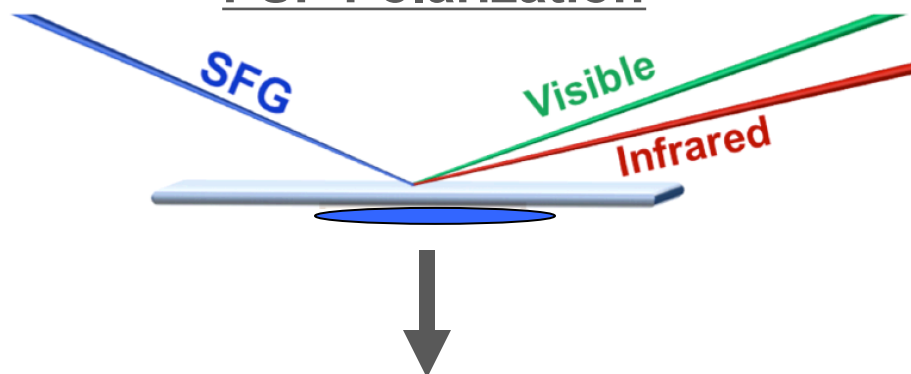
LK₇β (H₂O)
LK₇β (D₂O)
LK₇β (H₂¹⁸O)
LK₇β (D₂¹⁸O)



Dried overnight to obtain a thin film of LK₇β adsorbed on a glass slide



PSP Polarization



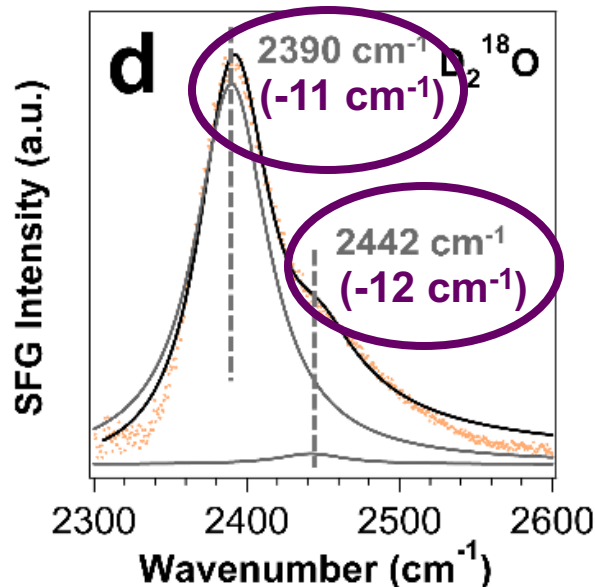
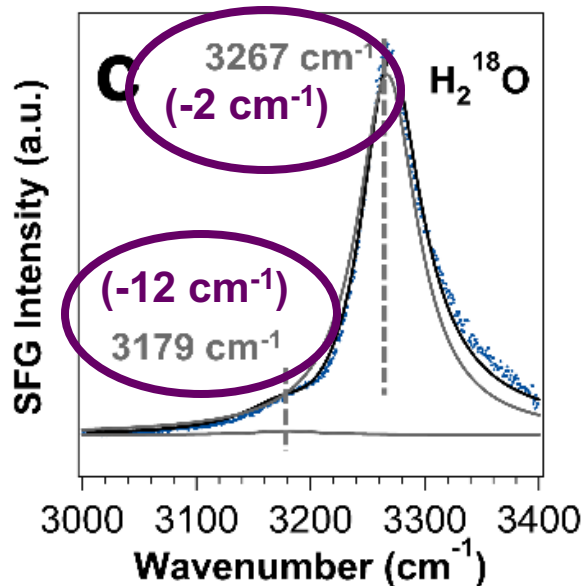
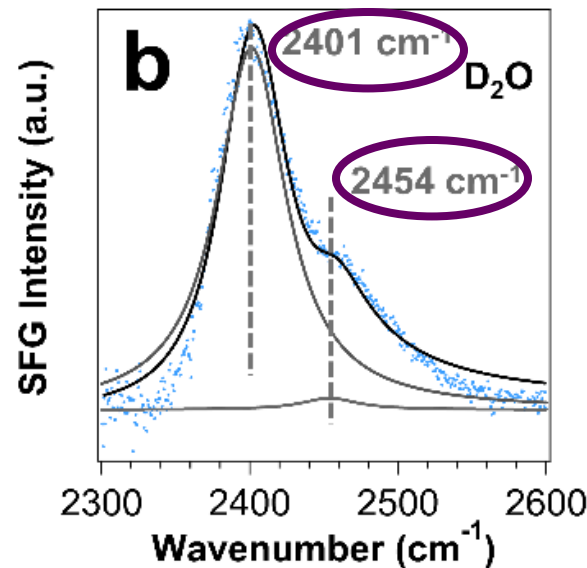
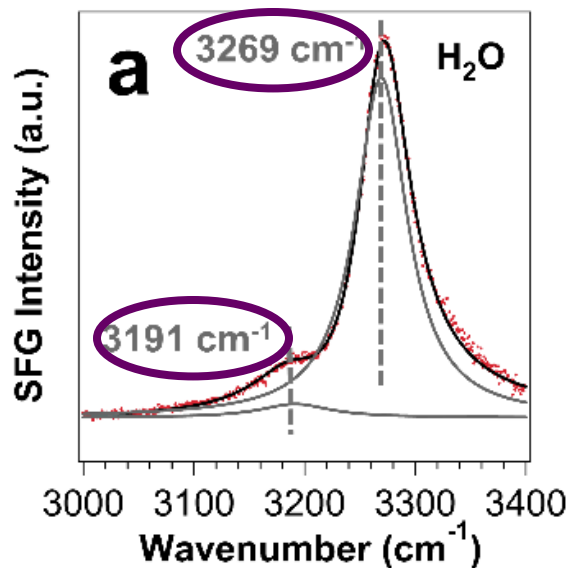
Chiral SFG spectra

1. ~3300 cm⁻¹ (O-H Stretch)
2. ~2400 cm⁻¹ (O-D stretch)

Isotopic Shifts of LK₇β Thin Film Spectra

One peak shifted by H₂¹⁸O

Both peaks shifted by D₂¹⁸O



Isotopic Shifts of LK₇β Thin Film Spectra

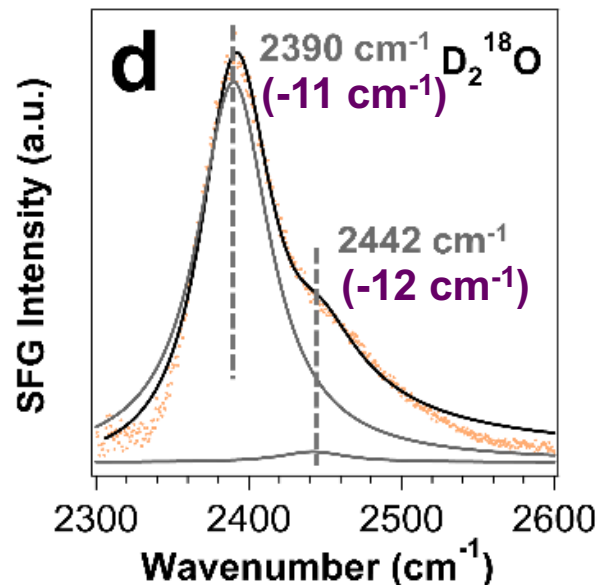
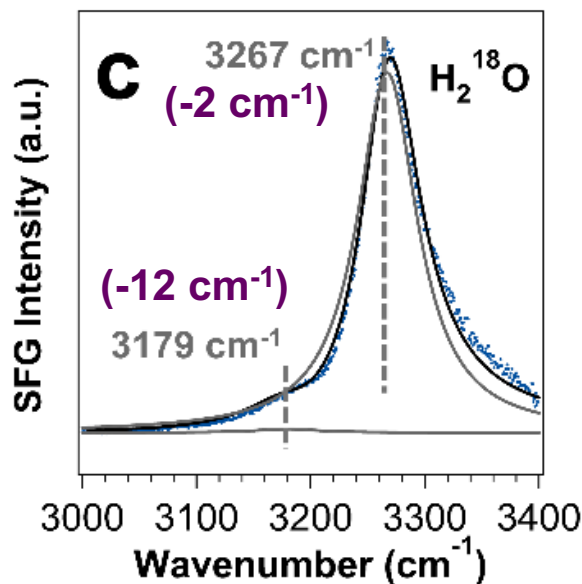
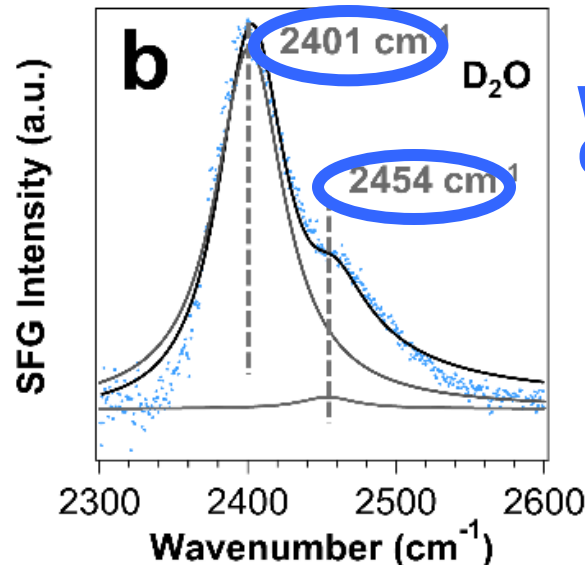
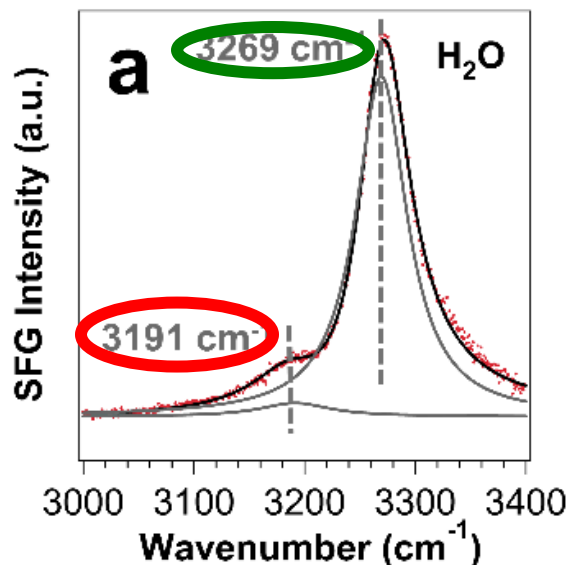
One peak shifted by H₂¹⁸O

Both peaks shifted by D₂¹⁸O

Peptide
N-H Stretch

Water
O-H Stretch

Water
O-D Stretch



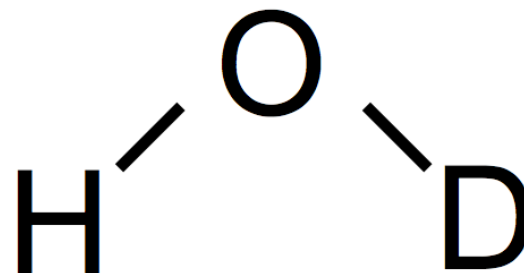
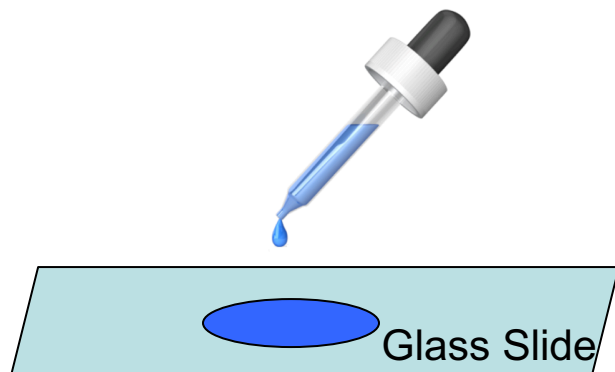
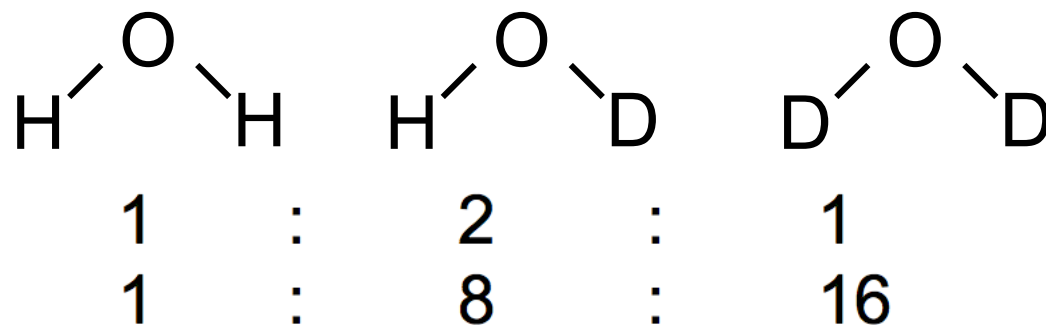
Isotopic Dilution

LK₇β (H₂O)

LK₇β (D₂O)

LK₇β (H₂O:D₂O 1:1)

LK₇β (H₂O:D₂O 1:4)



- Removing intramolecular coupling of two O-H stretches
- Changing the O-H stretch vibrational spectrum

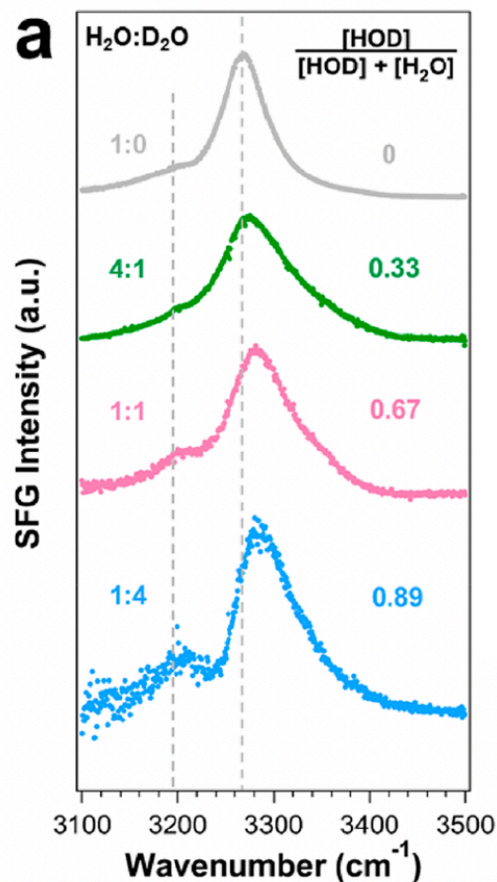
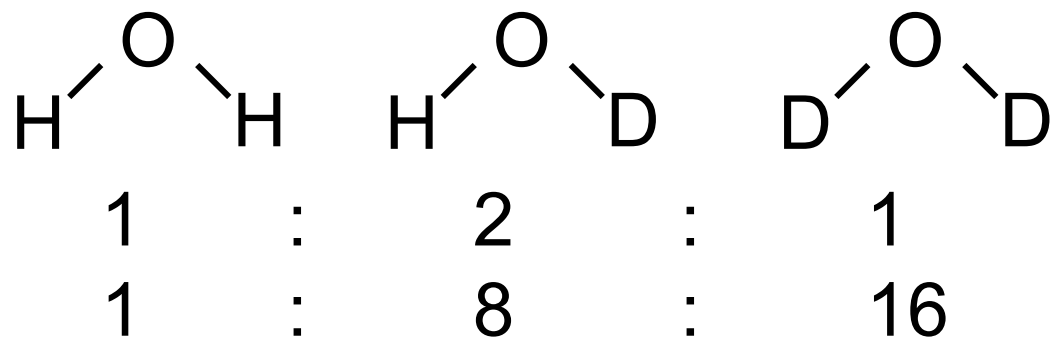
Isotopic Dilution

LK₇β (H₂O)

LK₇β (D₂O)

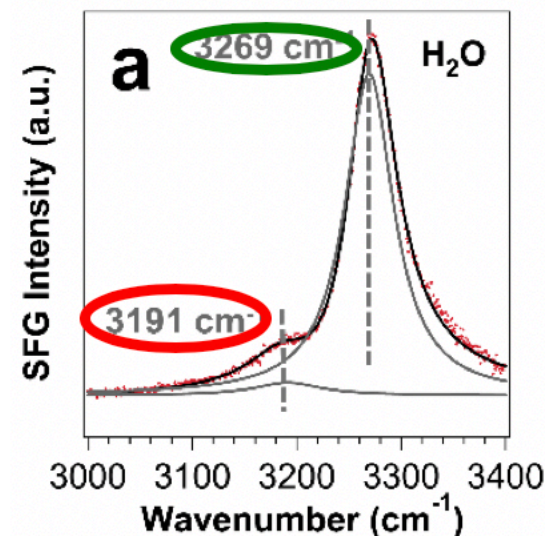
LK₇β (H₂O:D₂O 1:1)

LK₇β (H₂O:D₂O 1:4)



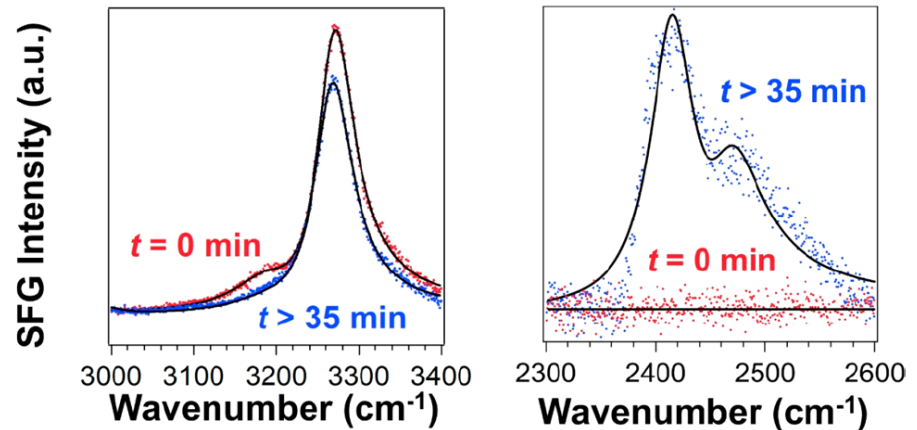
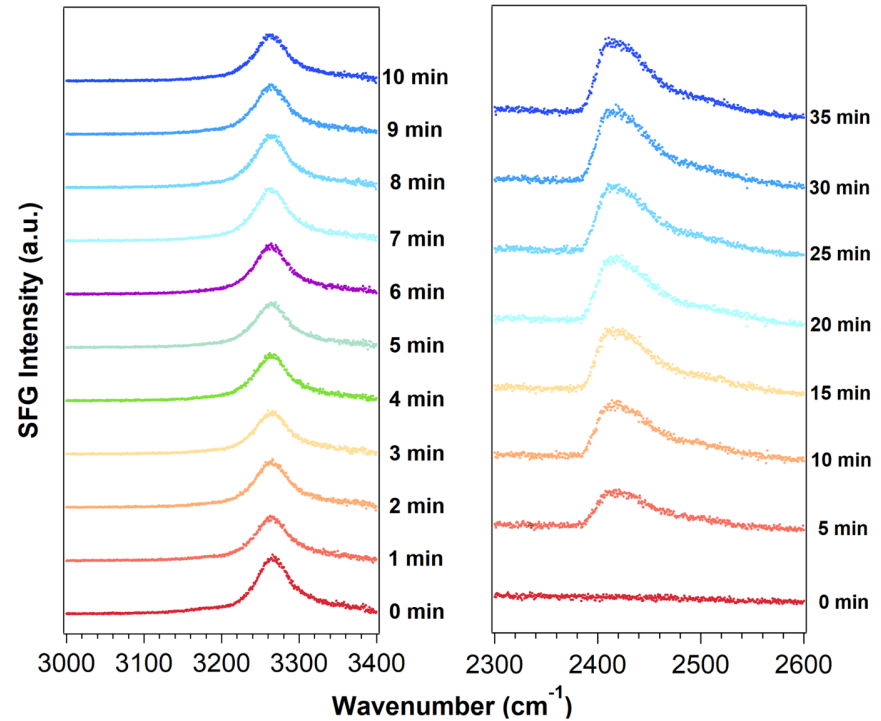
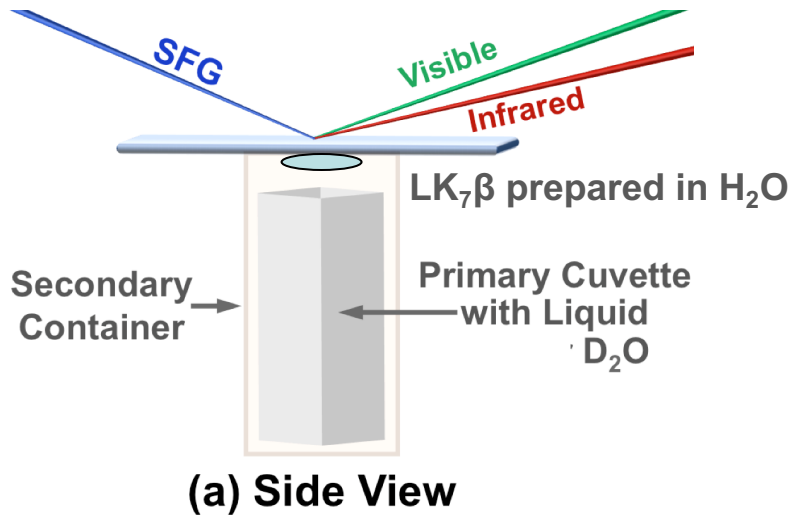
Peptide
N-H Stretch

Water
O-H Stretch



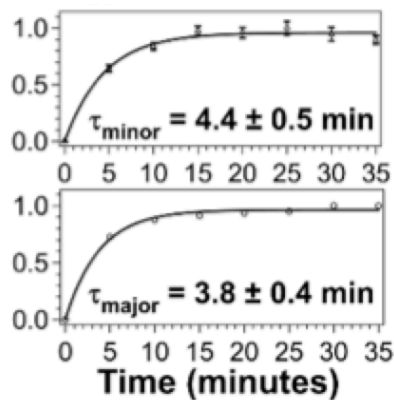
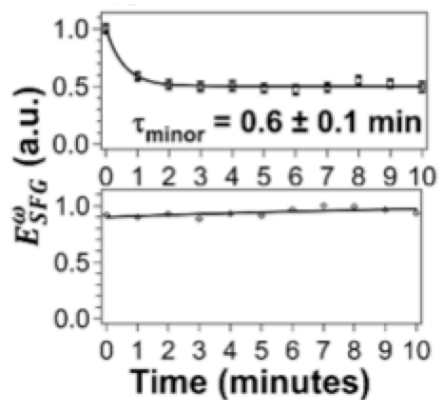
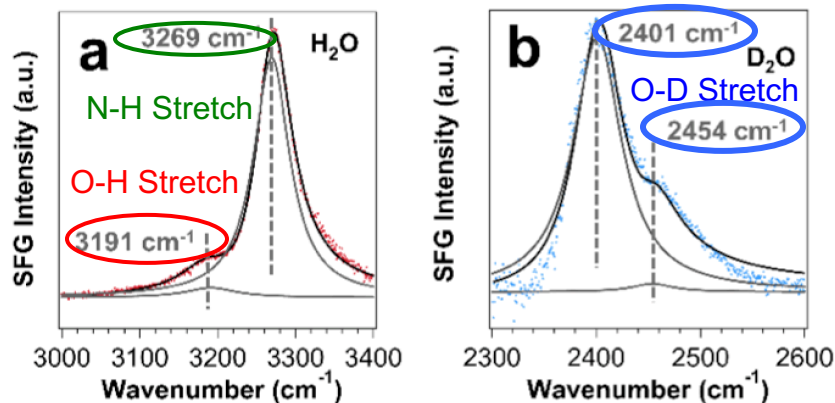
Kinetics of H₂O-D₂O Exchange

O-H Stretch Region O-D Stretch Region



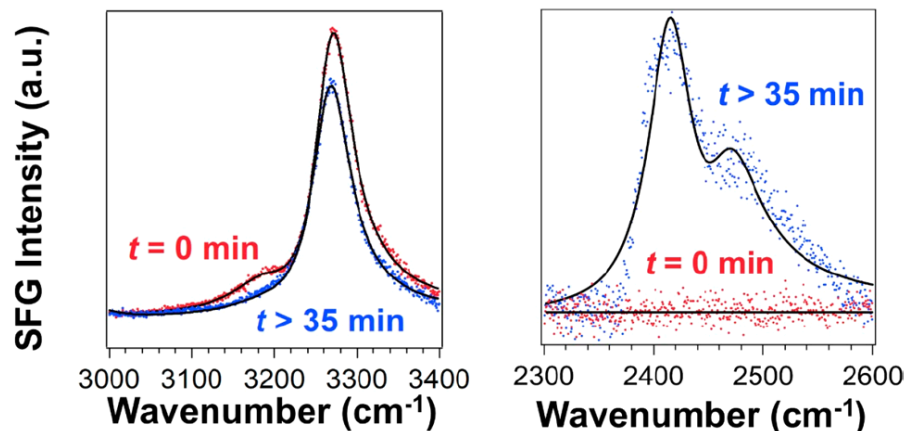
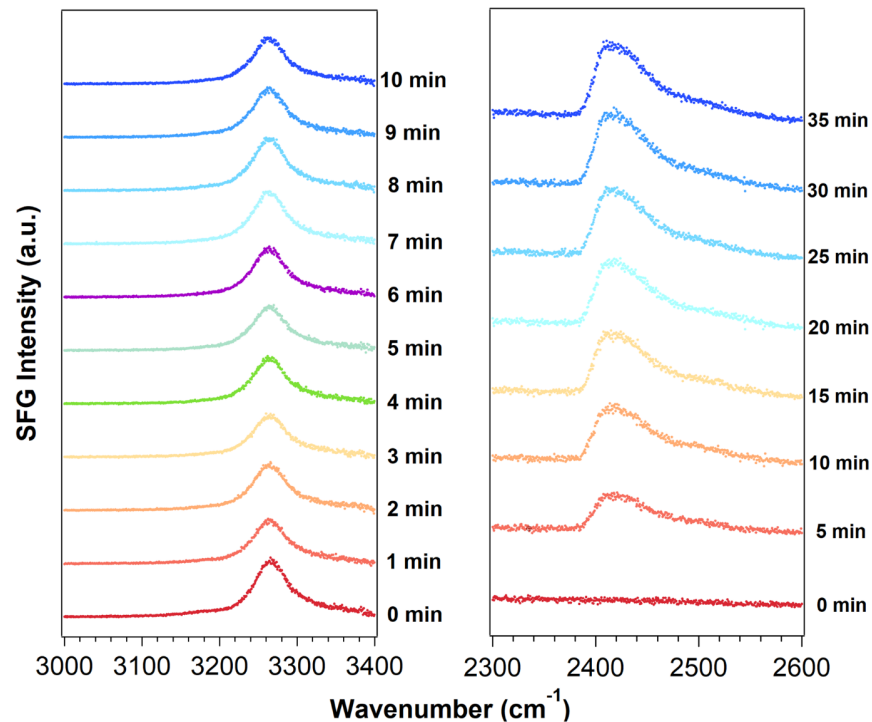
Kinetics of H₂O-D₂O Exchange

O-H Stretch Region O-D Stretch Region



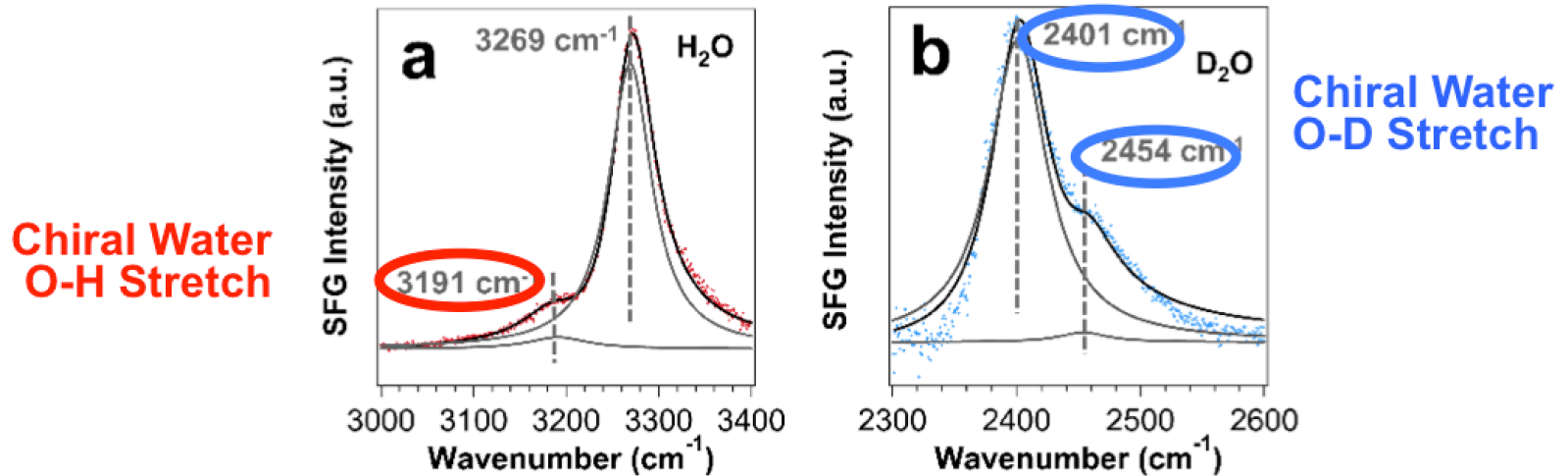
Two peaks from
two different
species

Possible that
both peaks
from a single
species



Chiral Water Superstructures observed by Chiral SFG

(1) Isotopic Shift, (2) Isotopic Dilution, (3) H₂O-D₂O Exchange

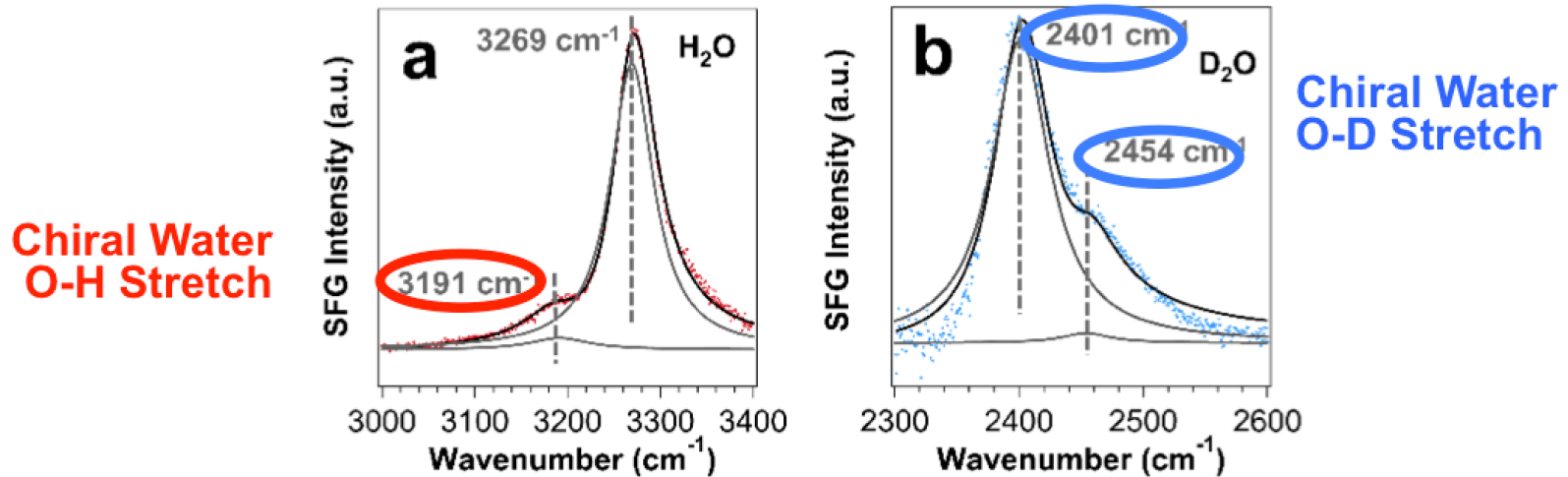


Implication:

1. Individual water molecules are achiral.
2. A protein secondary structure can imprint its chirality onto surrounding water
3. Water molecules form chiral supermolecular structures.

Chiral Water Superstructures observed by Chiral SFG

(1) Isotopic Shift, (2) Isotopic Dilution, (3) H₂O-D₂O Exchange



In progress:

1. Phase measurements using peptides made with L- and D-formed amino acids.
2. Protein denaturation mechanism.
3. MD simulations and computational studies:
 - to simulate the chiral SFG spectra
 - to explore the structures of water surrounding the peptide

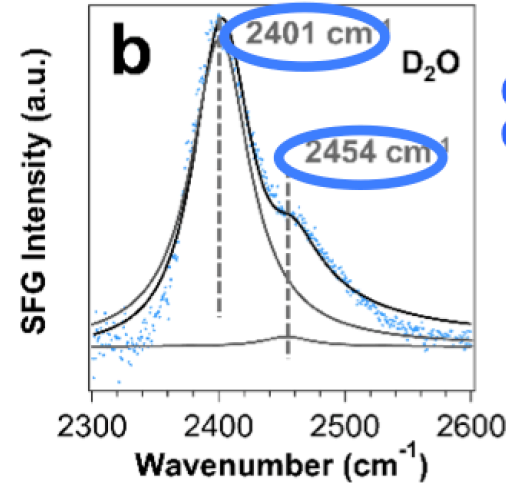
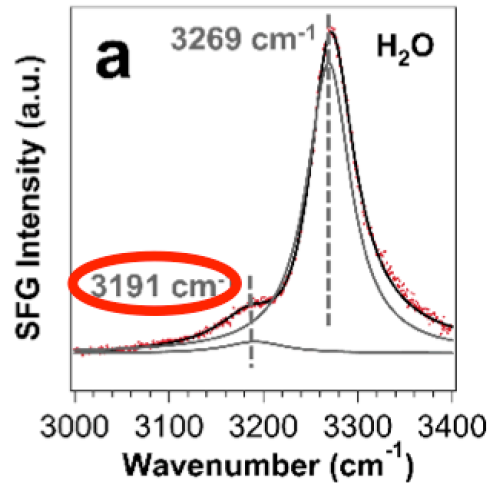


Prof. Sharon Dan Hammes-Schiffer
Konstantinovsky

Chiral Water Superstructures observed by Chiral SFG

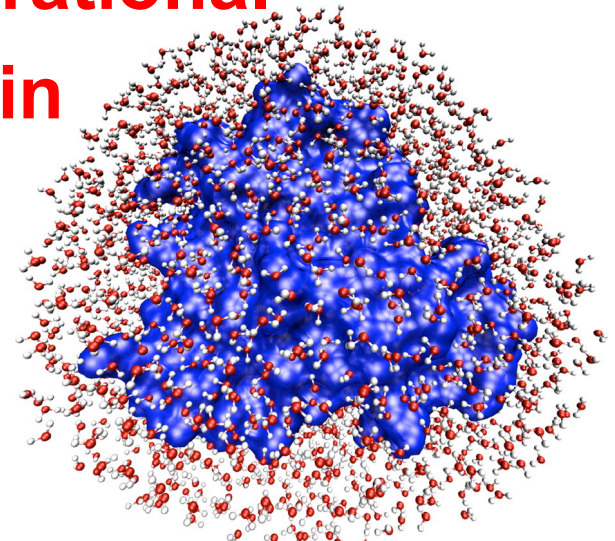
(1) Isotopic Shift, (2) Isotopic Dilution, (3) H₂O-D₂O Exchange

Chiral Water
O-H Stretch



Chiral Water
O-D Stretch

Chiral SFG for studying water vibrational
structures and dynamics in protein
hydration

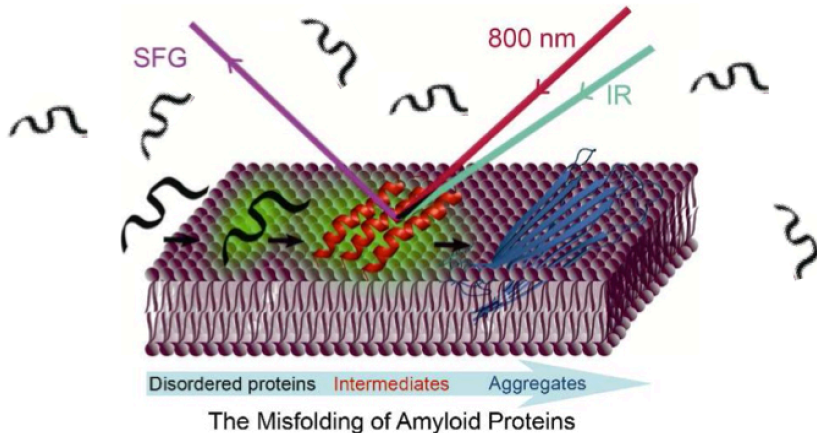


Summary

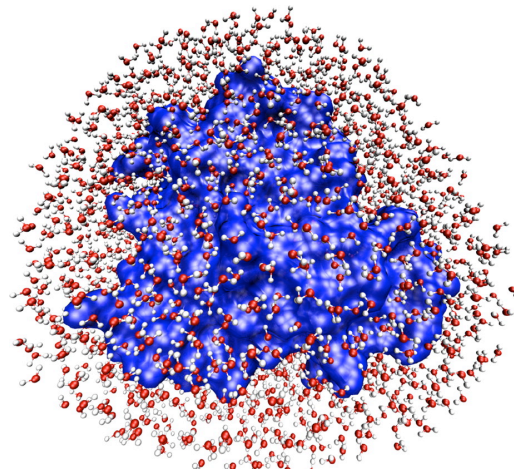
- I. Introduction to Chiral SFG Spectroscopy
- II. Protein Secondary Structures at Interfaces
- III. Water Structures in Protein Hydration

Conclusion

Protein Secondary Structures at Interfaces



Water Structures in Protein Hydration



Chiral SFG can introduce some new opportunities to probe protein structures and dynamics

Acknowledgements



Yan Lab Member

- Ya-Na (Anna) Chen
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- **Jaintao Chen**
- **Zachary Taitz**
- **Dan Konstantinovsky**

Yan Lab Alumni

- **Zhuguang Wang**
- **Li Fu**
- Ying Guo
- **Jian Liu**
- **Gang Ma**
- Victoria Mooney
- Yuting Liu
- **Wei Liu**
- Yingying Cai
- **Dr. Zahra Sophrabpour**
- Kelly Culhane

Collaborators

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- Prof. John Tully
- Prof. Sharon Hammes-Schiffer
- Prof. Roger Leblanc (U. Miami)

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